

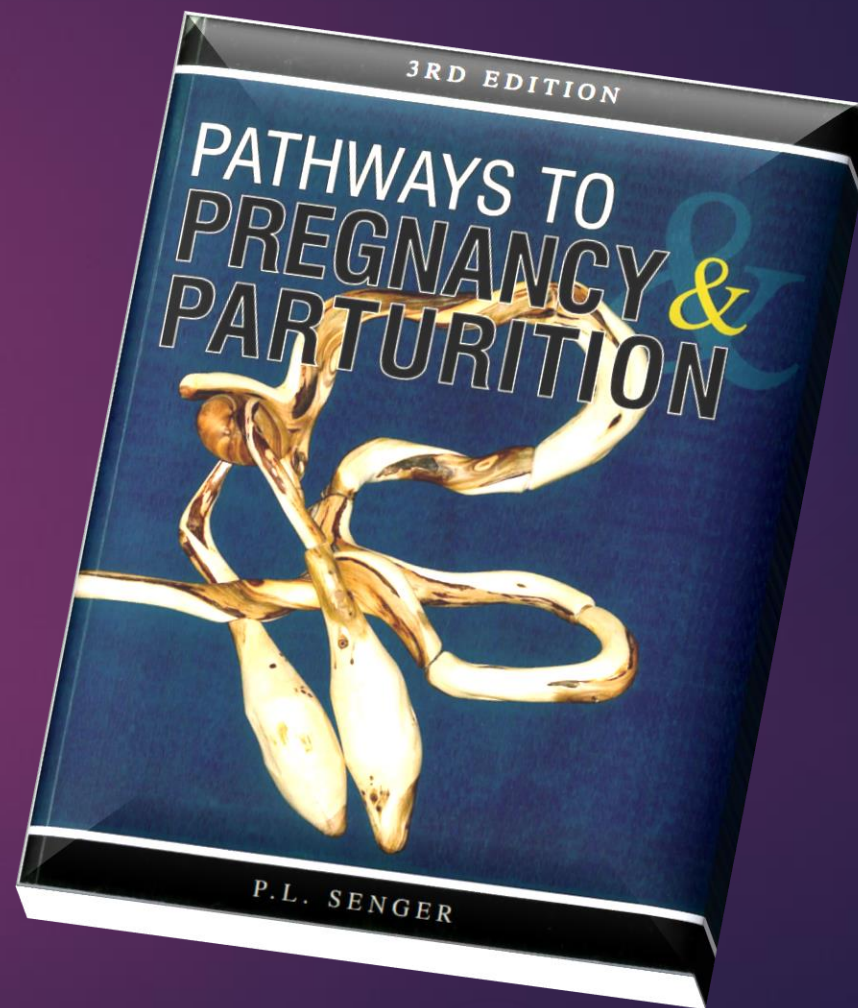
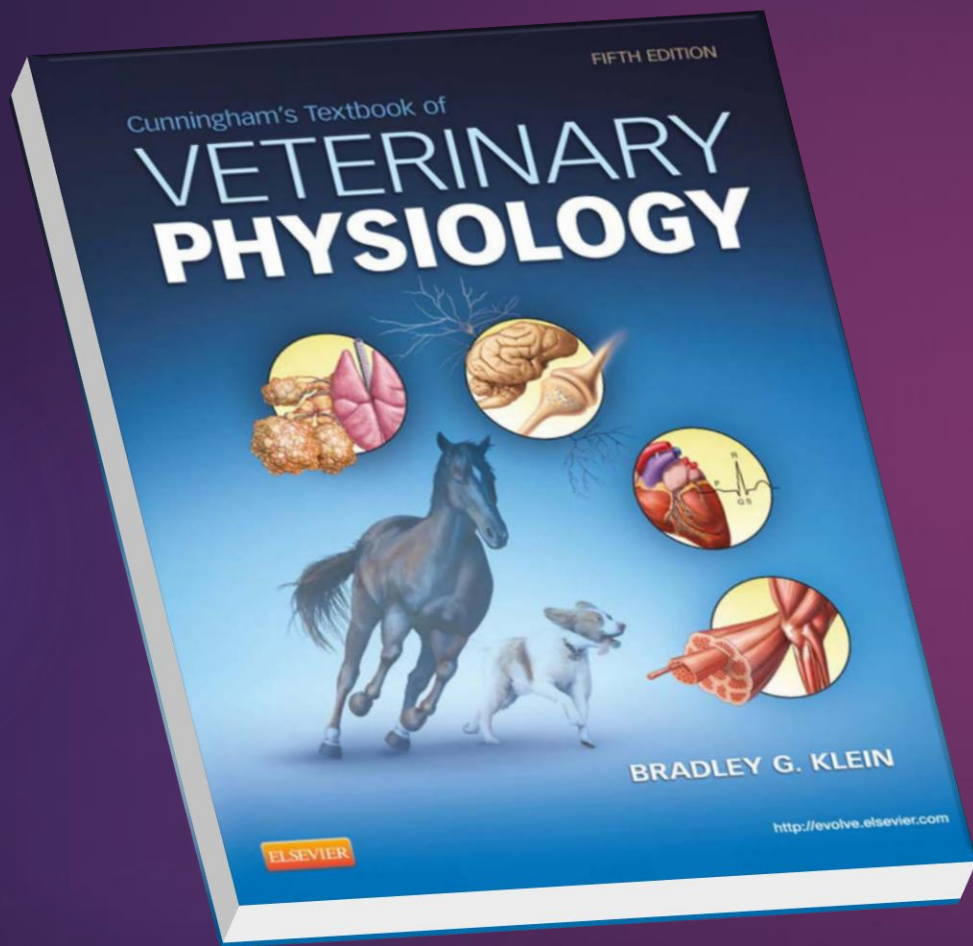


# *Reproductive Physiology in Domestic Animals*

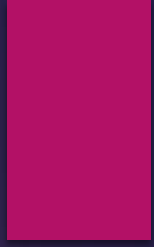
*Part one*

*A. K.Goudarzi, D.V.M. Ph.D.  
Department of Basic Sciences  
I.A. University*

# References



For PDF file, visit: [www.kajalpetclinic.com](http://www.kajalpetclinic.com)

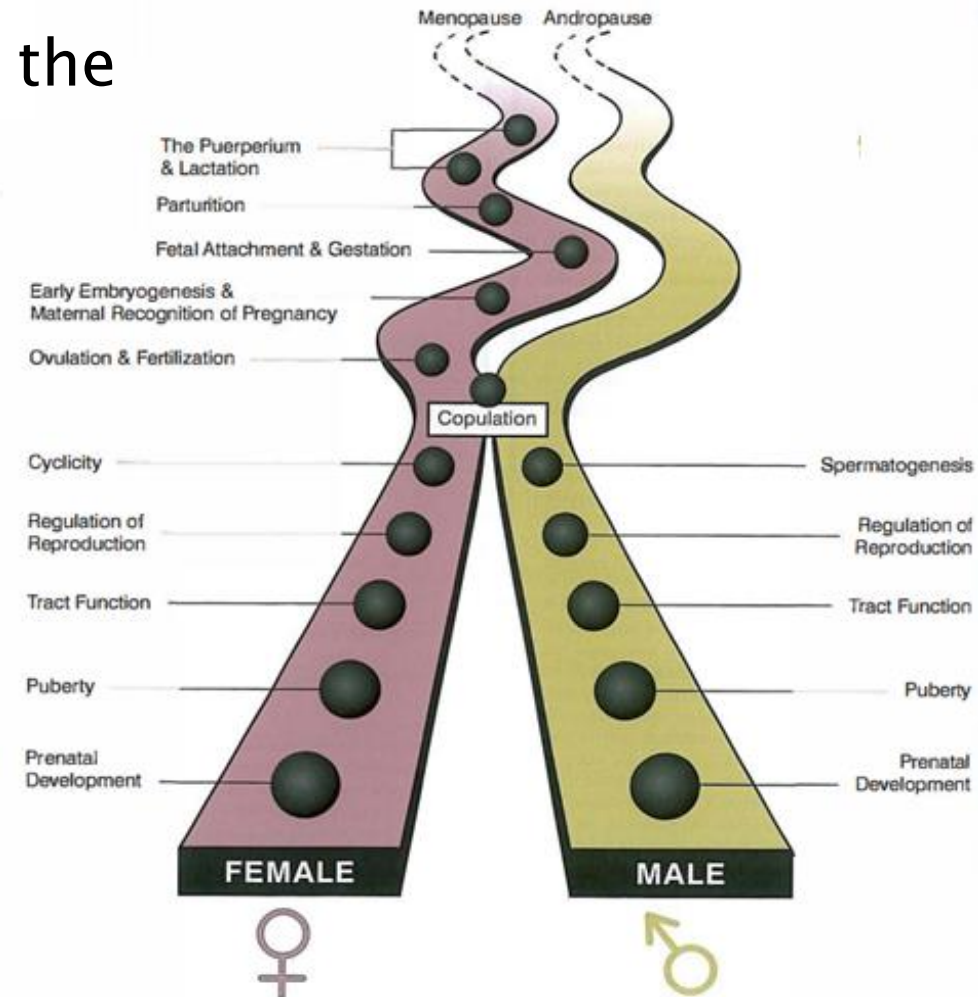


# *Introduction*

# Introduction

**Reproduction** is a sequence of events as follows:

- **Development** of the reproductive system in the embryo
- **Puberty**: ability to produce fertile gametes.
- **Cyclicity**
- Reproductive **behavior** and **copulation**.
- **Fertilization**
- **Placenta** formation.
- **Parturition**.
- **Lactation**
- Reestablishing **cyclicity**



# *Introduction*

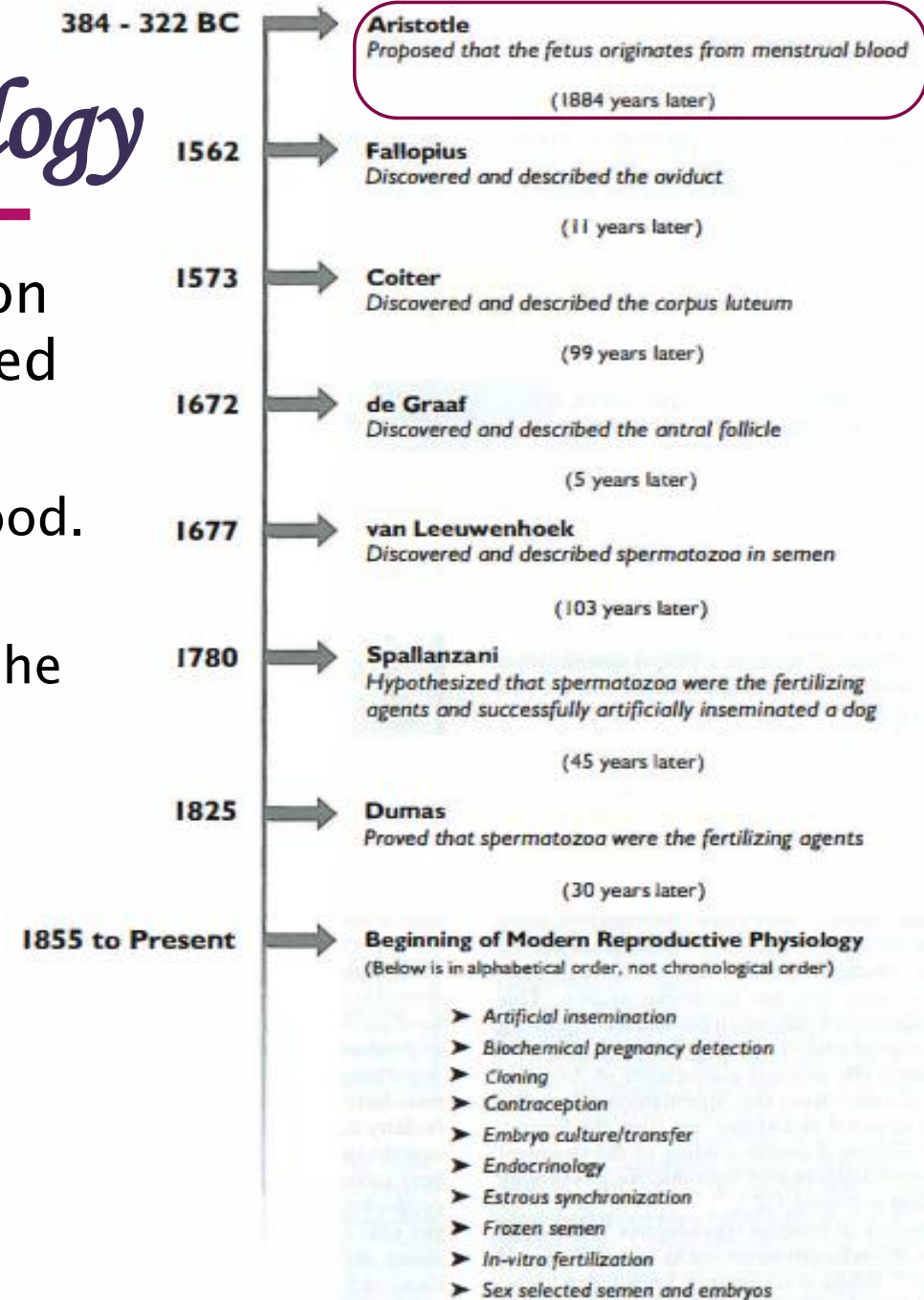
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**Reproductive Science** Consists of Several Subspecialties:

- **Andrology**
  - deals specifically with the study and treatment of male animals including humans.
- **Gynecology**
  - deals specifically with reproductive issues in women.
- **Theriogenology**
  - focuses on the reproductive system in animals.
- **Obstetrics**
  - a branch of reproductive physiology, veterinary medicine and/ or human medicine that specializes in the female before, during and after parturition.

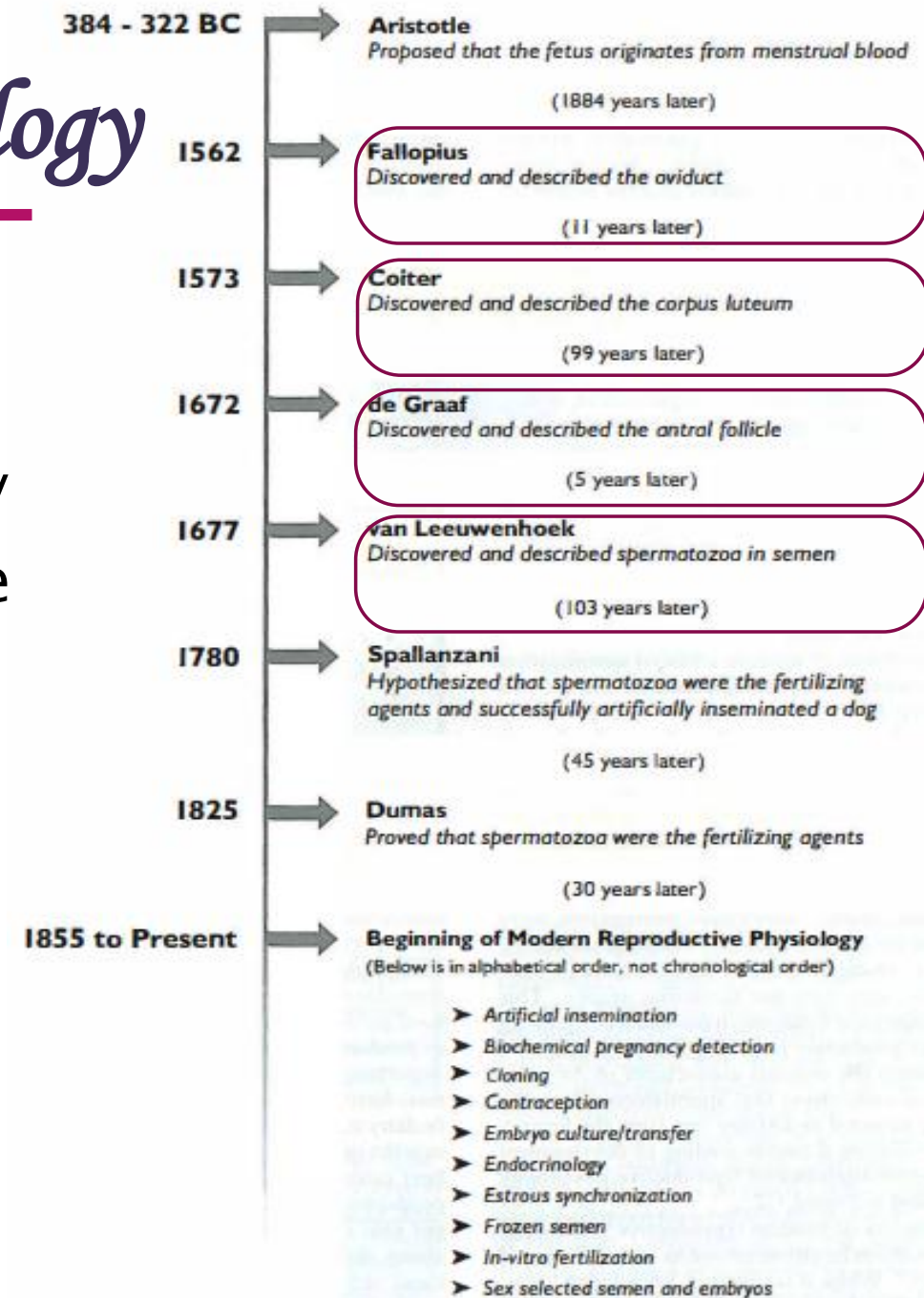
# The History of Reproductive Physiology

- **Aristotle** provided the first recorded information on how he thought the reproductive system functioned in his book entitled “**Generation of Animals**”.
  - He believed that the fetus arose from menstrual blood.
  - he concluded, based on the observation that menstruation did not occur during pregnancy that the **fetus was derived from menstrual blood**.
  - He also proposed that the conversion of menstrual blood to a fetus was initiated by **seminal fluid** deposited in the female during copulation.
  - Aristotle thought that **semen was derived from all parts of the body** and that the testes were simply pendular weights that kept the transport ducts (the ductus deferens) from becoming kinked or plugged with seminal fluid.



# The History of Reproductive Physiology

- About 2,000 years later **Fallopian** described the **oviducts**.
  - The name Fallopian tube reflects his discovery
- A student of Fallopian, **Coiter**, discovered the **corpus luteum** in 1573
- Almost 100 years later a scientist named **Regnier de Graaf** described the **antral follicle** that has been named the Graafian follicle in honor of his discovery.
- **van Leeuwenhoek** developed a simple **microscope** in 1677 which was a major technological breakthrough.



# The History of Reproductive Physiology

- A medical student suggested to van Leeuwenhoek that semen might contain living cells. Using his microscope, van Leeuwenhoek observed semen and discovered that it contained small particles that moved about. He referred to these particles as "animalcules".
  - van Leeuwenhoek found that similar "animalcules" were present in semen from males of many species and published a paper on his observations in 1677.
- The most widely accepted speculation of the day was that the "animalcules" (spermatozoa) contained fully formed individuals within their cellular confines. In other words, the sperm head was thought to contain a microscopic, yet fully formed individual

384 - 322 BC

**Aristotle**  
Proposed that the fetus originates from menstrual blood

(1884 years later)

1562

**Fallopian**  
Discovered and described the oviduct

(11 years later)

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**Coiter**  
Discovered and described the corpus luteum

(99 years later)

1672

**de Graaf**  
Discovered and described the antral follicle

(5 years later)

1677

**van Leeuwenhoek**  
Discovered and described spermatozoa in semen

(103 years later)

1780

**Spallanzani**  
Hypothesized that spermatozoa were the fertilizing agents and successfully artificially inseminated a dog

(45 years later)

1825

**Dumas**  
Proved that spermatozoa were the fertilizing agents

(30 years later)

1855 to Present

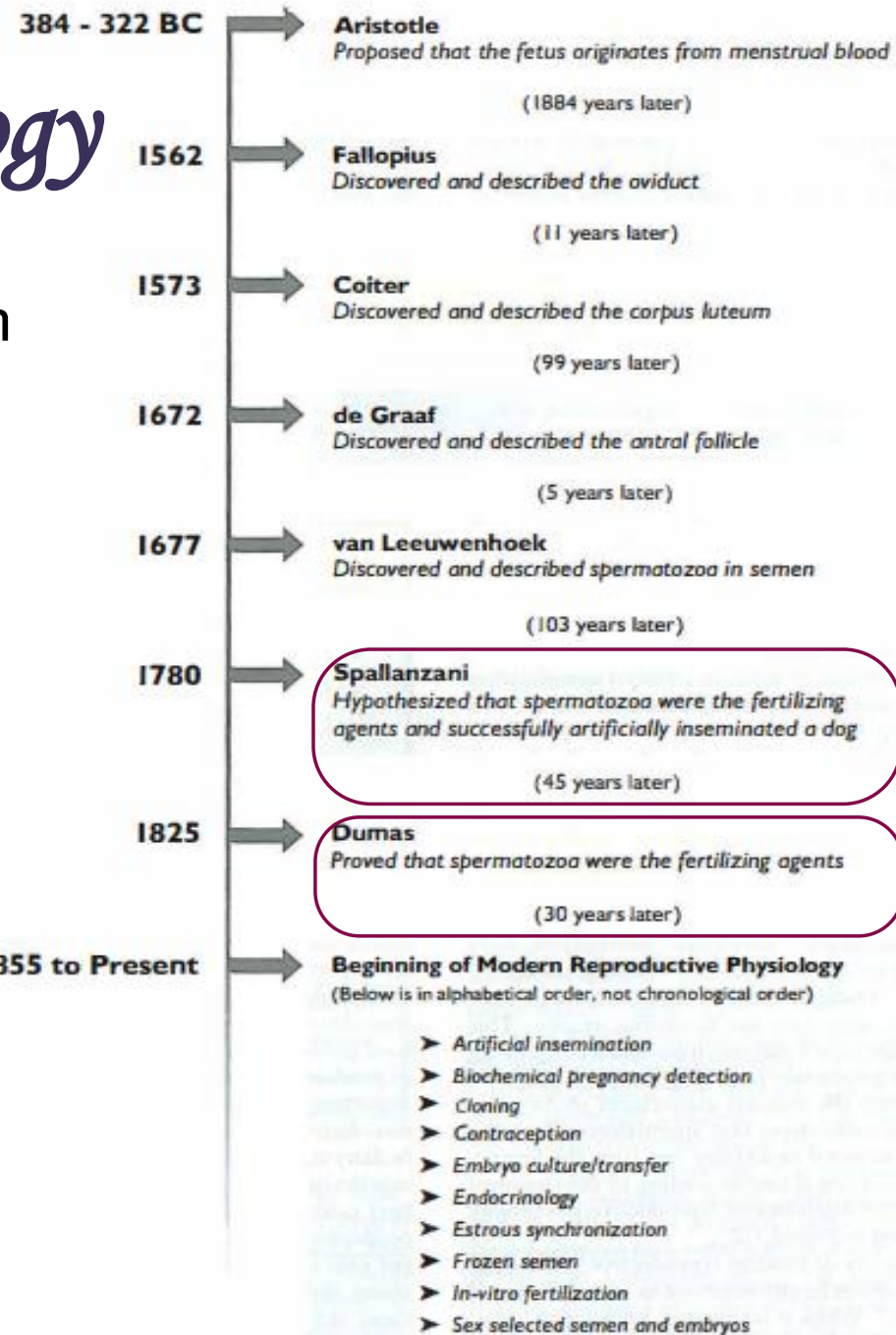
**Beginning of Modern Reproductive Physiology**  
(Below is in alphabetical order, not chronological order)

- Artificial insemination
- Biochemical pregnancy detection
- Cloning
- Contraception
- Embryo culture/transfer
- Endocrinology
- Estrous synchronization
- Frozen semen
- In-vitro fertilization
- Sex selected semen and embryos



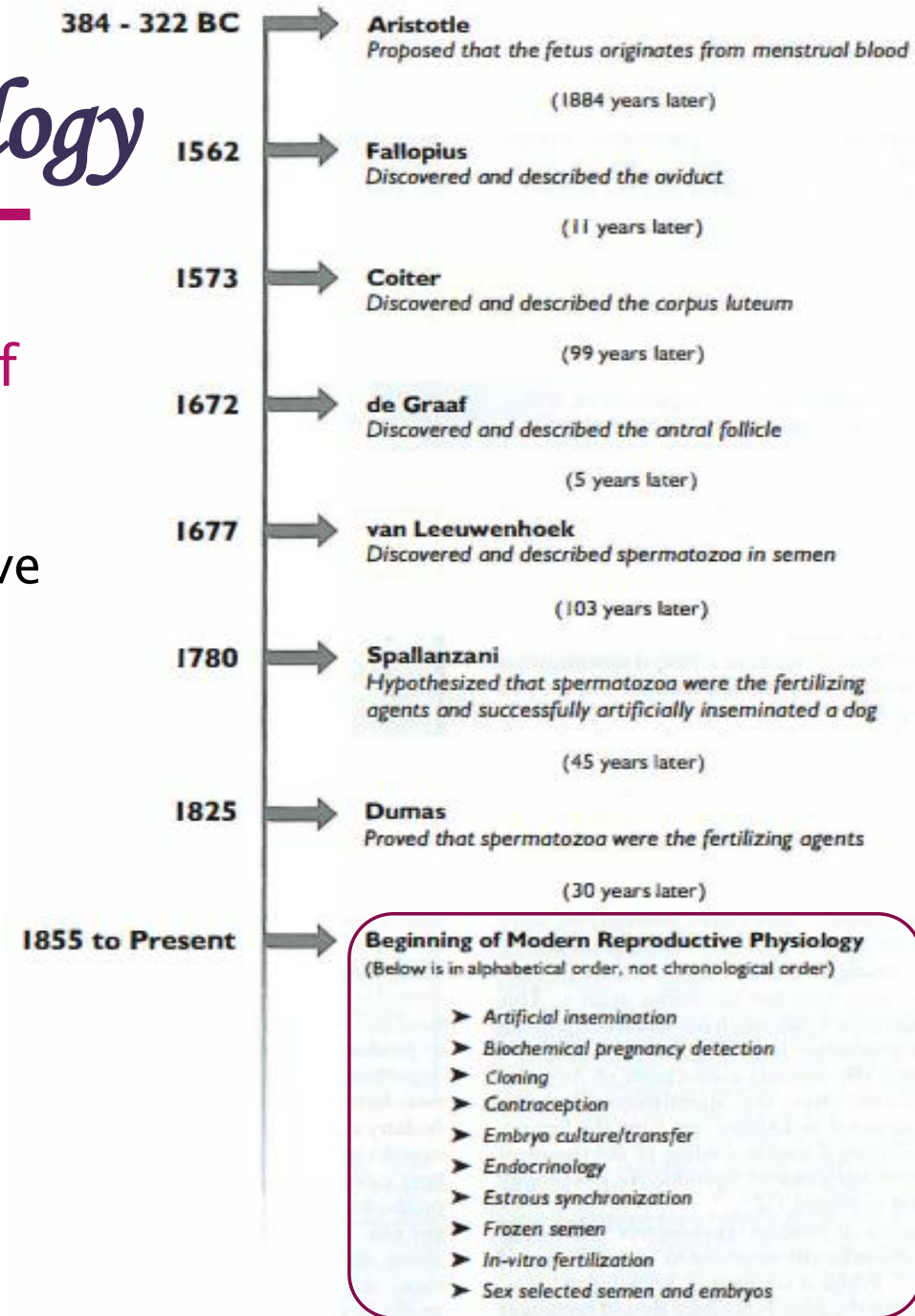
# The History of Reproductive Physiology

- The father of modern artificial insemination was an Italian priest named **Spallanzani**.
  - He showed that one drop of dog semen diluted with 25 pounds of fluid retained its ability to fertilize. Using the dog, he performed the first **artificial insemination**.
- A scientist named **Dumas** collected bodies about 1 mm in diameter from rabbit follicles. He discovered that follicles contained ova and were precursors to the early embryo. This discovery led Dumas to conclude that the "animalcules," now called spermatozoa, were responsible for uniting with the ovum and producing an embryo.
  - This early description of **fertilization** marked the beginning of **modern reproductive physiology**



# The History of Reproductive Physiology

- The era of modern reproductive physiology that followed can be characterized as an "explosion of knowledge."
  - recognition that the gonads produce steroid hormones that alter the function of the reproductive tissues and that the anterior pituitary controls the function of the gonads were major milestones of discovery.
  - The understanding that females experience reproductive cyclicity and that they ovulate with predictable frequency continued the explosion of knowledge.
  - Development of the radioimmunoassay for the measurement of hormones enabled the precise description of hormonal profiles in both the male and female



# The History of Reproductive Physiology

- These discoveries opened the door for the development of methods for **artificial manipulation of reproductive processes**.
  - In the 1940's and 1950's, understanding spermatozoal physiology and how these cells function in test-tube environments led to successful **artificial insemination** in several species.
  - It wasn't until the 1960's that it was understood that **prostaglandin  $F_{2\alpha}$**  regulated the length of the estrous cycle in most mammalian females.
  - The discovery that natural prostaglandin  $F_{2\alpha}$  caused **destruction of the corpus luteum** made it possible to manipulate and alter estrous cycles and to **control the time of ovulation**.
- Such application is now commonplace in dairy and beef enterprises throughout the world.

384 - 322 BC

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Proposed that the fetus originates from menstrual blood

(1884 years later)

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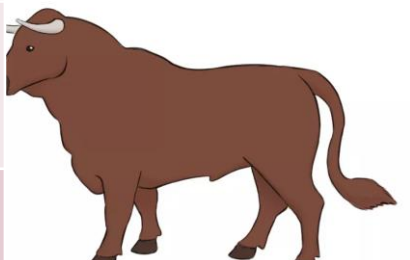
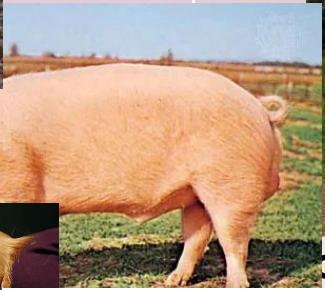
1855 to Present

**Beginning of Modern Reproductive Physiology**  
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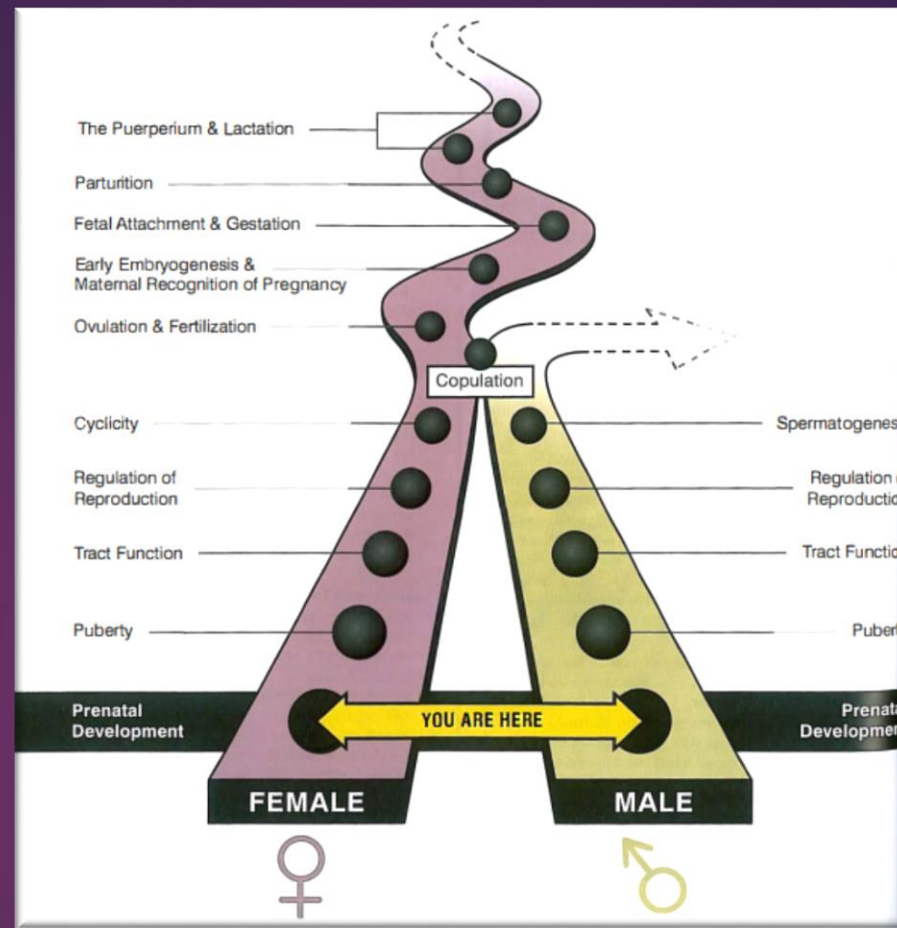
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- Biochemical pregnancy detection
- Cloning
- Contraception
- Embryo culture/transfer
- Endocrinology
- Estrous synchronization
- Frozen semen
- In-vitro fertilization
- Sex selected semen and embryos

| Eng. Name      | Per. name | Family  | Female name    | Male name       | Birth proc.              | M&F n.b. name      | Male n.b. name        | Fem. n.b. name        | Cast. male name |
|----------------|-----------|---------|----------------|-----------------|--------------------------|--------------------|-----------------------|-----------------------|-----------------|
| Cattle         | گاو       | Bovine  | Cow            | Bull            | Calving<br>(گوساله زایی) | Calf<br>(گوساله)   | Bull calf             | Heifer<br>(تلیسه)     | Steer           |
| Goat           | بز        | Caprine | Doe<br>(Nanny) | Buck<br>(Billy) | Kidding<br>(بزغاله زایی) | Kid<br>(بزغاله)    | Buckling              | Doeling               | Wether          |
| Sheep          | گوسفند    | Ovine   | Ewe<br>(میش)   | Ram<br>(قوچ)    | Lambing<br>(بره زایی)    | Lamb<br>(بره)      | Ram lamb<br>(بره قوچ) | Ewe lamb<br>(بره میش) | Wether          |
| Horse          | اسب       | Equine  | Mare           | Stallion        | Foaling                  | Foal<br>(کره اسب)  | Colt                  | Filly                 | Geling          |
| Swine<br>(pig) | خوک       | Porcine | Sow            | Boar            | Farrowing                | Piglet<br>(litter) | Boar                  | Gilt                  | Barrow          |
| Dog            | سگ        | Canine  | Bitch          | Dog             | Whelping                 | Puppy<br>(litter)  | -                     | -                     | -               |
| Cat            | گربه      | Feline  | Queen          | Tom             | Kindling                 | Kitten<br>(litter) | -                     | -                     | -               |

| Eng. Name | Per. name | Family | Female name    | Male name       | Birth proc.              | M&F n.b. name      | Male n.b. name | Fem. n.b. name           | Cast. male name |
|-----------|-----------|--------|----------------|-----------------|--------------------------|--------------------|----------------|--------------------------|-----------------|
| Cattle    | گاو       | Bovine | Cow            | Bull            | Calving<br>(گوساله زایی) | Calf<br>(گوساله)   | Bull calf      |                          |                 |
|           |           |        | Doe<br>(Nanny) | Buck<br>(Billy) | Kidding<br>(بزغاله زایی) | Kid<br>(زغال)      |                | g                        | Wether          |
|           |           |        |                |                 |                          |                    |                | Ewe<br>lamb<br>(بره میش) | Wether          |
|           |           |        |                |                 |                          |                    | Colt           | Filly                    | Geling          |
|           |           |        |                |                 |                          |                    | Boar           | Gilt                     | B               |
| Dog       | سگ        | Canine |                |                 |                          |                    |                |                          |                 |
| Cat       | گربه      | Feline | Queen          | Tom             | Kindling                 | Kitten<br>(litter) |                |                          |                 |



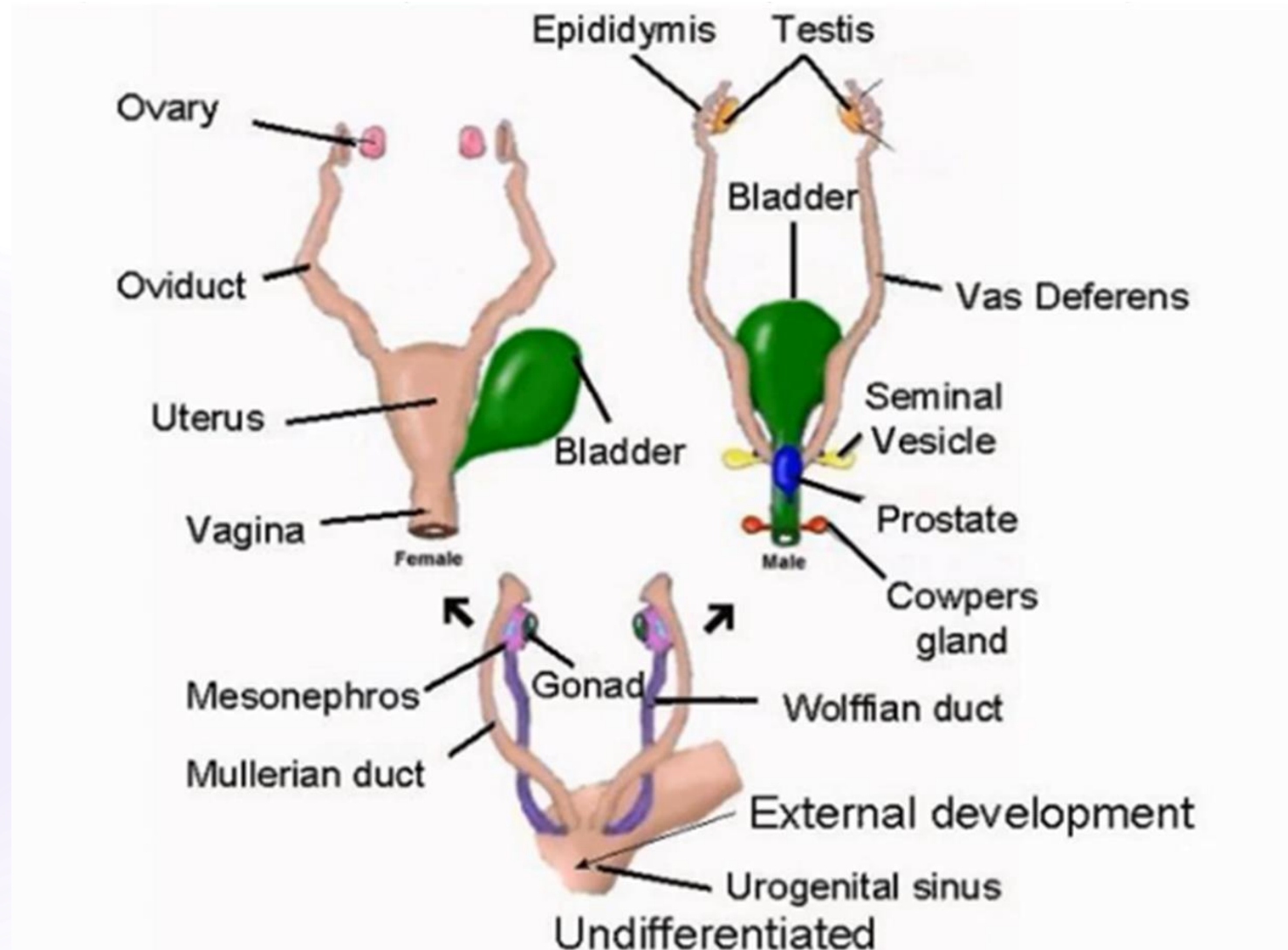
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## Section I

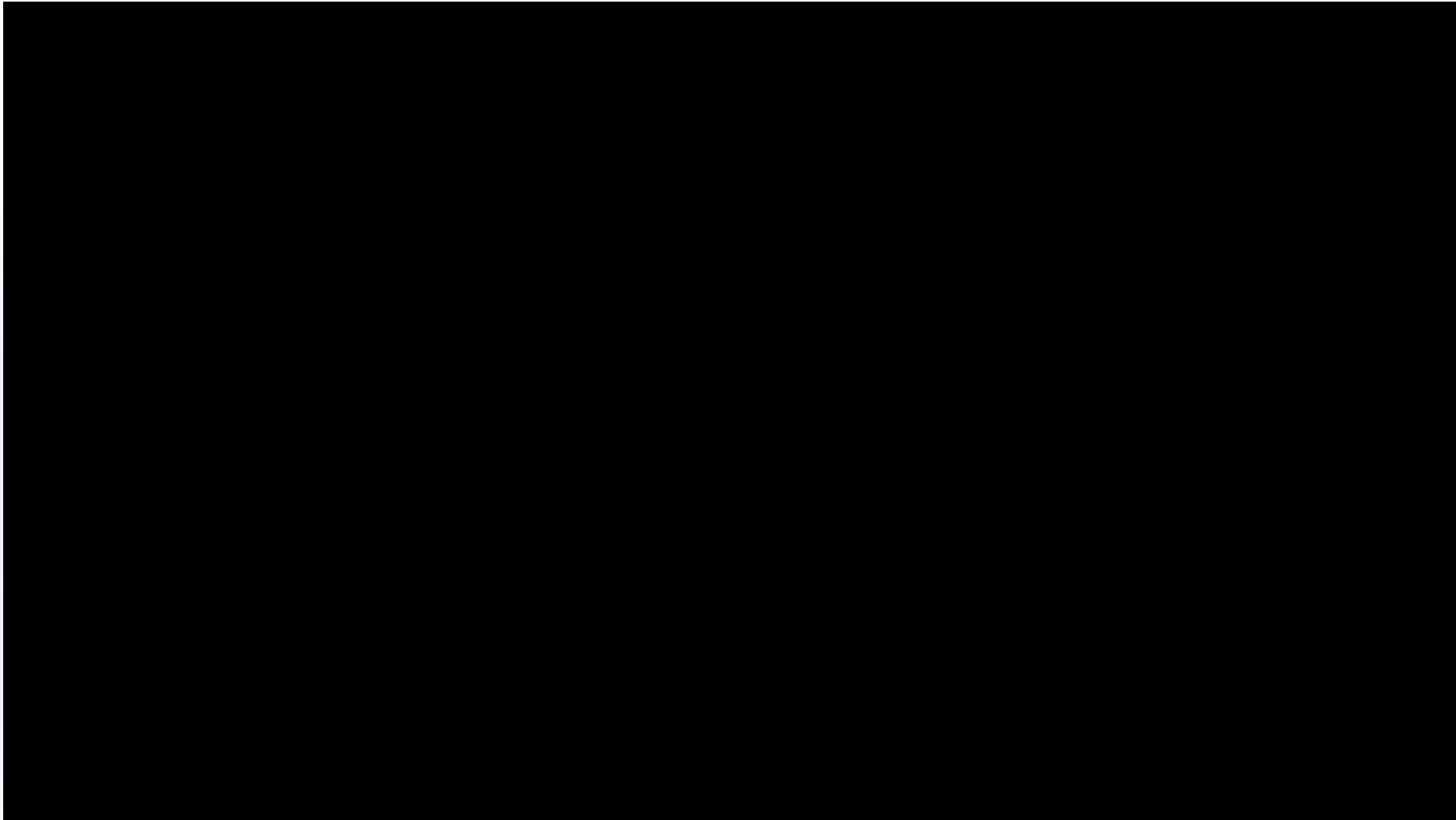
# *Prenatal Development*

# Ontogenesis of the Female Genitalia



# *Sex Determination and Differentiation*

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# *Sex Determination and Differentiation*

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# Journal Club (for more reading)

▶ <https://www.nejm.org/doi/pdf/10.1056/NEJMra022784?articleTools=true>

**REVIEW ARTICLE**

**MECHANISMS OF DISEASE**

## Sex Determination and Differentiation

David T. MacLaughlin, Ph.D., and Patricia K. Donceel

**SEX DETERMINATION**, which depends on the complement of the embryo, is established by multiple molecular events. In the presence of the Y chromosome (or absence of the X chromosome and the presence of a second Y chromosome), the sex of the embryo is determined. Sex determination sets the stage for sex differentiation, the process by which the gonads and the rest of the body develop. A number of genes have been discovered that control the process of sex determination and differentiation. In many cases, these genes have been discovered through studies of either spontaneous or engineered mouse mutants. We will examine how these genes have been discovered and how they have influenced clinical syndromes (Table 1 and Fig. 1) and discuss important clinical syndromes at the molecular level. We will continue to elude classification of the molecular level. Knowledge of disorders of sex determination and differentiation pathways, strong influence on the diagnosis and management of these disorders when possible, adheres to that used in the Online Mammalian Genome Project (http://www.ncbi.nlm.nih.gov).

**GERM CELLS**

Primordial germ cells, which eventually localize in the gonad, differentiate and migrate from the epiblast, the outer ectodermal layer of the embryo, whence they migrate along the primitive streak and then to the base of the allantois (Fig. 2). In the urogenital ridge, the site of the future gonad (Fig. 2). Interactions between these primordial germ cells have recently been elucidated. In the urogenital ridge, where its expression is highest, the fate of these primordial germ cells are regulated by genes that are unique to the differentiating germ cells: *Sox9*, which is expressed in the proximal epiblast, and *Sox2*, which is expressed in the distal epiblast. Inactivation of *Sox9* in the urogenital ridge causes a severe form of intersex, in which the gonads are of mixed sex. Inactivation of *Sox2* in the urogenital ridge causes a severe form of intersex, in which the gonads are of mixed sex. Inactivation of *Sox9* in the urogenital ridge causes a severe form of intersex, in which the gonads are of mixed sex. Inactivation of *Sox2* in the urogenital ridge causes a severe form of intersex, in which the gonads are of mixed sex.

**MALE GERM CELLS**

The proliferation patterns of male and female germ cells differ. XY germ cells undergo mitosis during embryonic development, but soon after reaching the gonads, their migration becomes arrested and they remain within the testis in the quiescent (G<sub>0</sub>) phase of the cell cycle until after birth under the influence of an unknown inhibitory factor (referred to as meiosis inhibitory factor) secreted by either Sertoli or myoid cells (Fig. 2). After birth, the male germ cells resume the cell cycle and undergo meiotic division, which halves the number of chromosomes to produce haploid cells, which complete spermatogenesis at puberty.

**SYNDROMES OF ABSENT GERM CELLS AND RELATION OF GERM CELLS TO STEM CELLS**

Germ cells are absent in the mutant strain of piebald mice<sup>10</sup> and in "Sertoli-only"<sup>11</sup> testes of infertile men who have deletions in the long arm of the Y chromosome in the azoospermia factor (AZF) region. The Y chromosome is the sex-determining region of the genome that controls spermatogenesis.<sup>12</sup> The recent elucidation of the sequence of the human Y chromosome<sup>13</sup> will provide a template to further our understanding of the structure and function of this chromosome, particularly of the elusive long arm (q) region. Stem-cell factor,<sup>14</sup> a ligand also known as mast-cell growth factor that is encoded by the *steel* locus on chromosome 11 and is important in the development of stem cells. Stem-cell factor, a ligand also known as mast-cell growth factor that is encoded by the *steel* locus on chromosome 11 and is important in the development of stem cells.

**Table 1. Mutations in Genes Involved in Sex Determination and Differentiation**

| Gene (Locus)                             | Protein and Proposed Function  |
|--|--|
| WT1 (11p13)                              | Transcription factor   |
| SF-1 (9q33)                              | Transcription factor, nuclear receptor                                       |
| SOX9 (17q24)                             | High-mobility-group transcription factor                                     |
| DAXI (9p21.3)                            | Transcriptional regulator, nuclear-receptor protein                          |
| SRY (Yp11)                               | High-mobility-group transcription factor                                     |
| MIS, or AMH, type II receptor (12q12-13) | Serine/threonine kinase receptor   |
| MIS, or AMH (19p13)                      | Secreted protein, causes regression of müllerian duct; Leydig-cell inhibitor |
| AR (Xq11-12)                             | Androgen receptor, a ligand transcription factor                             |
| HSD17B3 (9q22)                           | 17β-Hydroxysteroid dehydrogenase, 17-ketosteroid reductase                   |
| SRD5A2 (5p15)                            | 5α-Reductase type 2  |
| CYP17 (10q24-25)                         | 17-Hydroxylase; 20-22 lyase  |
| CYP21 (6q21.3)                           | 21-Hydroxylase   |
| HSD3B2 (1p13.1)                          | 3β-Hydroxysteroid dehydrogenase type II                                      |
| CYP11B1 (8q24)                           | 11β-Hydroxylase  |
| SAR (9p11.2)                             | Steroidogenic acute regulatory protein                                       |

\* Virilization may occur at puberty.

**Figure 1. Clinical Examples of Intersex Abnormalities.**

Panel A shows the internal structures of a 46,XY patient with pure gonadal dysgenesis. Panel B shows the internal structures of a 46,XX patient with femoral hypoplasia and congenital adrenal hyperplasia. Panel C shows micropenis and penoscrotal hypospadias in a 46,XY patient with persistent müllerian duct syndrome. Panel D shows internal genitalia of a 46,XY patient with persistent müllerian duct syndrome as well as wolffian structures (i.e., fallopian tubes and uterus) with the permission of the publisher.

**Figure 2. Migration and Proliferation of Germ Cells during Embryonic and Fetal Development.**

Germ cells are first detected in the epiblast, where they are activated by bone morphogenetic protein (BMP) from the notochord. Migration occurs through the primitive streak to the base of the allantois. Subsequently, the cells migrate to the urogenital ridge, where the gonads for

**Figure 3. Syndromes of Dysgenesis during the Development of the Urogenital Ridge.**

Mutations in various genes can lead to a variety of syndromes of dysgenesis involving the müllerian or wolffian ducts, gonads, kidneys, and adrenal glands as a result of a deficiency or excess of the proteins shown. DAX1 denotes the gene for duplicated in adrenal hypoplasia congenita on the X chromosome 1; Emx2 the empty spiracle homeobox gene 2; GATA4 the gene encoding a protein that binds to a GATA DNA sequence; HMOX1 homeobox protein 1; Lmo2 a homeobox gene important for limb development; Lhx9 a LIM homeobox family member; Pax2 paired box homeotic gene; Sf-1 the gene for steroidogenic factor 1; Sry sex-determining region of the Y chromosome; Sox9 SRY homeobox 9; Wnt-4 a protein that induces the development of the müllerian mesenchyme; and Wt1 Wilms' tumor-suppressor gene 1.

**GONADAL AND RENAL ABNORMALITIES**

The Frasier syndrome is characterized by both gonadal and renal abnormalities that result in streak gonads coupled with the nephrotic syndrome (Fig. 3). If it occurs in the XY genotype then there is sex reversal. Study of the phenotype of Wt1-deficient mice revealed that the gene is involved in the early steps of the differentiation of both gonads and kidneys, helping to explain the association of gonadal and kidney malfunction in the Frasier syndrome.

Alternative splicing of the Wt1 gene in mice can result in up to 24 protein isoforms. Mutations of two of these isoforms lead to striking clinical manifestations of the phenotypes of mice in which candidate genes have been inactivated by homologous recombination (knockout mice), have increased our understanding of the pathophysiology of some of the

from fetal specimens<sup>17</sup> or from excess blastocysts generated by in vitro fertilization protocols.<sup>18</sup> These developments have increased our understanding of factors affecting pluripotency and have fueled hopes that therapeutic cloning can be used to create differentiated cell types for replacement therapy. Thus germ-cell biology has contributed to the development of stem-cell biology. In turn, discoveries regarding pluripotency have also led to myriad ethical controversies<sup>19</sup> and initiated steps to ensure that cells will not be used unlawfully for reproductive cloning of humans.

**SYNDROMES OF GONADAL DYSGENESIS**

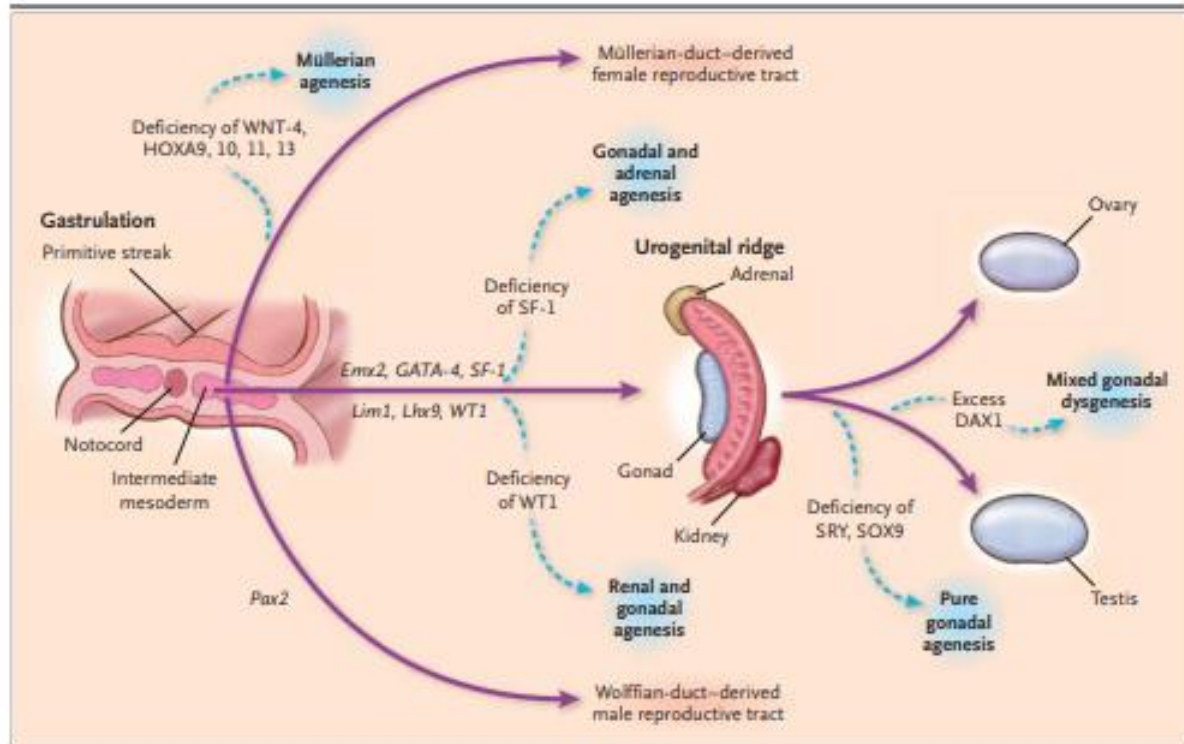
Investigations of the molecular events that occur during sex determination, coupled with an analysis of the phenotypes of mice in which candidate genes have been inactivated by homologous recombination (knockout mice), have increased our understanding of the pathophysiology of some of the

maintenance (+), respectively, of three amino acids: lysine (K), threonine (T), and serine (S) between the third and fourth zinc fingers of the DNA-binding domain of this transcription factor. Hammes et al.<sup>20</sup> found that altering the expression of KTS in mice influences both kidney and testicular function. In the Frasier syndrome, the splice site of Wt1 that normally preserves the KTS triplet is mutated, therefore, patients with the syndrome produce only Wt1 protein without KTS. Gonads lacking KTS have decreased production of the sex-determining region of the Y chromosome (SRY), a urogenital ridge protein that is critical for testicular differentiation. In these -KTS gonads there is also a decrease in müllerian-inhibiting substance, a glycoprotein hormone derived from Sertoli cells that causes regression of the male müllerian ducts and whose presence is an early marker of testicular differentiation.<sup>21</sup> The findings

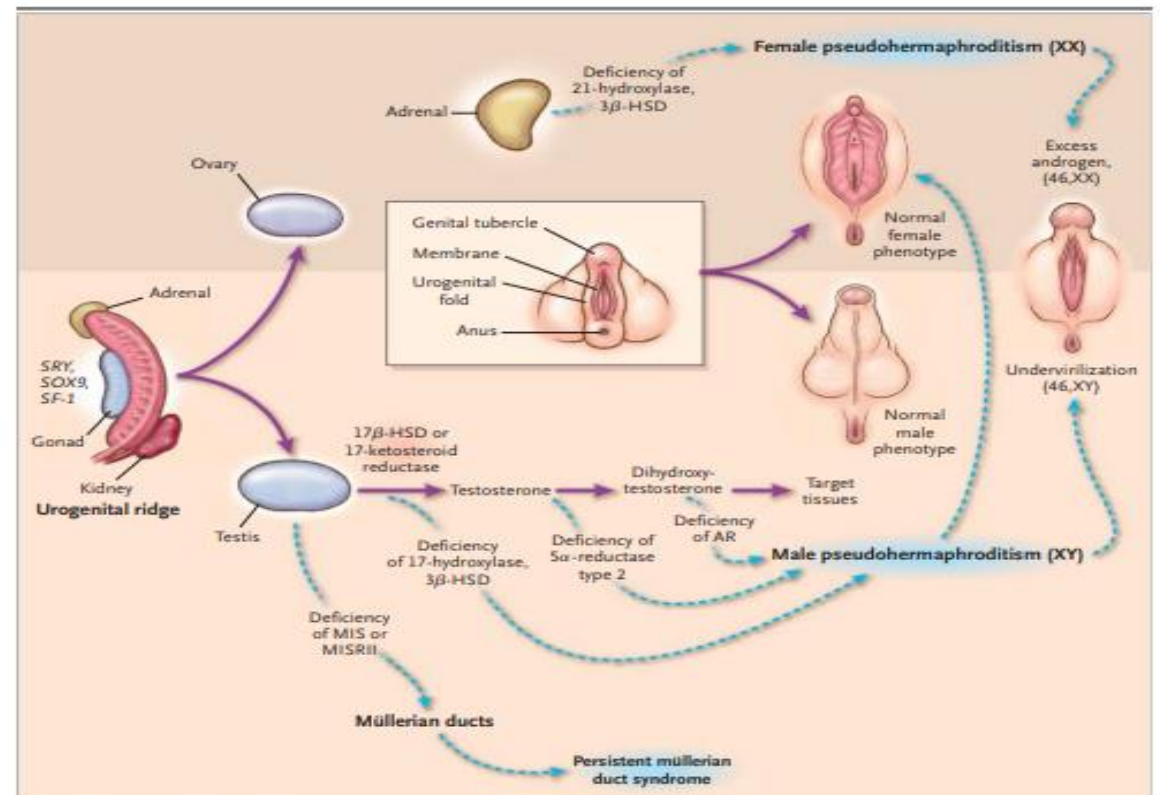
# Journal Club (for more reading)



► <https://www.nejm.org/doi/pdf/10.1056/NEJMra022784?articleTools=true>



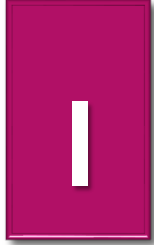
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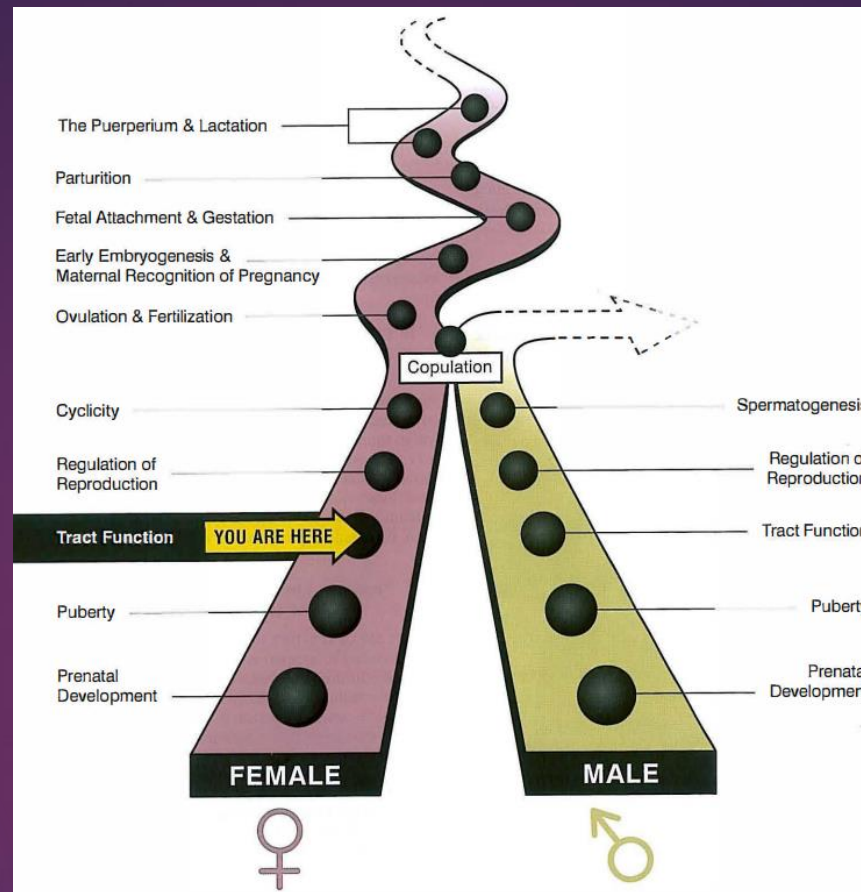


**Figure 4. Functional Abnormalities of the Synthesis and Action of Hormones.**  
 After the gonads have formed, reduced hormonal activity or signaling of specific receptors can lead to functional abnormalities of the reproductive tract, including persistent müllerian duct syndrome; male pseudohermaphroditism, causing undervirilization; and müllerian agenesis. After adrenal development, reduced enzymatic activity can result in female pseudohermaphroditism with excessive virilization. HSD denotes hydroxysteroid dehydrogenase, MIS müllerian inhibiting substance, MISRII müllerian inhibiting substance type II receptor, *SF-1* the gene for steroidogenic factor 1, *SRY* the gene for the sex-determining region of the Y chromosome, *SOX9* the gene for SRY homeobox 9, and *AR* androgen receptors.

# *Sex and Gender (for more reading)*

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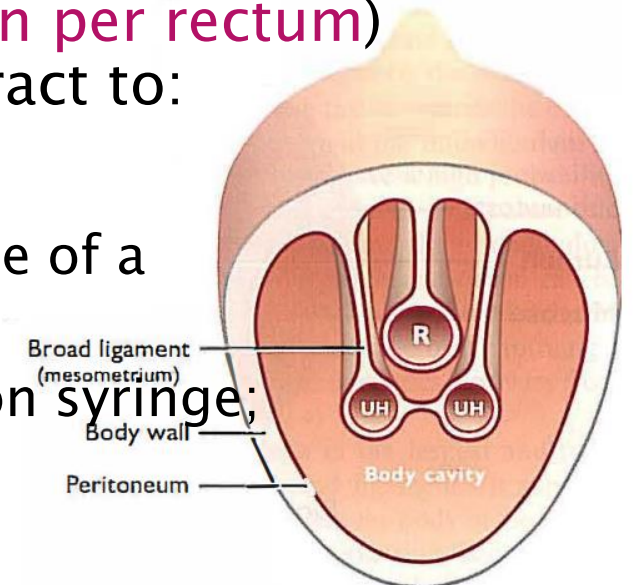
## Section II

# *The Organization and Function of the Female Reproductive Tract*

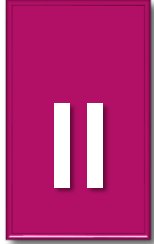
# Female Reproductive Tract



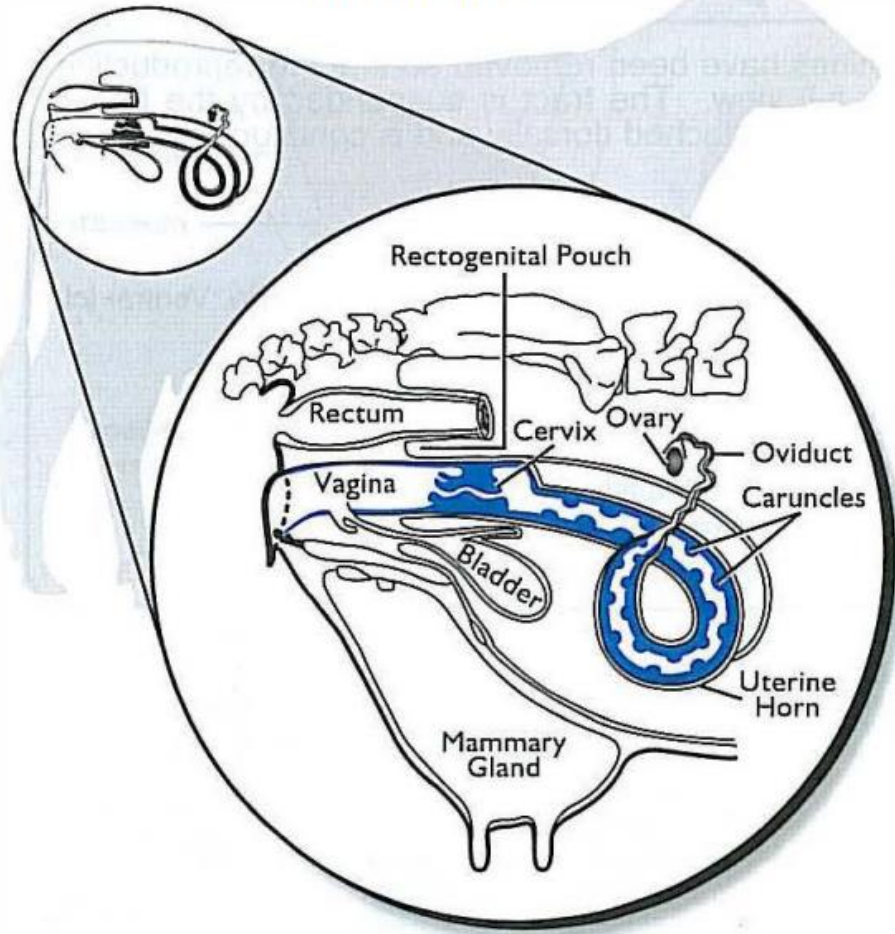
- The major structures of the female reproductive tract include the **ovaries** (the female gonads), **oviducts**, **uterus**, **cervix**, **vagina** and **external genitalia**. Each of these organs may be subdivided into components that represent specific anatomical regions.
- In all domestic species, the reproductive tract lies directly beneath the rectum and is separated from it by the **rectogenital pouch**.
- In the **cow**, **mare**, and **camel** this fortuitous anatomical relationship provides the opportunity for manual palpation (**manipulation per rectum**) and/or **ultrasonic examination** of the female reproductive tract to:
  - 1) diagnose the ovarian status of the female;
  - 2) diagnose pregnancy by determining the presence or absence of a fetus or of fetal membranes located within the uterus;
  - 3) manipulate the tract for insertion of an artificial insemination syringe;
  - 4) recover embryos using nonsurgical techniques
  - 5) identify reproductive tract abnormalities



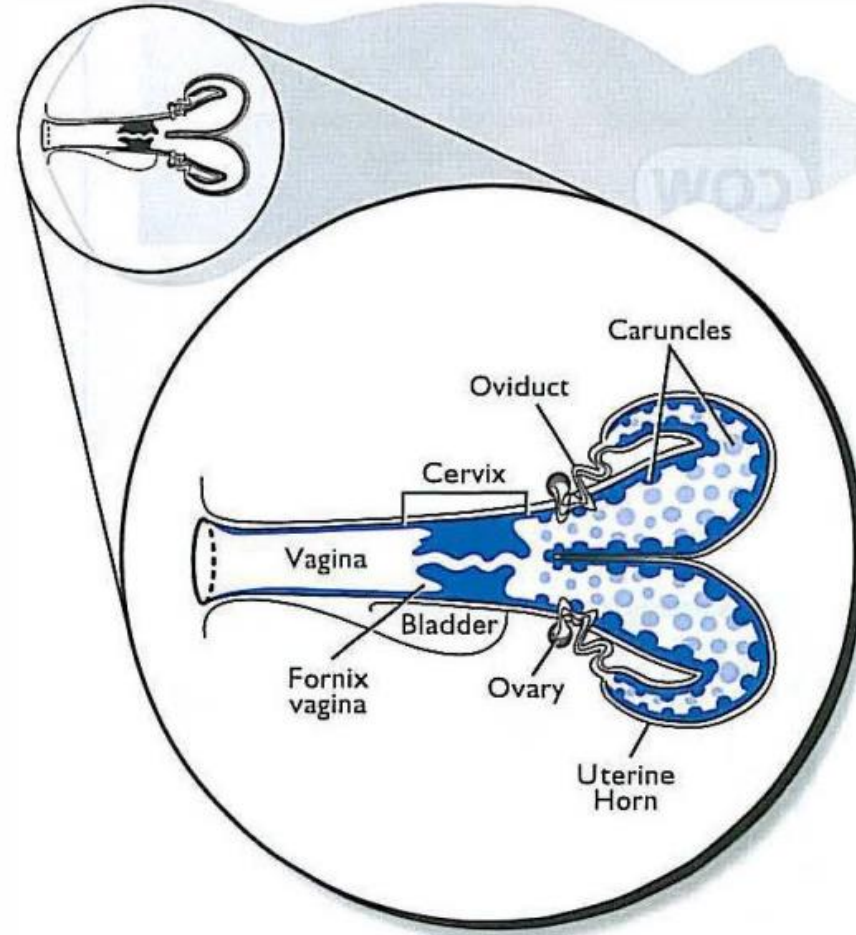
# Ruminant (Cow)



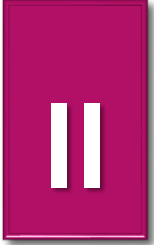
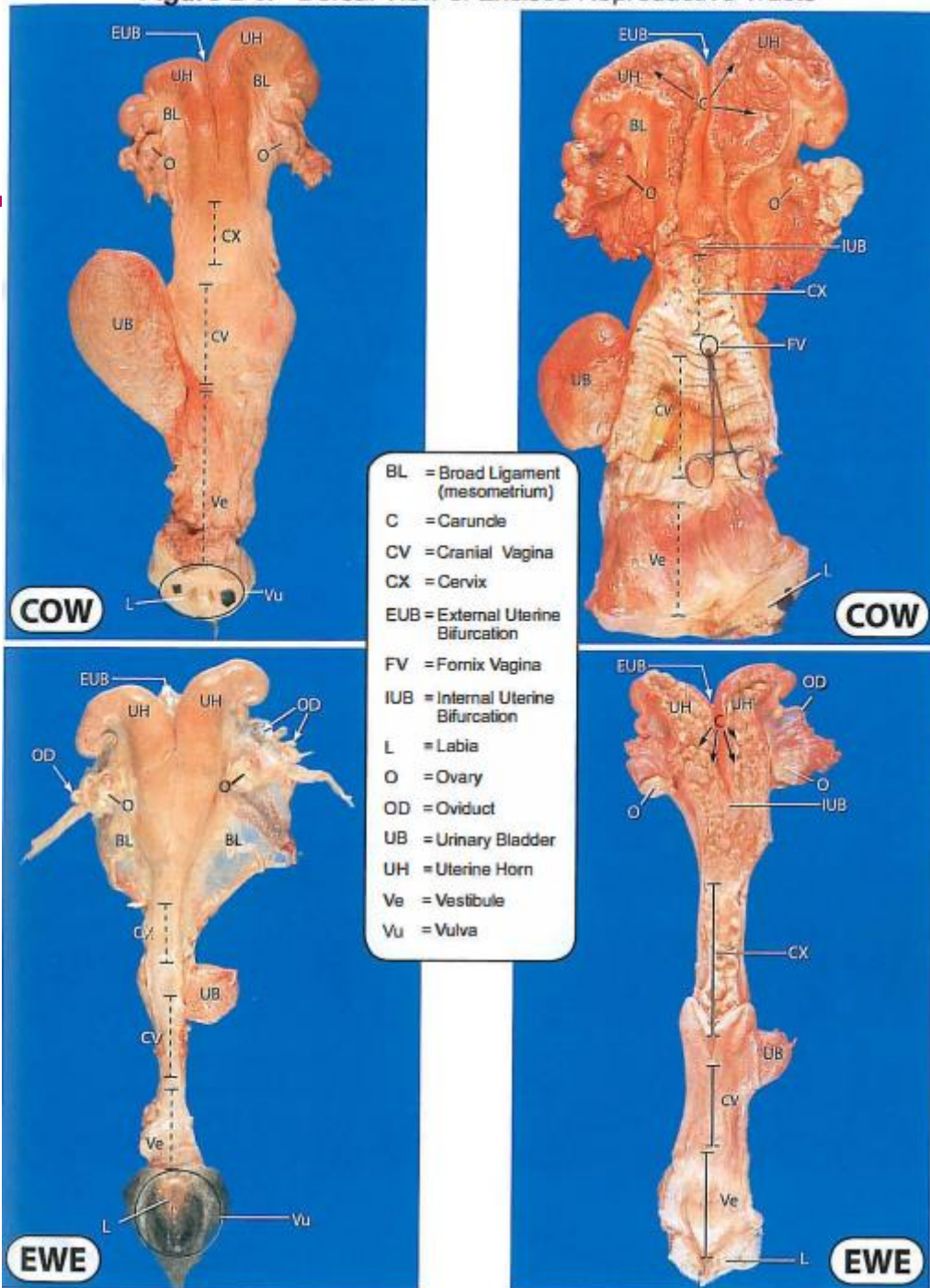
Lateral view



Dorsal view

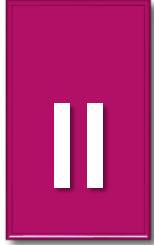


# Ruminant



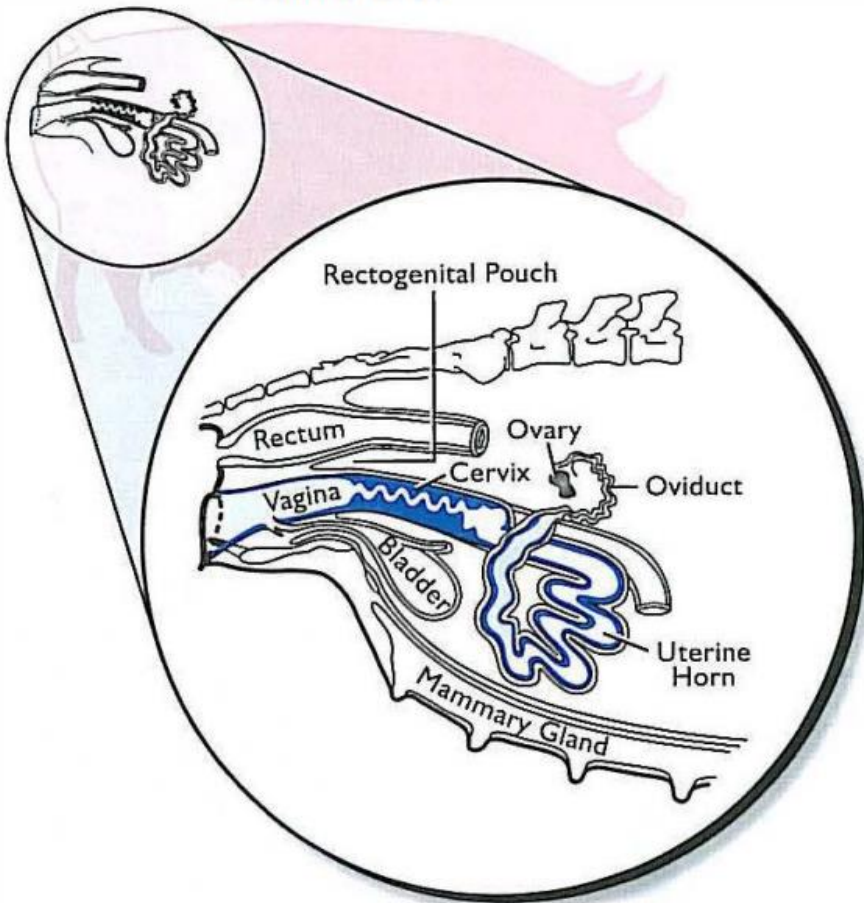


# Sow

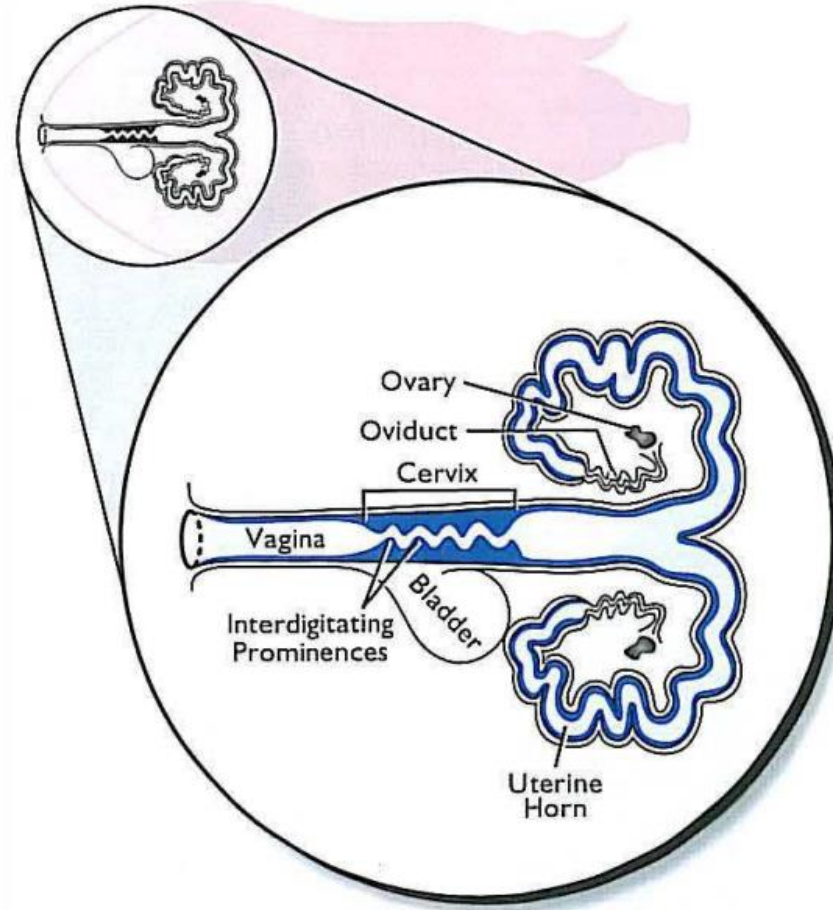


## Sow

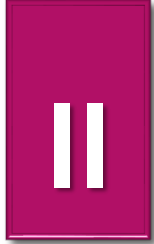
Lateral view



Dorsal view

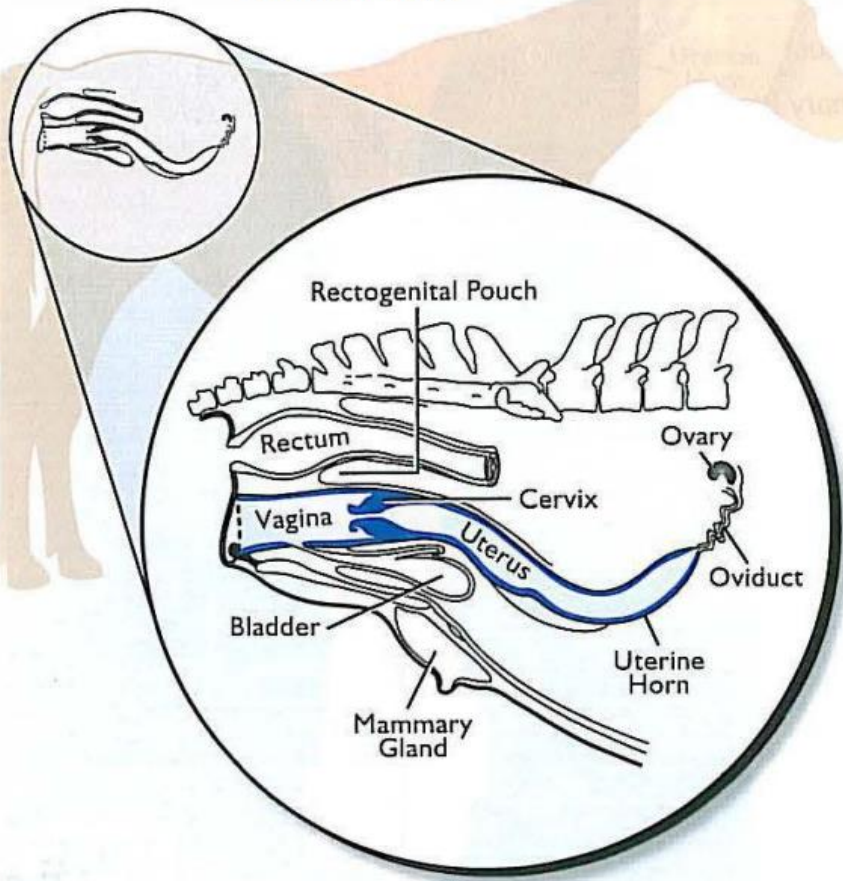


# Mare

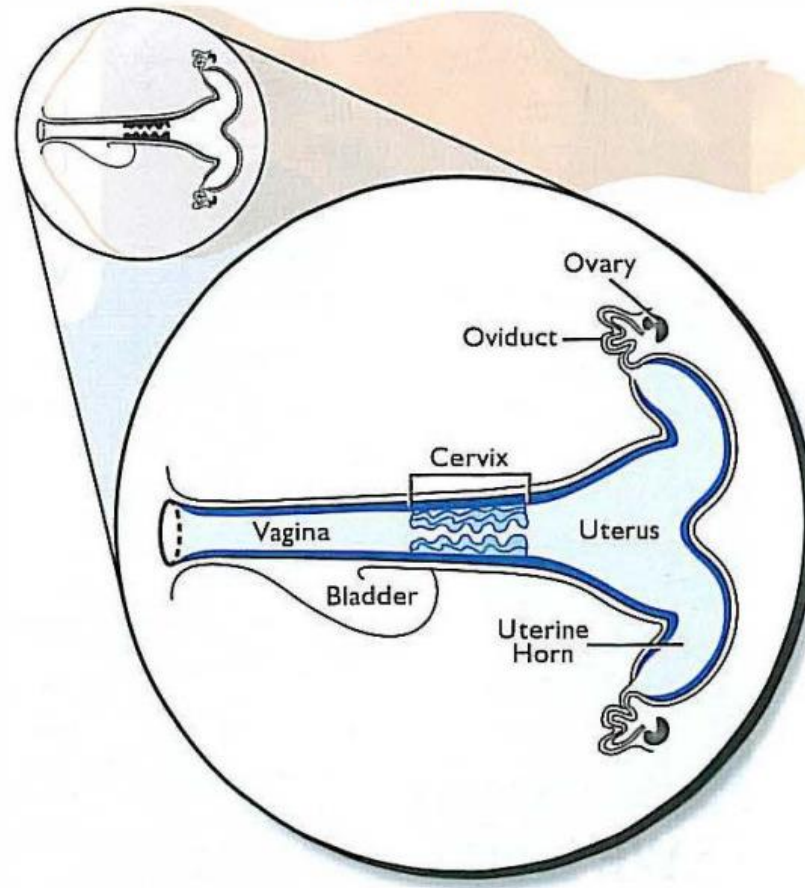


## Mare

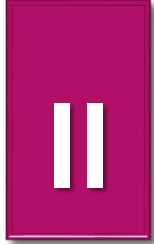
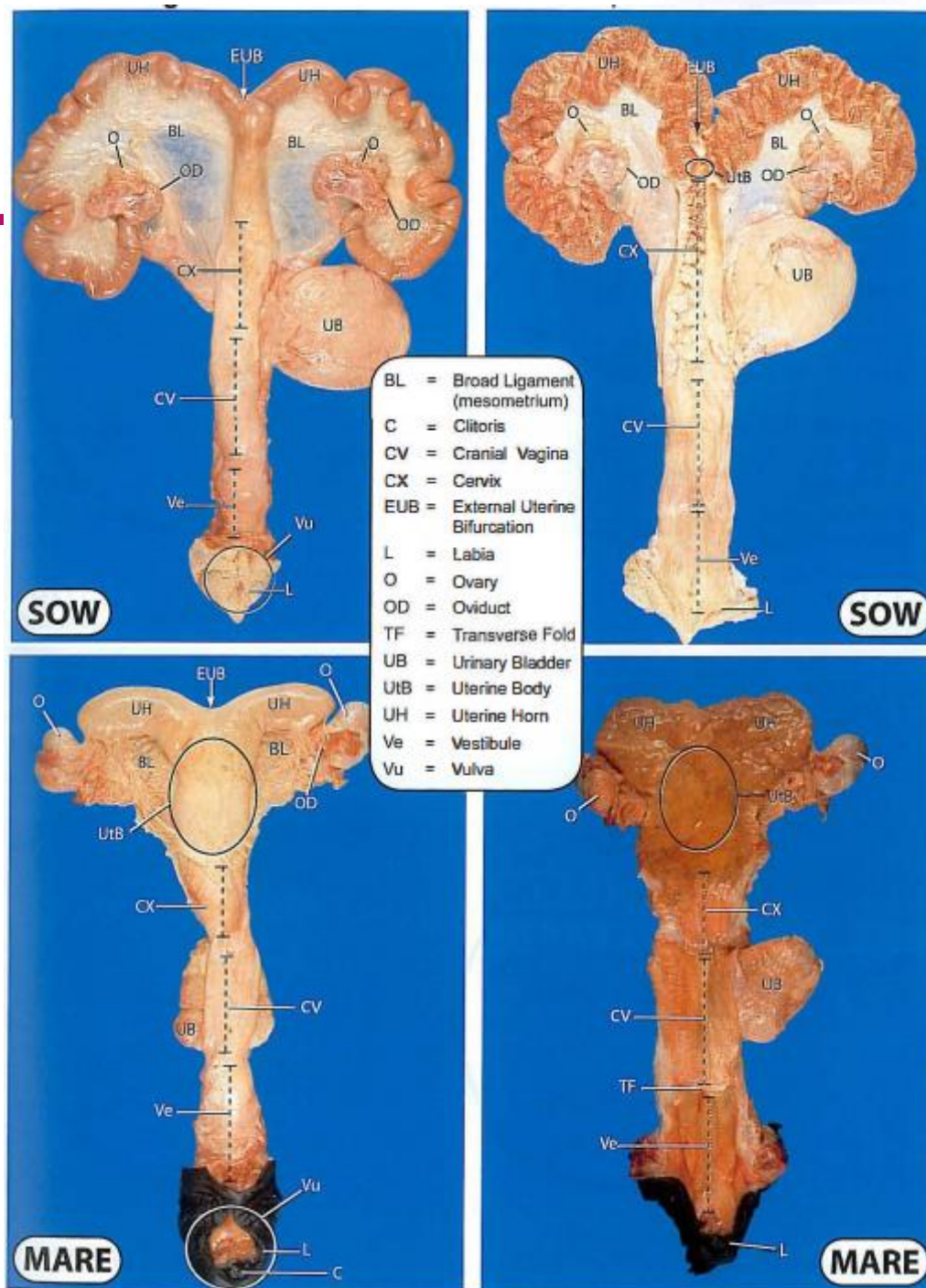
Lateral view



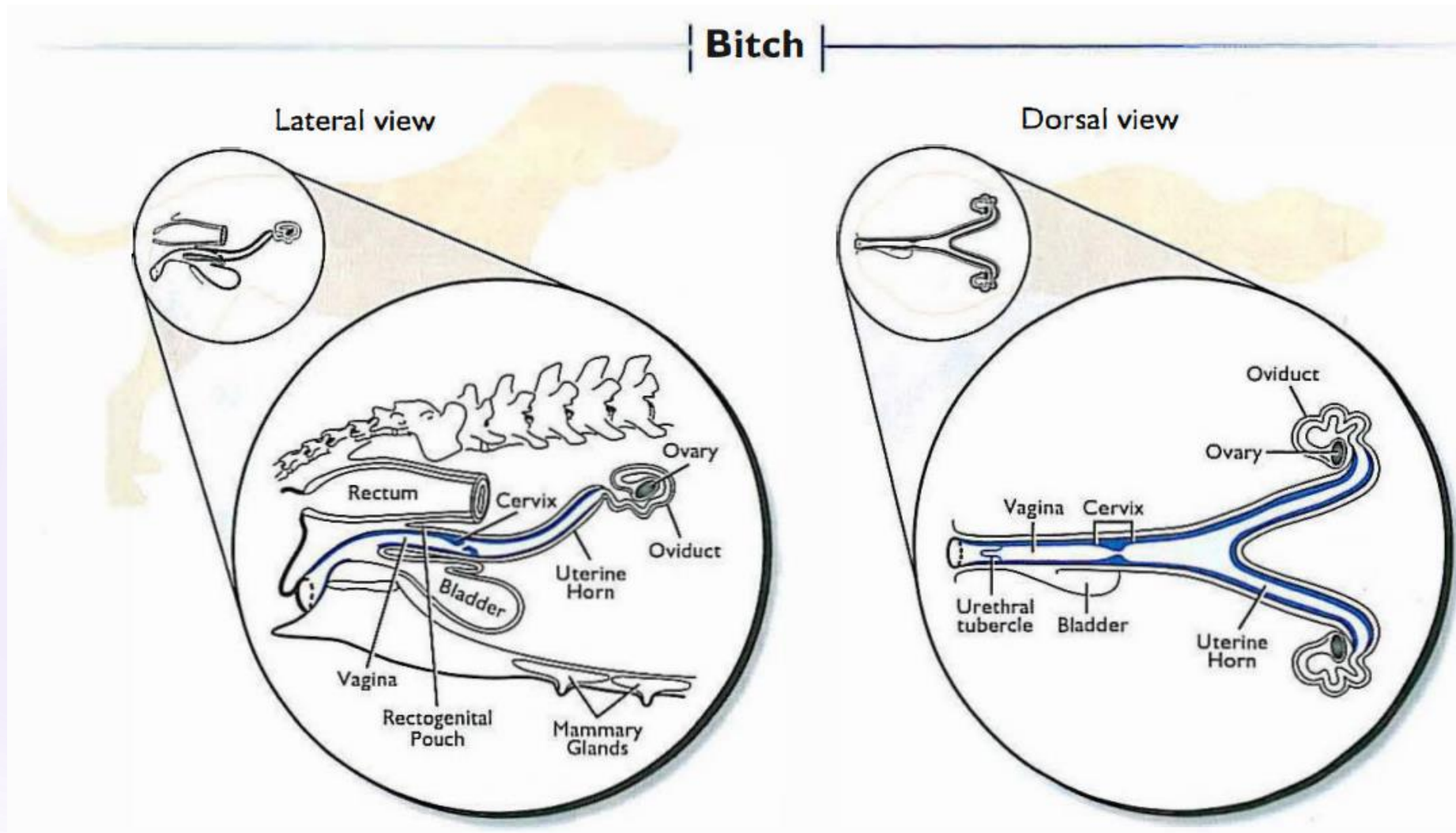
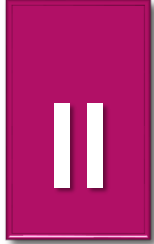
Dorsal view



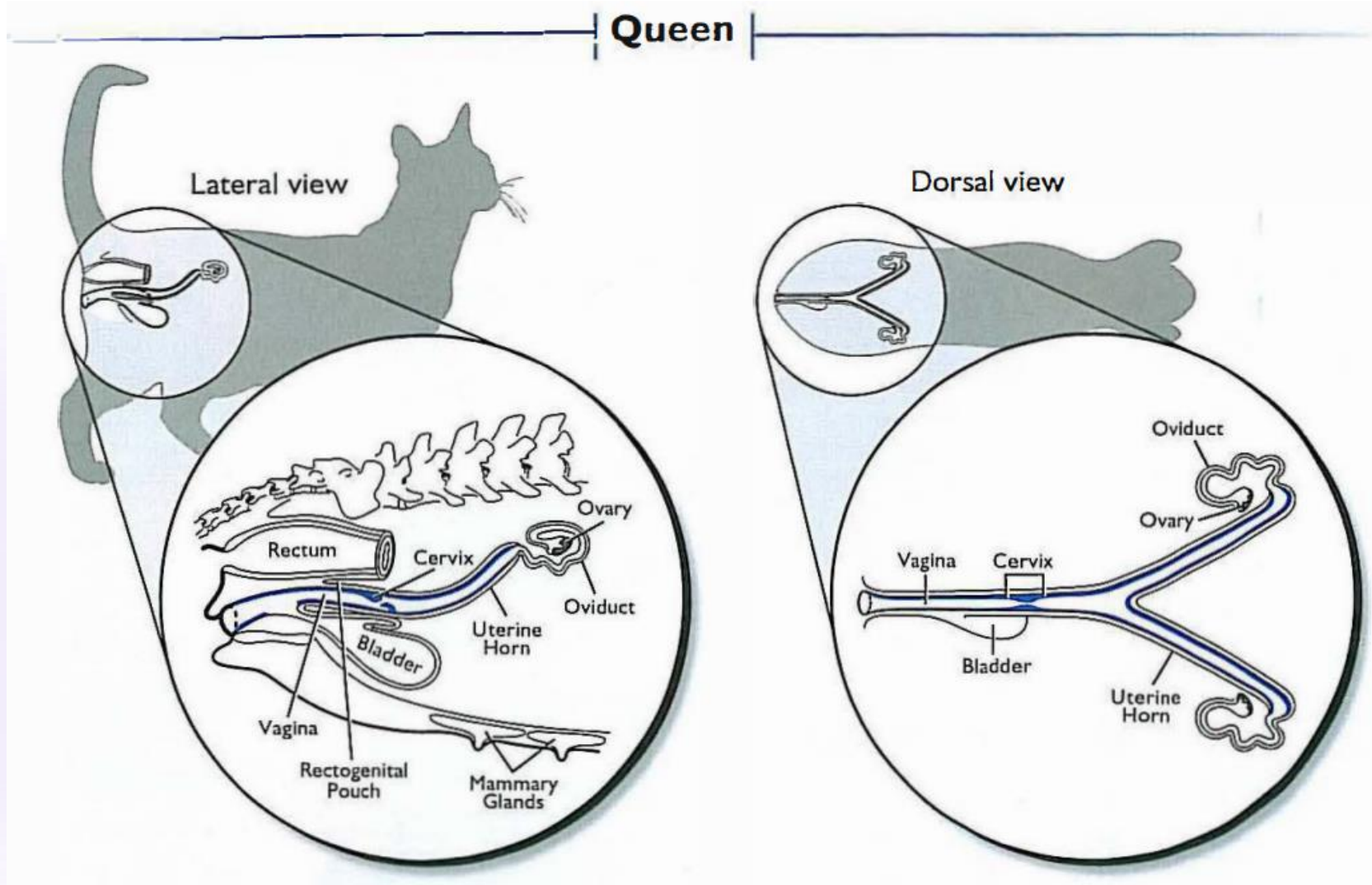
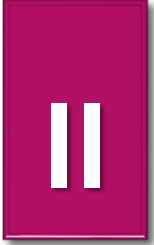
# Sow and Mare



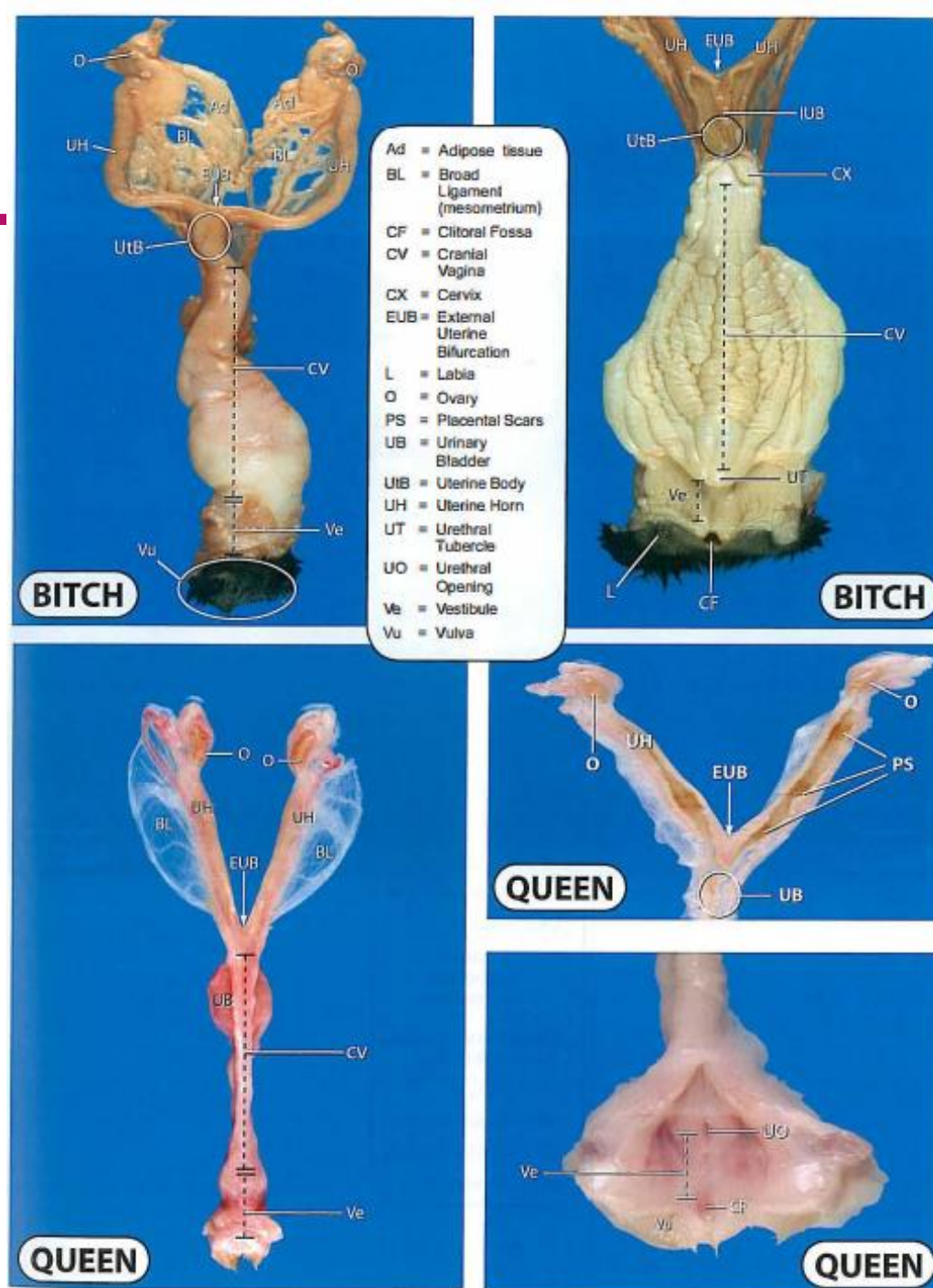
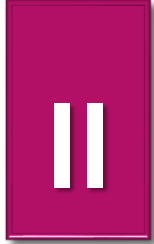
# Carnivores (*Bitch*)



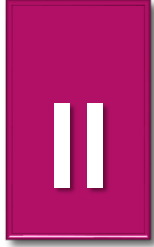
# Carnivores (Queen)



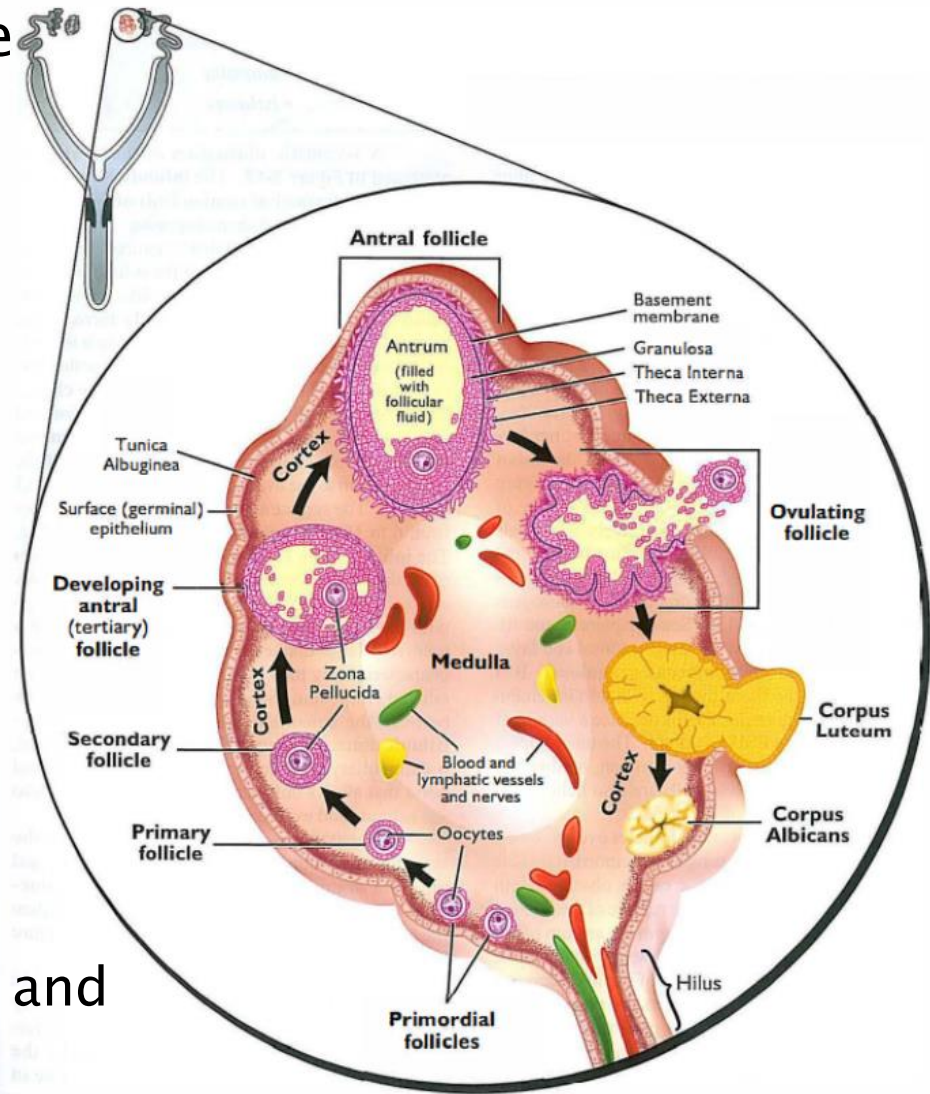
# *Bitch and Queen*



# The ovary

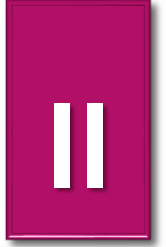


- The **ovary** is an ovoid relatively dense structure, the primary functions of which are to **produce female gametes** (ova) and the hormones **estrogen** and **progesterone**. The **corpus luteum** also produces **oxytocin, relaxin, inhibin** and **activin**.
- The ovary is composed of:
  - Outer connective tissue surface called the **tunica albuginea**
  - Single layer of cuboidal cells called the **germinal epithelium** (wrongly named)
  - The **ovarian cortex** houses the population of oocytes
    - **follicles** that will mature and eventually ovulate.
    - **corpus luteum** (CL)
    - **corpus albicans**
  - The **ovarian medulla**: vasculature, nerves, lymphatics and relatively dense connective tissue.



# The ovary

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- **Ovarian follicles:** The process whereby immature follicles develop into more advanced follicles and become candidates for ovulation is referred to as folliculogenesis.
  - **Primordial follicles** that are microscopic, are the most immature and are the smallest encountered in the ovarian cortex. The oocyte ( egg) within the primordial follicle is surrounded by a single layer of flattened (squamous) cells.
  - **Primary follicle** is characterized by having an oocyte that is Surrounded by a single layer of cuboidal or follicular cells.
    - they either develop into a more advanced secondary follicle or they degenerate.
  - **Secondary follicle**, is characterized as having two or more layers of follicle cells, but without an antrum or cavity.
    - the oocyte within a secondary follicle is characterized as being surrounded by a relatively thick translucent layer called the **zona pellucida**.



# The ovary

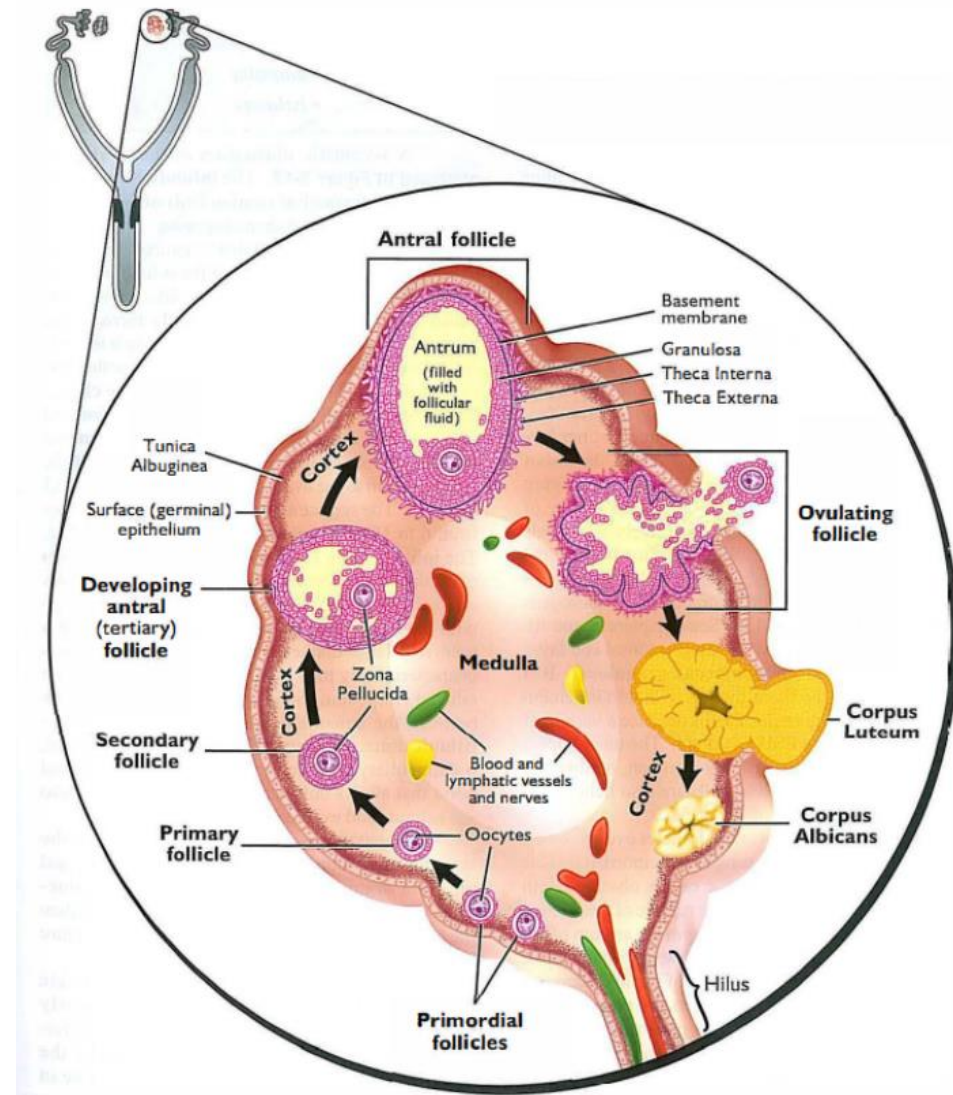
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- **Antral follicle** is characterized by a fluid-filled cavity called the **antrum**. The fluid within the antrum is called **follicular fluid**. Sometimes the antral follicle is referred to as a **tertiary follicle**.
- **Graafian follicle** is the dominant tertiary preovulatory follicle.
  - Antral follicles consist of three distinct layers. Theca externa, the theca interna and the granulosa cell layer.
    - **The theca externa**: loose connective tissue that completely surrounds and supports the follicle.
    - **the theca interna**: responsible for the production of androgens under the influence of LH.
    - **The granulosa cells**: produce a variety of materials and have FSH receptors. The most important products of these cells are estrogen, inhibin and follicular fluid. Granulosa cells are also believed to govern the maturation of the oocyte

# The ovary

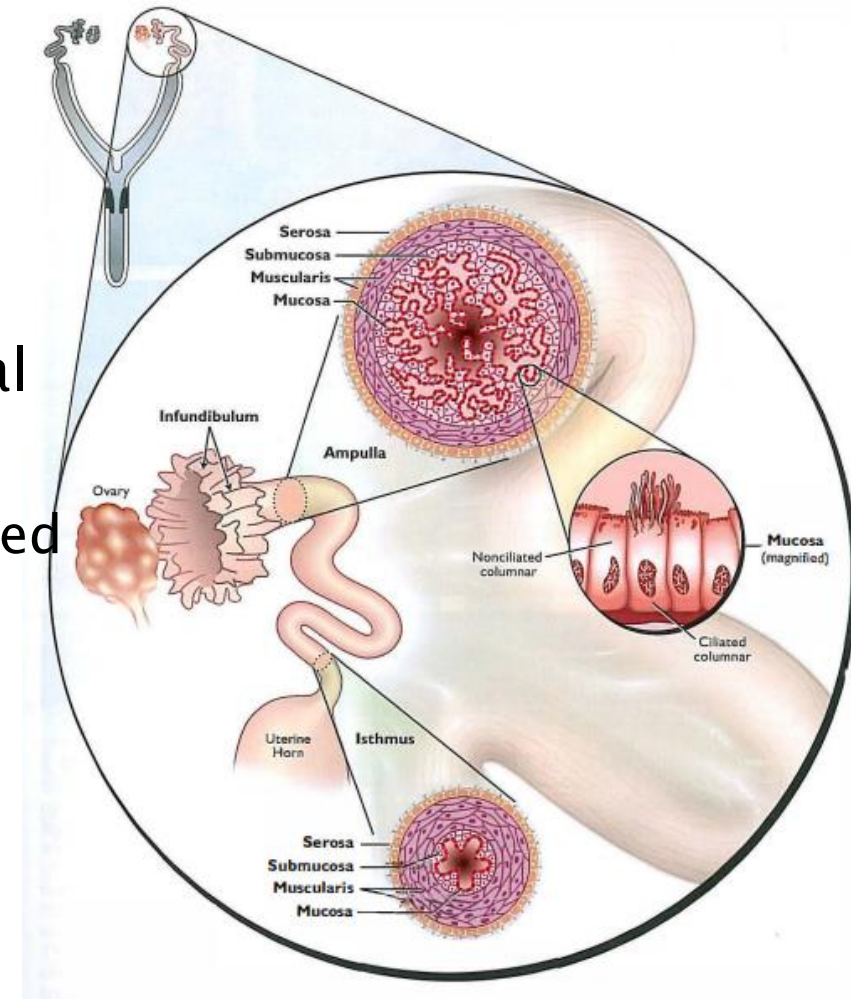
- **corpus hemorrhagicum** forms When dominant antral follicles ovulate and small blood vessels rupture, causing local hemorrhage.
- After the formation of the corpus hemorrhagicum ("bloody body"), the cells of the theca interna and the granulosa cells differentiate into luteal cells to form a **corpus luteum**.



# The Oviduct

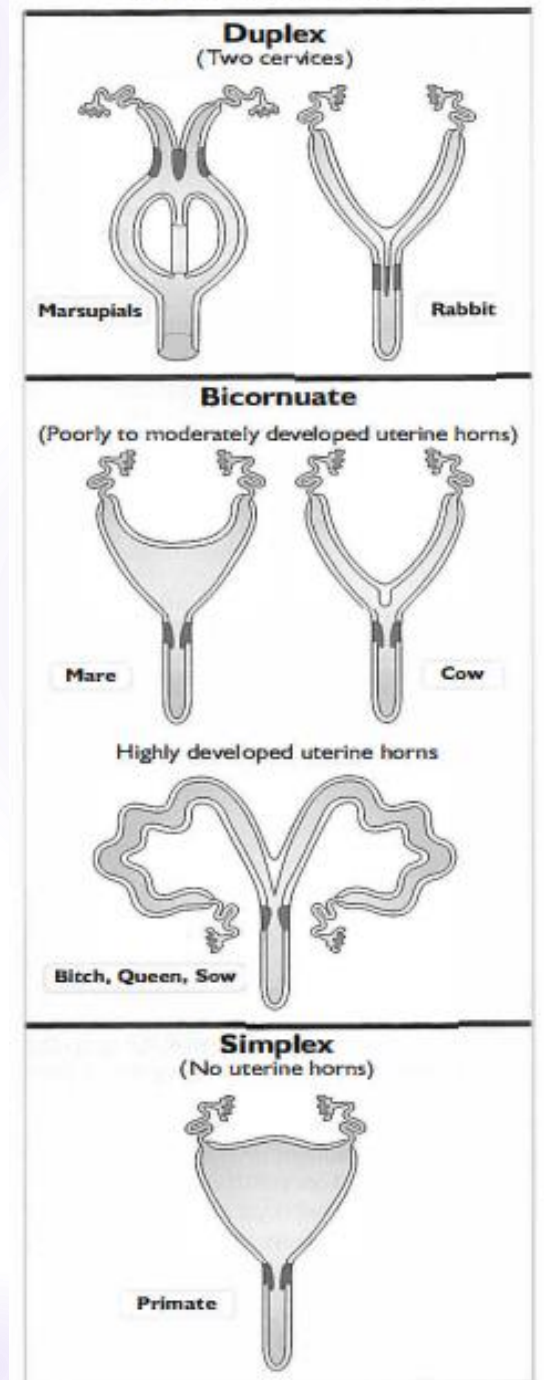


- The **infundibulum** is the terminal end (cranial or ovarian end) of the oviduct and consists of a funnel-shaped opening that "captures" the newly ovulated oocyte.
  - The surface of the infundibulum is covered with many velvety, finger-like projections called **fimbriae**.
- The **ampulla** occupies one-half or more of the oviductal length and merges with the isthmus of the oviduct
  - The **ampullary-isthmus junction** (AIJ) is generally ill-defined and serves as a control point that allows only fertilized oocytes to pass into the isthmus and eventually into the uterus.
- The **isthmus** is smaller in diameter than the ampulla, thicker muscular wall and has fewer mucosal folds.

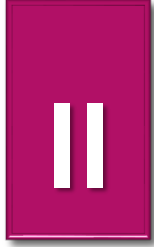


# The Uterus

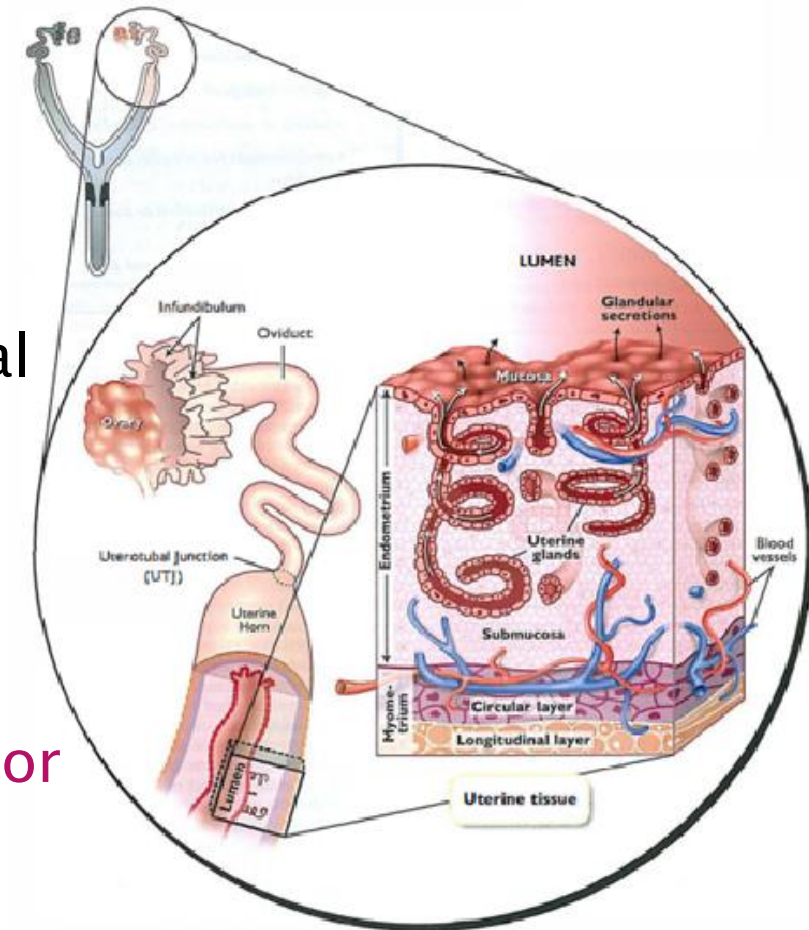
- The uterus connects the oviducts to the cervix. In most mammals, the uterus consists of two **uterine horns** or **cornua**.
- there are three distinct anatomical types of uteri:
  - **Duplex uterus**, characterized as having two cervical canals.
  - **Bicornuate uterus** is characterized by having two uterine horns and a small uterine body
  - **Simplex uterus** is characterized as having a single uterine body



# The Uterus



- The uterus consists of a serosal layer called **the perimetrium** that is part of the peritoneum.
- **The myometrium** consists of two layers of outer longitudinal and inner circular muscle layers.
  - Motility of the uterus a high degree of tone (a partial state of contraction) when **estrogen** is the predominant steroidal hormone.
  - Under the influence of **progesterone**, the myometrium has a low degree of tone
  - During parturition, the myometrium becomes a **major driving force** for expulsion of the fetus and fetal membranes.



# The Uterus

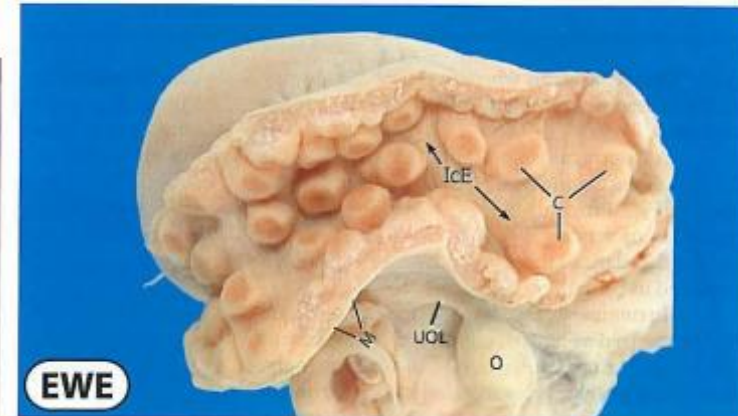
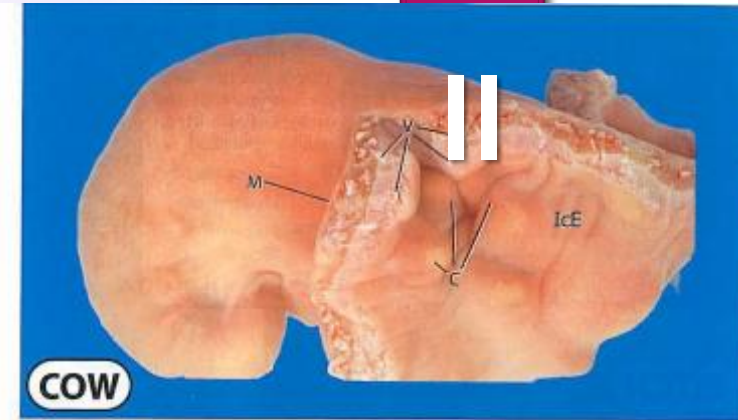
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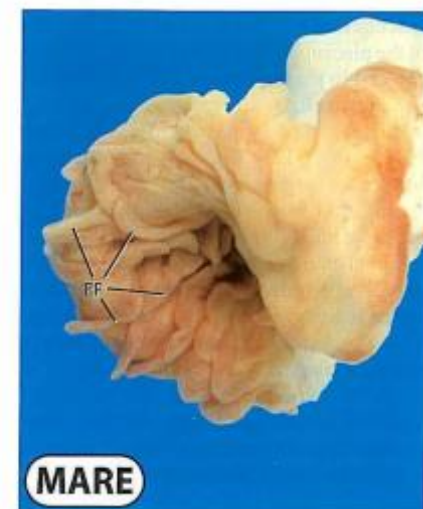
- The inner portion of the uterus is composed of the mucosa and submucosa called **the endometrium**.
  - The mucosal epithelium is responsible for secreting materials into the lumen of the uterus that enhance **embryo development** and **sperm viability**.
  - A distinct difference between lower mammals and primates, particularly humans, is that the endometrium of the uterus in the human is **sloughed to the exterior**.
  - At a critical time during the estrous cycle the cells of the uterine endometrium produce prostaglandin  $F_{2\alpha}$ .
    - $PGF_{2\alpha}$  causes luteolysis or regression of the corpus luteum if the animal is not pregnant.
- **The primary functions of the uterus are:** 1) sperm transport, 2) luteolysis and control of cyclicity, 3) environment for preattachment embryo, 4) maternal contribution to the placenta, 5) expulsion of the fetus and fetal placenta

# The Uterus

- The uterus has been incised so that the endometrial surface can be visualized.
  - In the **cow** and the **ewe**, **caruncles** (C) can be observed as protrusions from the endometrial surface. Blood vessels (V) are white, cord-like structures located beneath the surface of each caruncle.
  - The endometrium of the **sow** and **mare** is characterized as having many **endometrial folds** (EF).
  - Both the caruncles and the endometrial folds contribute to the **maternal placenta** if pregnancy occurs



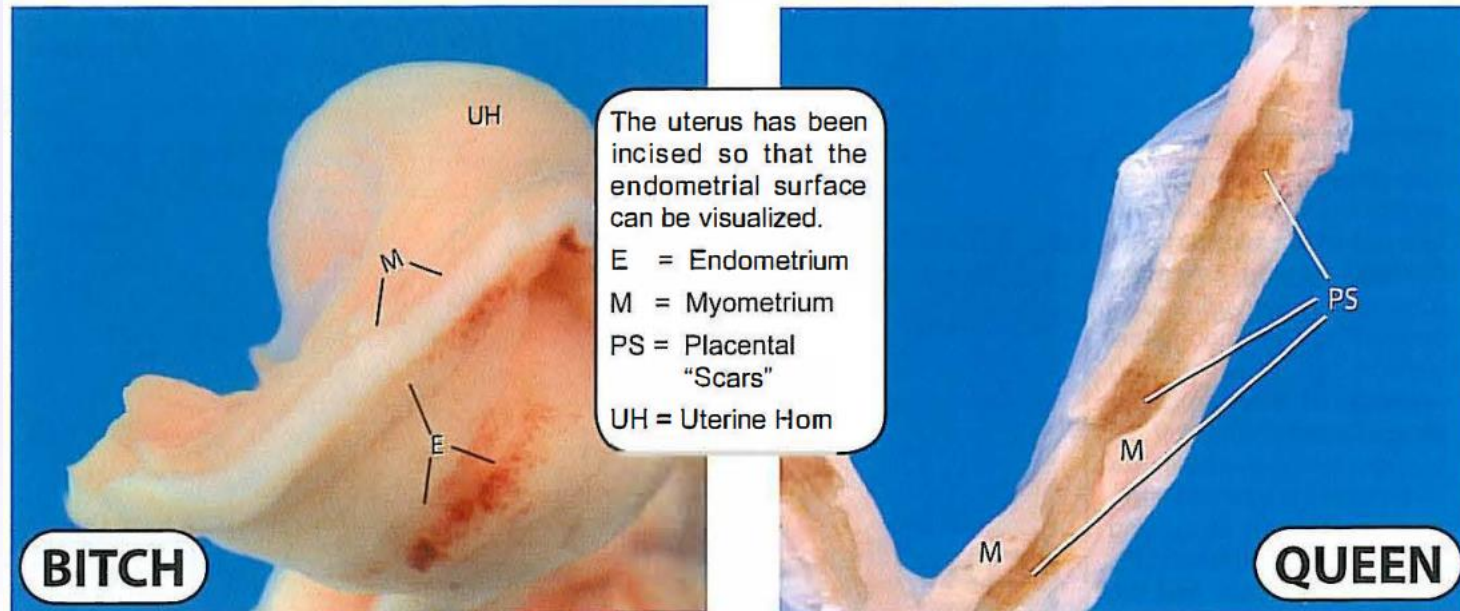
|     |                               |
|-----|-------------------------------|
| C   | = Caruncles                   |
| EF  | = Endometrial Folds           |
| IcE | = Intercaruncular Endometrium |
| M   | = Myometrium                  |
| O   | = Ovary                       |
| UOL | = Utero-Ovarian Ligament      |
| V   | = Blood Vessels               |



# The Uterus

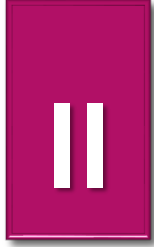


- Placental "scars" in the uterus of the queen represent sites of **previous placental attachment**. These sites are not true scars that are permanent fibrous replacements of normal tissue.
- They are useful to wildlife biologists who use them in postmortem evaluation of wild animals to approximate the number of young produced by a female within a certain period of time.

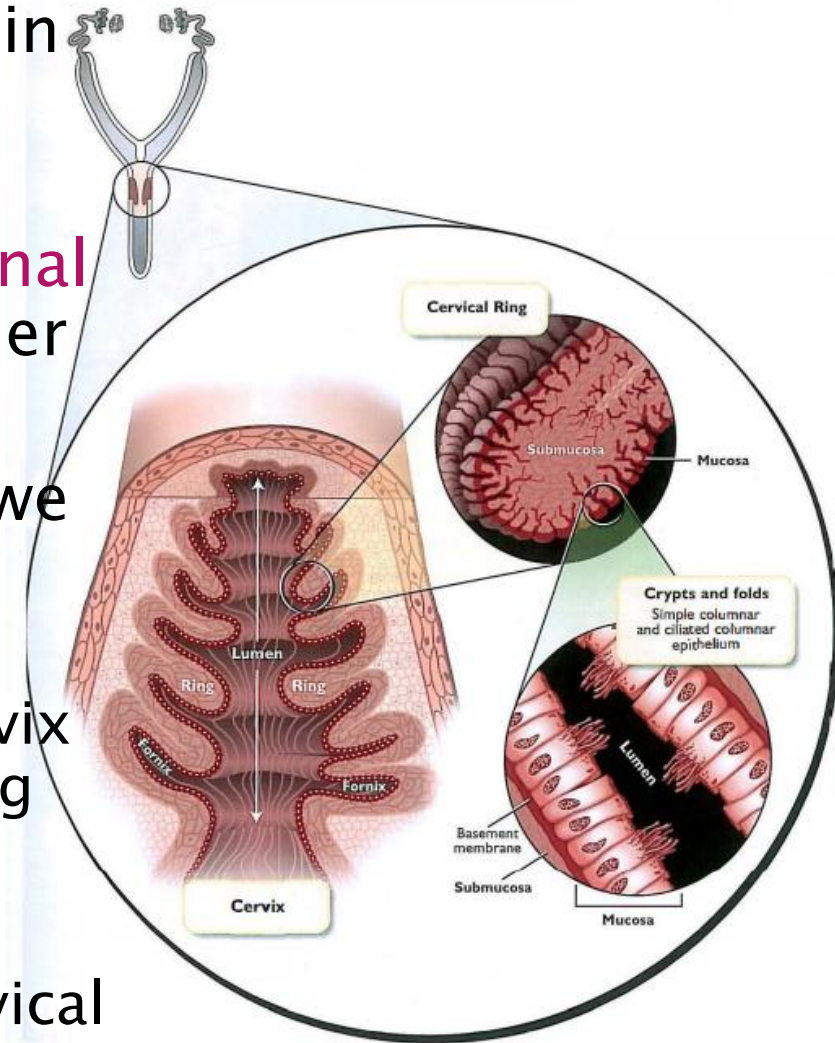




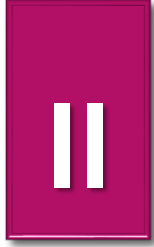
# The Cervix



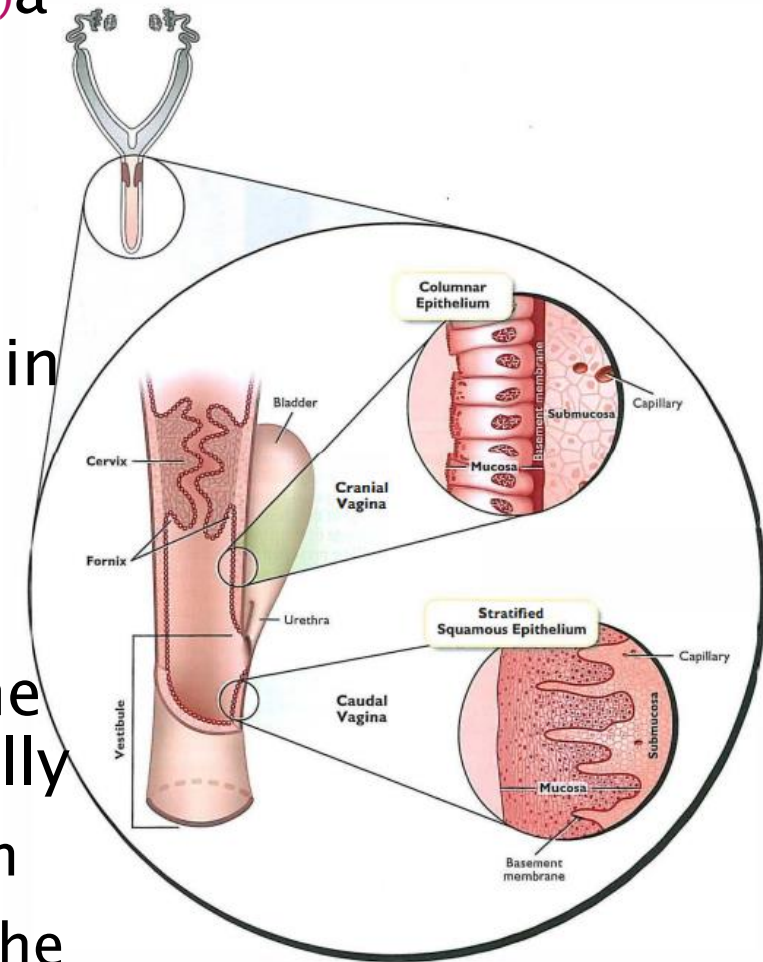
- The **cervix** is a relatively thick-walled, noncompliant organ that serves as a **barrier to sperm** transport in the ewe, cow, bitch and queen but not in the sow and mare.
- The cervix also **isolates the uterus from the external environment** during pregnancy by forming a barrier consisting of highly viscous mucus.
- A primary function of the cervix in the cow and ewe is to **produce mucus during estrus**.
  - In the sow and mare, a much smaller quantity of mucus is produced. This mucus flows from the cervix toward the exterior and lubricates the vagina during copulation.
  - Foreign material introduced during copulation (including sperm) is flushed out of the tract by cervical mucus.



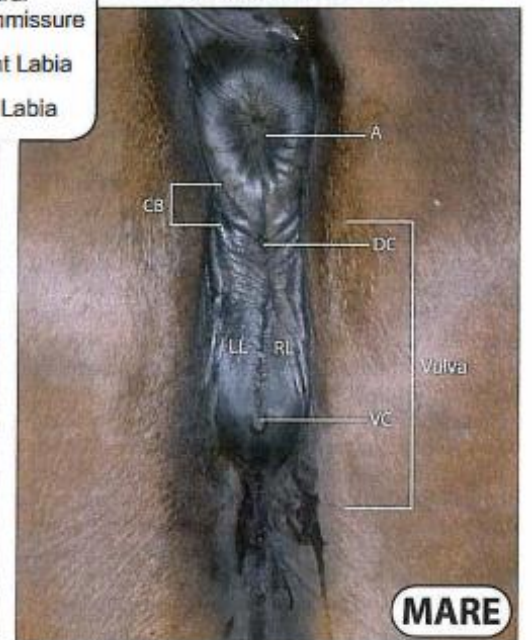
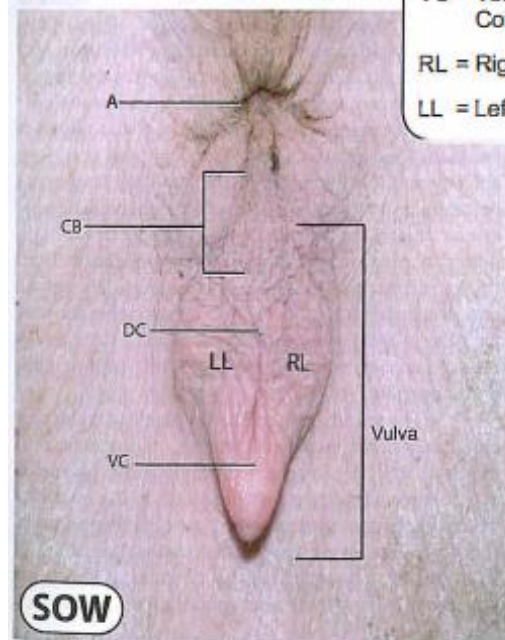
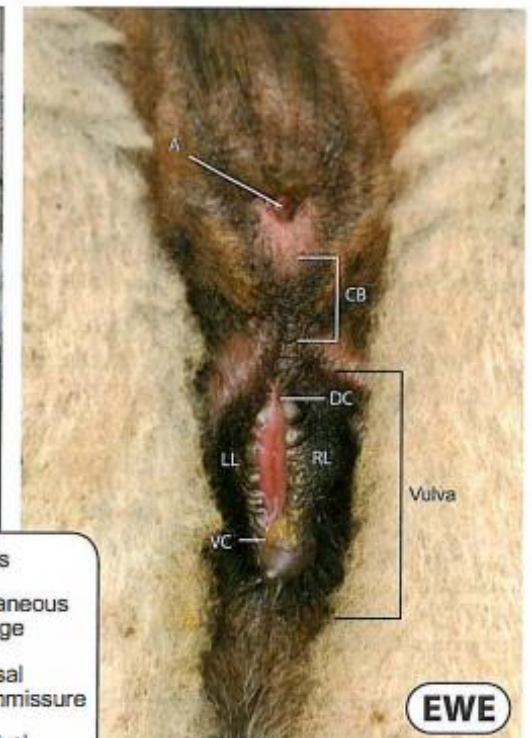
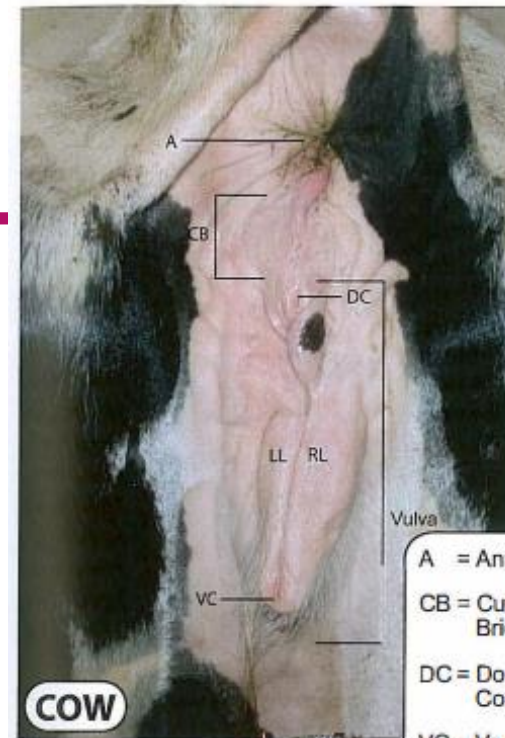
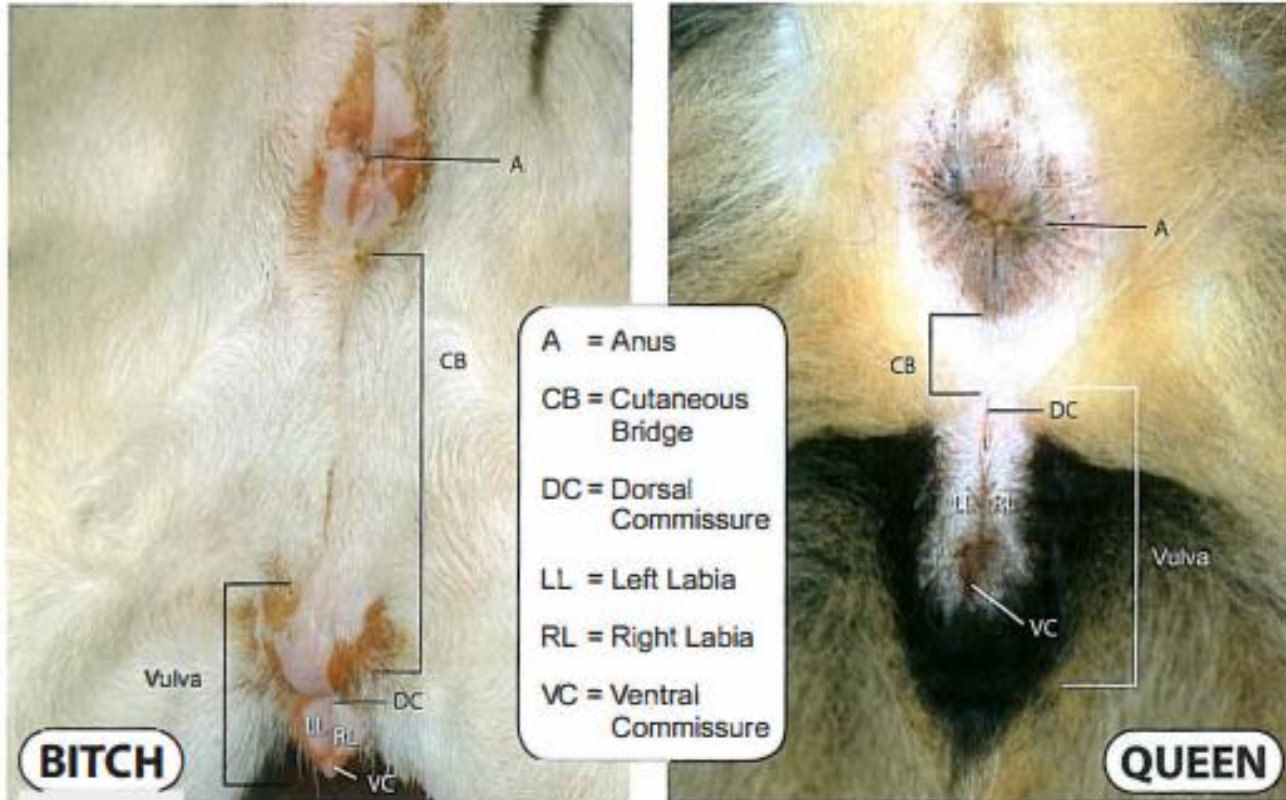
# The Vagina

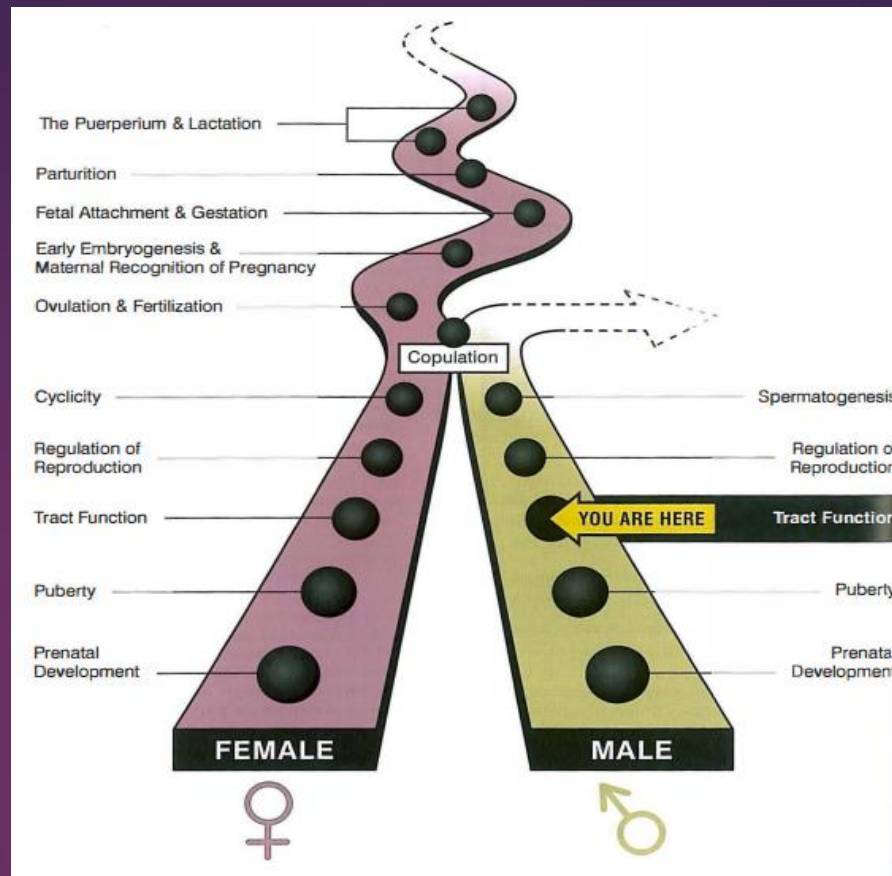


- The primary function of the **vagina** is to serve as: 1) a copulatory organ, 2) the site for expulsion of urine during micturition 3) a passive birth canal during parturition.
- The luminal epithelium near the cervix (cranial vagina) is generally columnar and highly secretory in nature.
- The caudal vagina is characterized as having stratified squamous epithelium
- During the time of estrogen dominance (estrus), the stratified squamous epithelium thickens dramatically
  - it mechanically protects the vagina during copulation
  - prevents microorganisms from gaining entrance to the vasculature in the submucosa.



# The External Genitalia



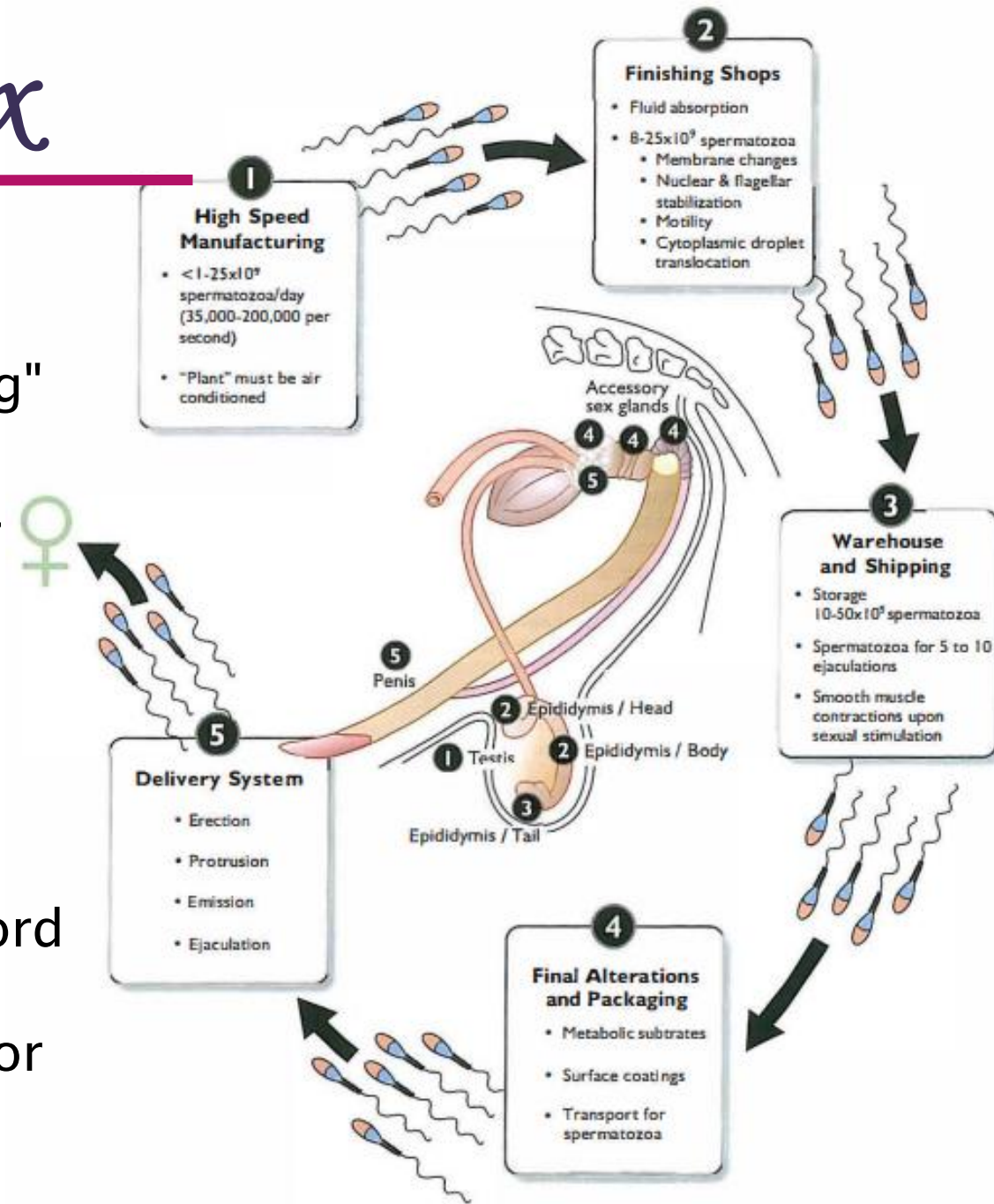


## Section III

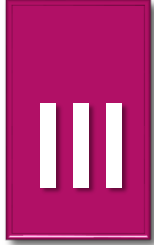
# *The Organization and Function of the Male Reproductive Tract*

# A Manufacturing Complex

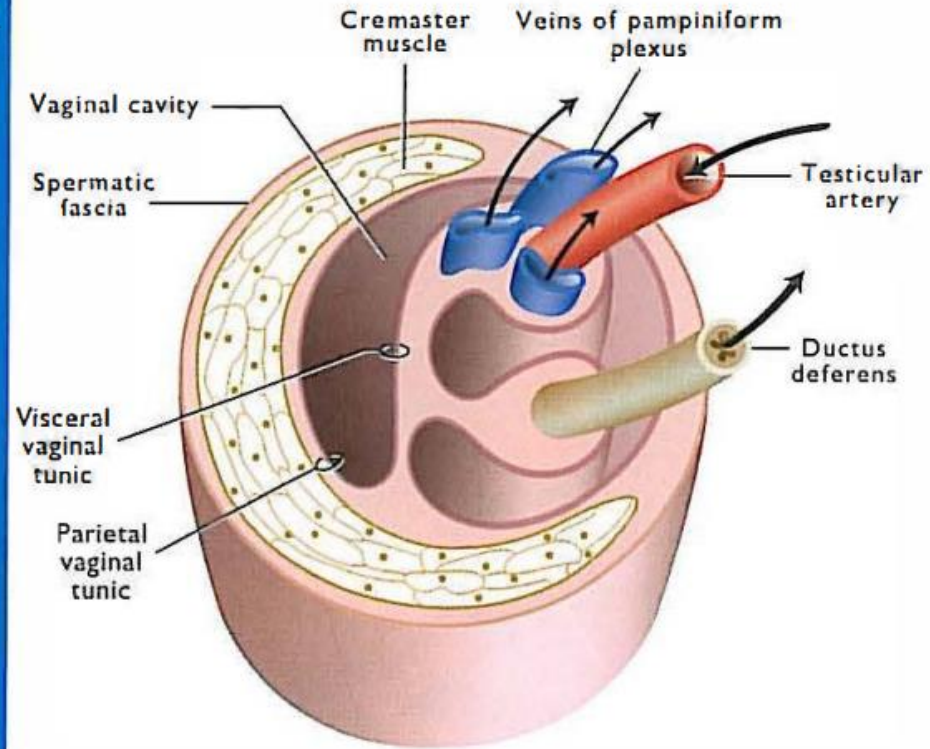
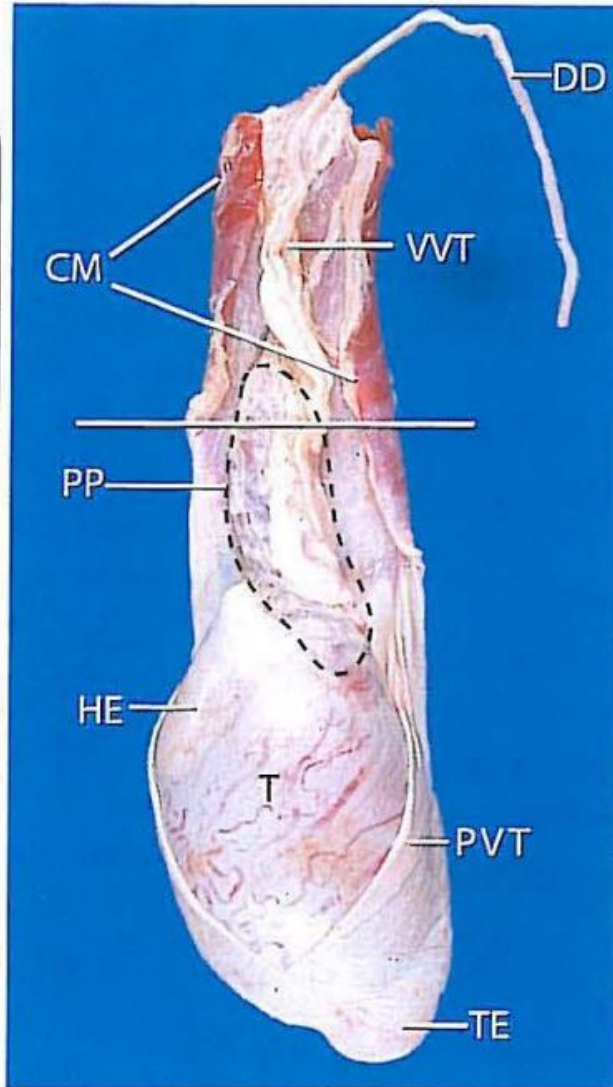
- The male reproductive system is analogous to a **manufacturing complex**.
- The primary products of the "manufacturing" process are fertile **spermatozoa**.
- **Hormones** (such as testosterone) and other secretory products (epididymal fluid and seminal plasma) of the male system contribute to the efficiency of the overall manufacturing and delivery process
- The **basic components of the male reproductive system** are the: 1) spermatic cord 2) scrotum 3) testis 4) excurrent duct system 5) accessory sex glands 6) penis 7) muscles for protrusion, erection and ejaculation



# The Spermatic Cord

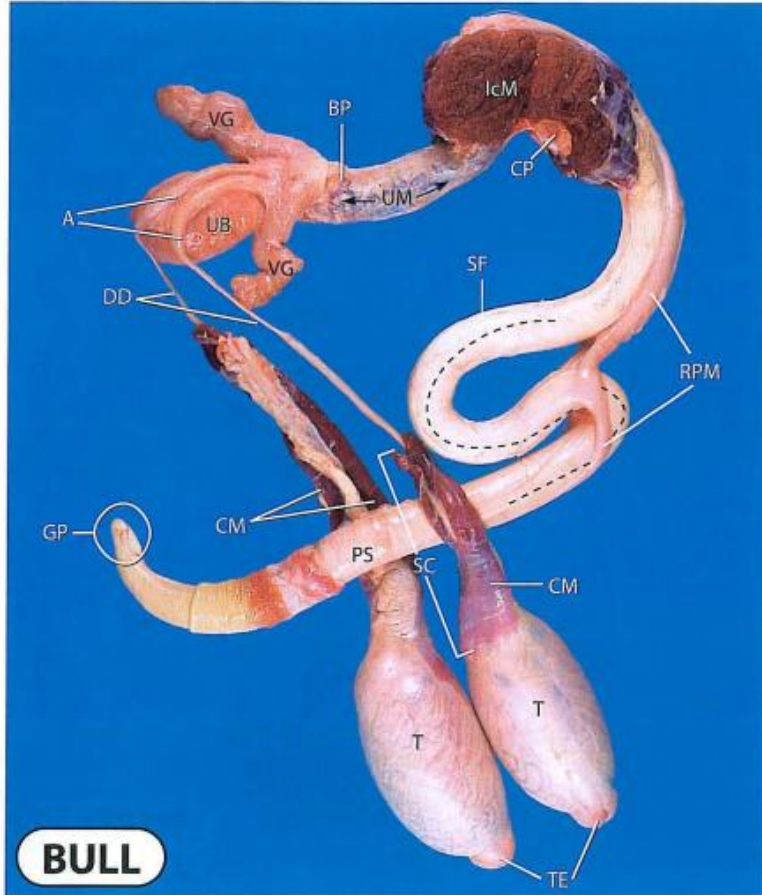
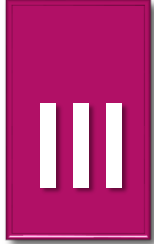


- CM = Cremaster Muscle
- DD = Ductus Deferens
- HE = Head of Epididymis
- PP = Pampiniform Plexus
- PVT = Parietal Vaginal Tunic
- T = Testis
- TE = Tail of the Epididymis
- WT = Visceral Vaginal Tunic



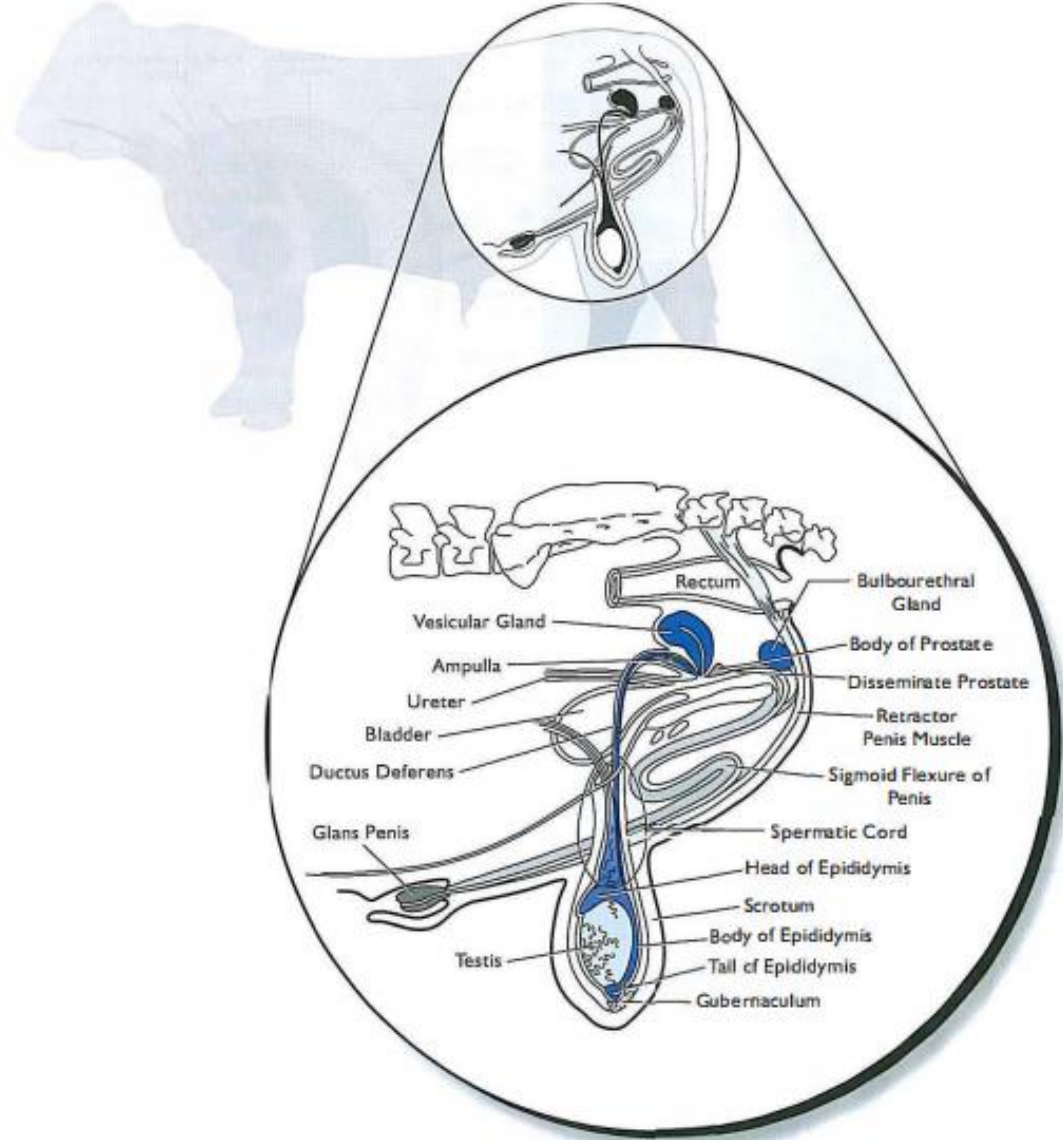
The line across the excised spermatic cord (photo at left) indicates the approximate plane of the cross-sectional schematic. Arrows indicate direction of fluid flow.

# Bull

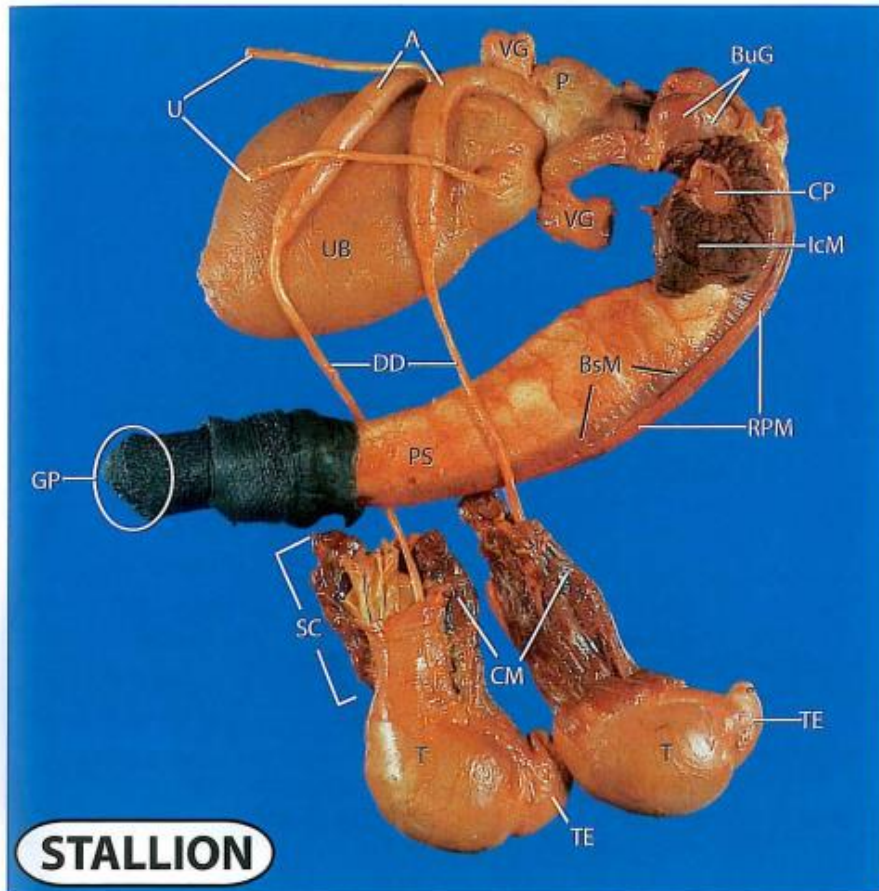


**BULL**

- |                       |                               |                         |
|-----------------------|-------------------------------|-------------------------|
| A = Ampulla           | IcM = Ischiocavernosus Muscle | TE = Tail of Epididymis |
| BP = Body of Prostate | PS = Penile Shaft             | UB = Urinary Bladder    |
| CM = Cremaster Muscle | RPM = Retractor Penis Muscle  | UM = Urethralis Muscle  |
| CP = Crus Penis       | SC = Spermatic Cord           | VG = Vesicular Gland    |
| DD = Ductus Deferens  | SF = Sigmoid Flexure          |                         |
| GP = Glans Penis      | T = Testis                    |                         |

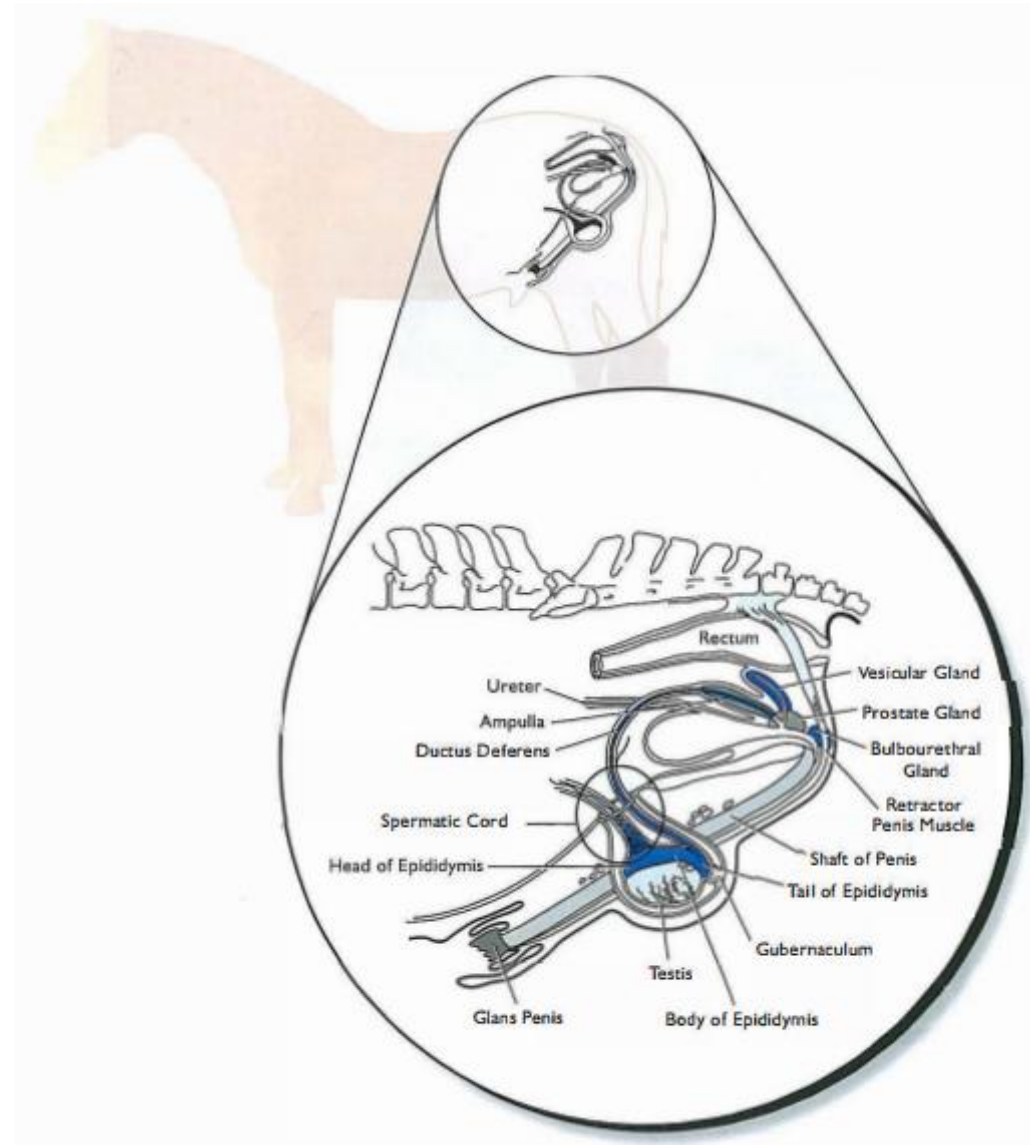


# Stallion



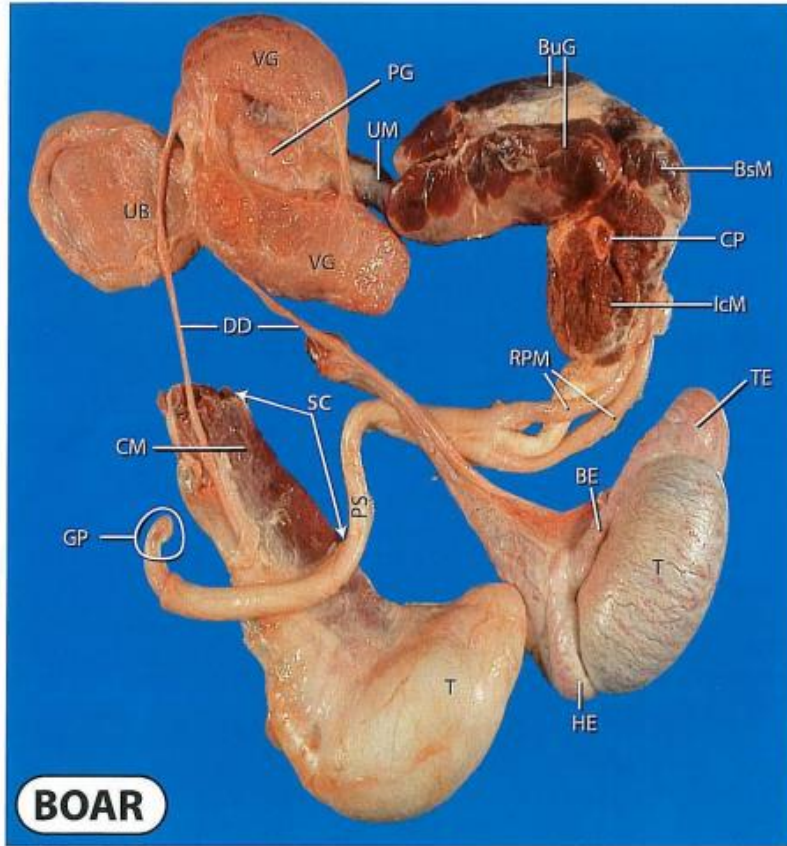
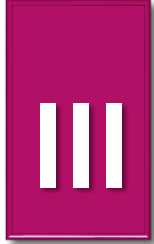
**STALLION**

- |                              |                               |                         |
|------------------------------|-------------------------------|-------------------------|
| A = Ampulla                  | GP = Glans Penis              | T = Testis              |
| BsM = Bulbospongiosus Muscle | IcM = Ischiocavernosus Muscle | TE = Tail of Epididymis |
| BuG = Bulbourethral Gland    | P = Prostate                  | U = Ureters             |
| CM = Cremaster Muscle        | PS = Penile Shaft             | UB = Urinary Bladder    |
| CP = Crus Penis              | RPM = Retractor Penis Muscle  | VG = Vesicular Gland    |
| DD = Ductus Deferens         | SC = Spermatic Cord           |                         |



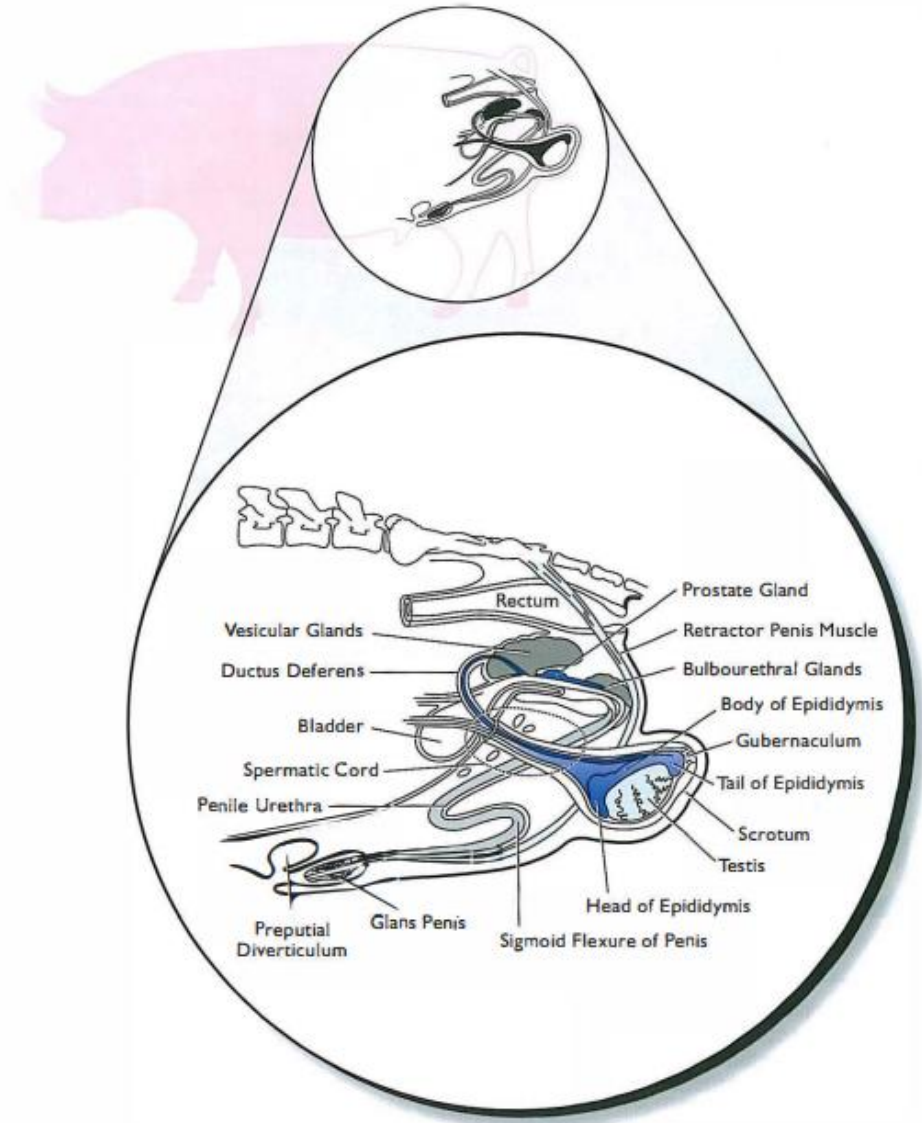


# Boar

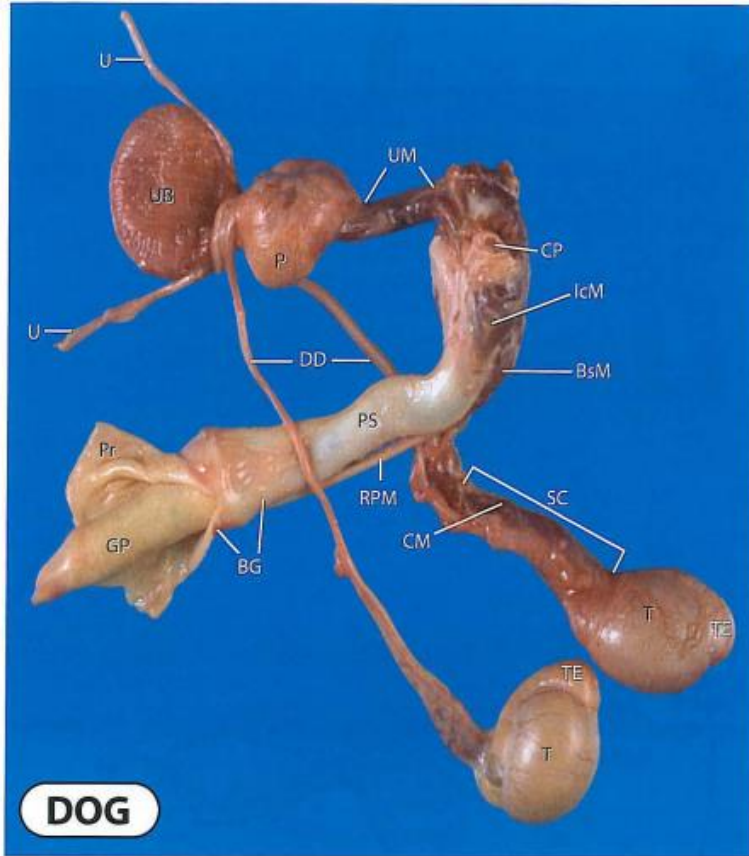
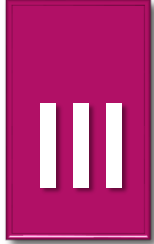


**BOAR**

- |                              |   |                         |
|------------------------------|---|-------------------------|
| BE = Body of Epididymis      | HE = Head of Epididymis   | TE = Tail of Epididymis |
| BsM = Bulbospongiosus Muscle | IcM = Ischiocavernosus Muscle   | UB = Urinary Bladder    |
| BuG = Bulbourethral Gland    | PG = Prostate Gland   | UM = Urethralis Muscle  |
| CM = Cremaster Muscle        | PS = Penile Shaft   | VG = Vesicular Gland    |
| CP = Crus Penis              | RPM = Retractor Penis Muscle  |                         |
| DD = Ductus Deferens         | SC = Spermatic Cord   |                         |
| GP = Glans Penis             | T = Testis (left T-parietal vaginal tunic intact; right T-parietal vaginal tunic removed) |                         |

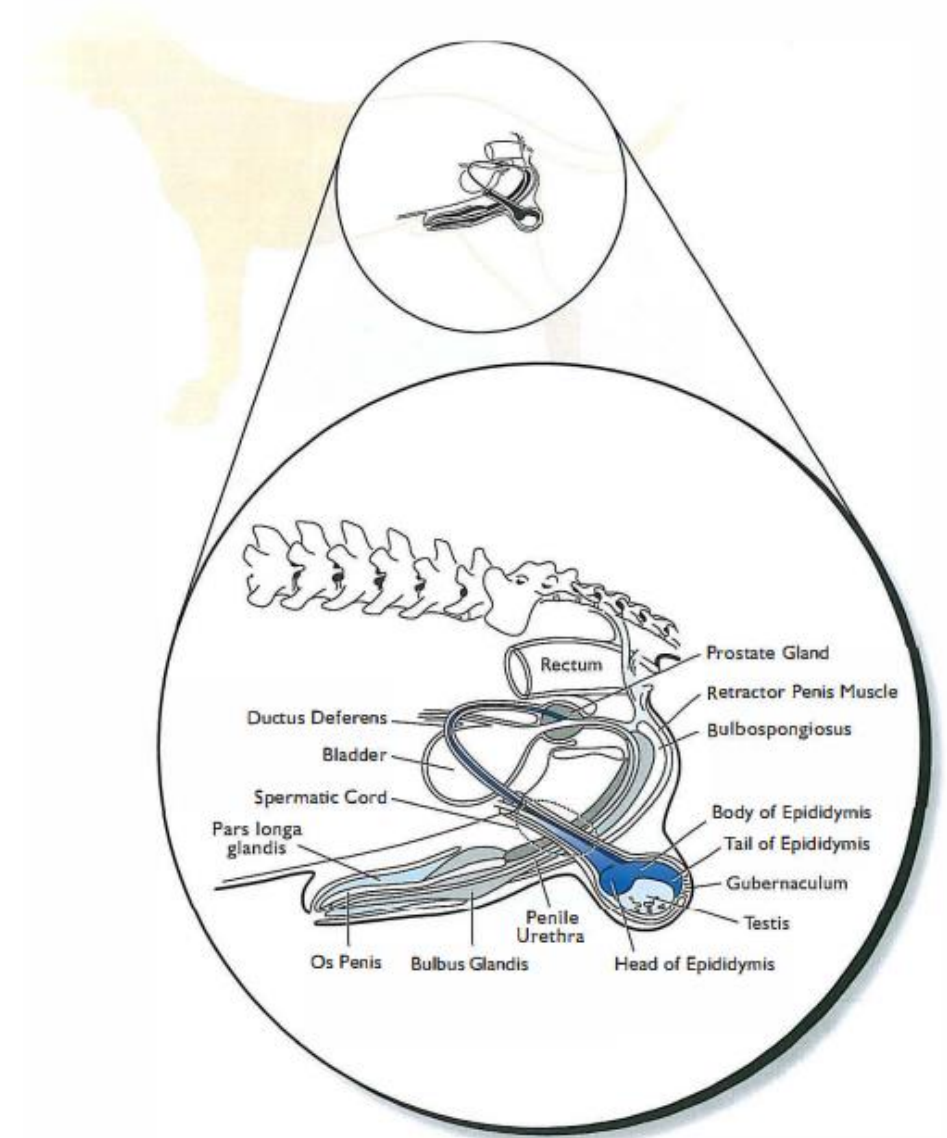


# Dog

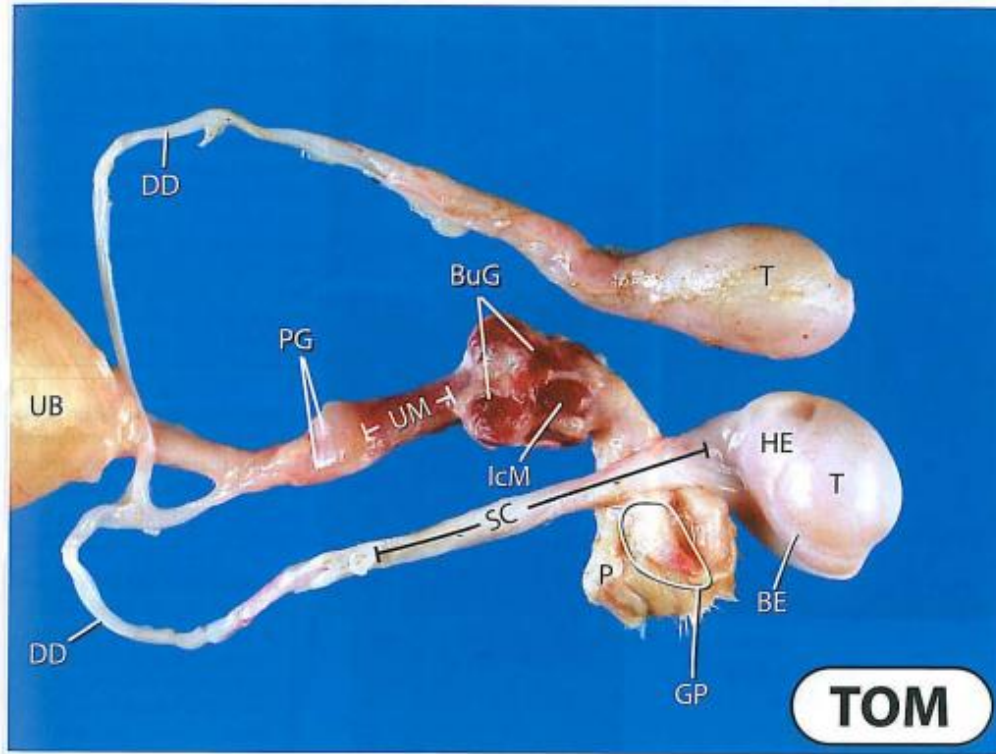
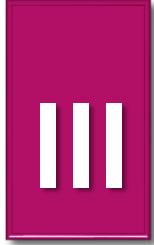


**DOG**

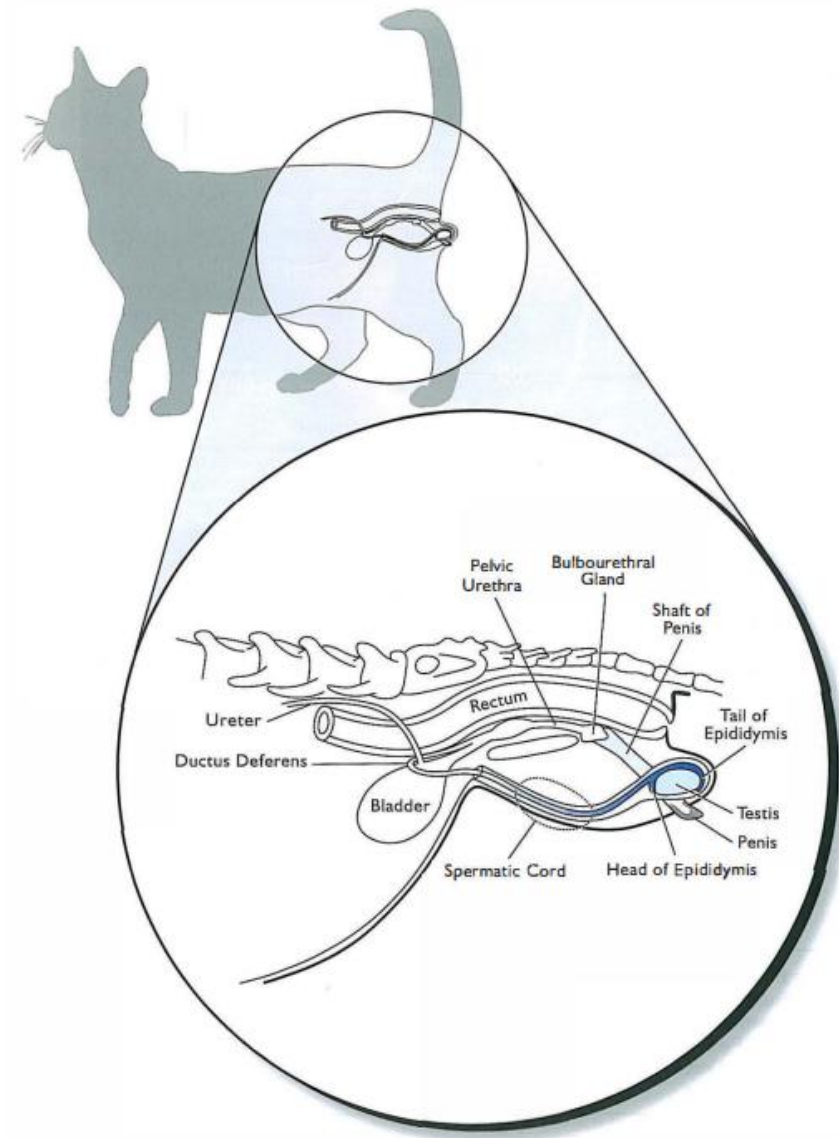
- |                              |                               |                         |
|------------------------------|-------------------------------|-------------------------|
| BG = Bulbus Glandis          | IcM = Ischiocavernosus Muscle | T = Testis              |
| BsM = Bulbospongiosus Muscle | P = Prostate Gland            | TE = Tail of Epididymis |
| CM = Cremaster Muscle        | PS = Penile Shaft             | U = Ureter              |
| CP = Crus Penis              | PR = Prepuce                  | UB = Urinary Bladder    |
| DD = Ductus Deferens         | RPM = Retractor Penis Muscle  | UM = Urethralis Muscle  |
| GP = Glans Penis             | SC = Spermatic Cord           |                         |



# Tom



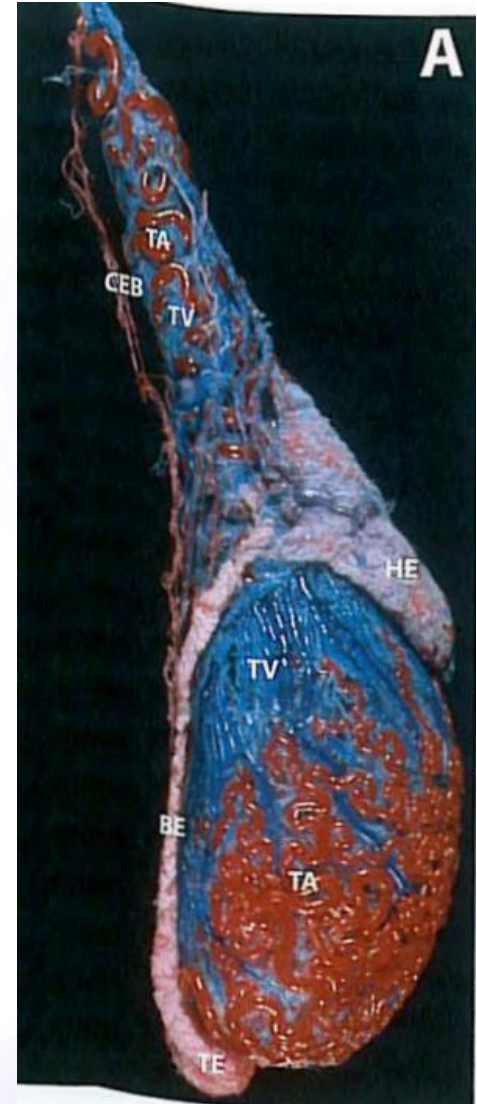
- |                            |                               |                        |
|----------------------------|-------------------------------|------------------------|
| BE = Body of Epididymis    | HE = Head of Epididymis       | SC = Spermatic Cord    |
| BuG = Bulbourethral Glands | IcM = Ischiocavernosus Muscle | T = Testis             |
| DD = Ductus Deferens       | P = Prepuce                   | U = Urinary Bladder    |
| GP = Glans Penis           | PG = Prostate Gland           | UM = Urethralis Muscle |



# *The pampiniform plexus*



- The **testicular artery** (TA) is highly convoluted and passes through the spermatic cord and surrounds the testis in the ventromedial area.
- In the spermatic cord, the **testicular veins** (TV) are in close proximity to the torturous testicular artery. The testicular veins (TV) seen on the surface of the testicle return venous blood to the spermatic cord.
  - A branch of the testicular artery, the caudal epididymal branch (CEB) can be observed. The head of the epididymis (HE), body of the epididymis (BE) and tail of the epididymis (TE) can be seen.



# *The pampiniform plexus*



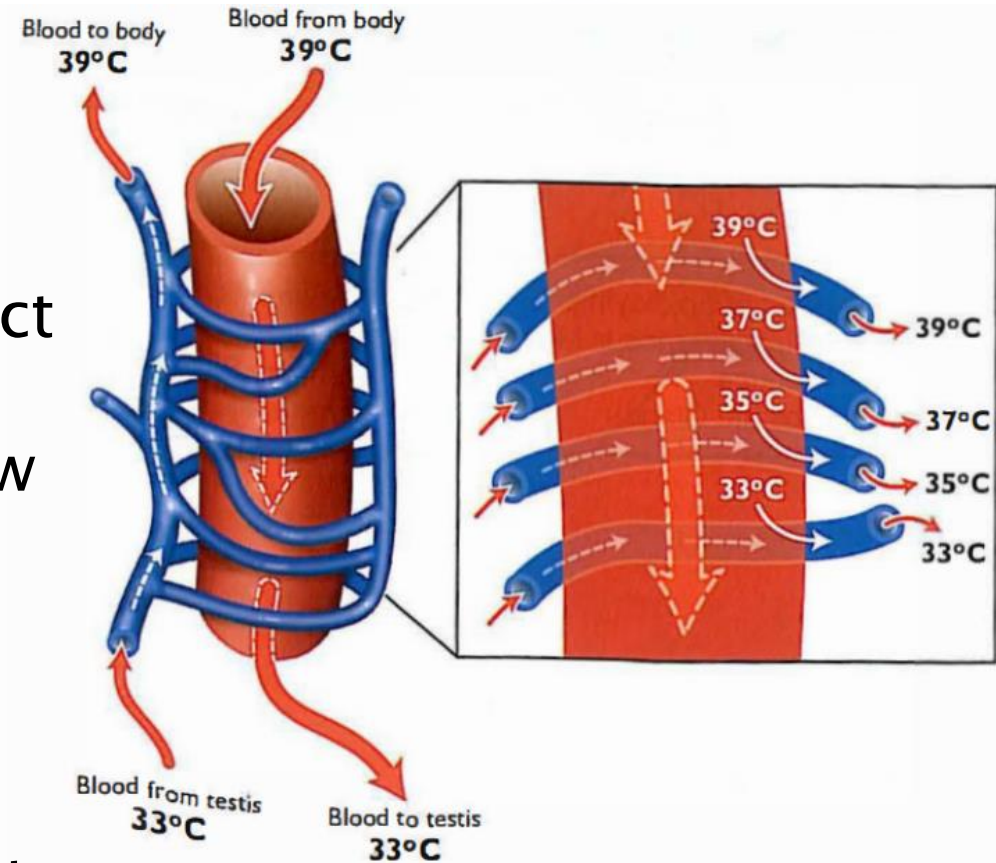
- An enlarged view of a portion of the vascular cone.
- The **highly convoluted testicular artery** (TA) has an intimate relationship with the **veins** of the pampiniform plexus (PP).
- A highly enlarged photograph showing the intimate relationship of the pampiniform plexus with the testicular artery (TA).
- Notice the finger-like "wrappings" (arrows) of the pampiniform plexus surrounding the testicular artery (TA). This intimate relationship provides the **anatomical basis for the countercurrent heat exchanger**.



# The pampiniform plexus



- Heat from the warm (39°C) arterial blood from the body is transferred to the cooler (33°C) venous blood leaving the surface of the testes.
- This venous blood has been cooled by direct heat loss from the testicular veins through the skin of the scrotum. Maintenance of low testicular temperature is obligatory for spermatogenesis in domestic animals and man.
- Disruption or modification of this cooling mechanism will severely compromise, if not completely suppress, spermatogenesis.



# The Testes

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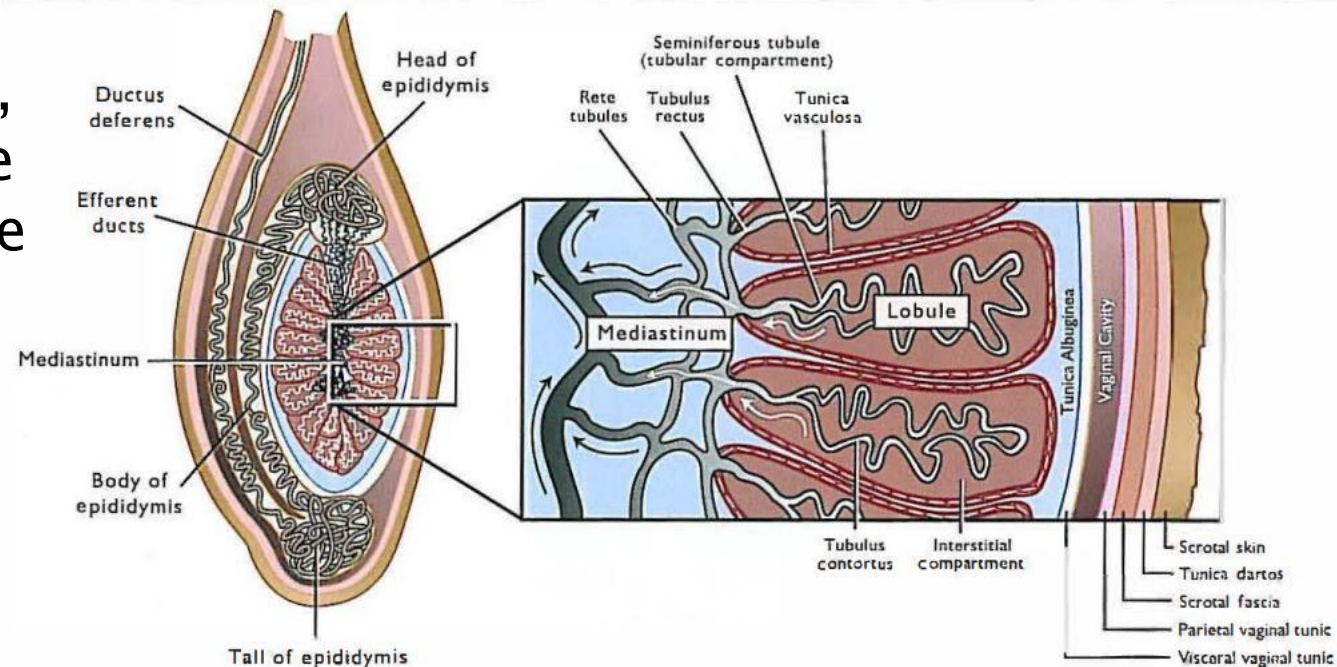


- The **testes** are paired organs that vary considerably in size and shape among species.
- They are considered the primary reproductive organs in the male because they produce both **spermatozoa** and the androgen **testosterone**.
- In addition, they produce **inhibin**, **estrogens** and a variety of **proteins** believed to be important to spermatozoal function.
- They also produce **fluid** that originates primarily from the seminiferous tubules. This fluid serves as a vehicle in which spermatozoa are suspended and facilitates their removal from the testes. The fluid produced by the testes (sometimes called rete fluid) also contains products synthesized by the Sertoli cells.

# The Testes



- The word **parenchyma** refers to the specific cellular mass of a gland or organ that is supported by a connective tissue network
  - **Tubular** parenchyma: **seminiferous tubules** and all of the cells and material inside them.
  - **Interstitial** parenchyma: consists of all cells and materials outside the seminiferous tubules, such as blood vessels, connective tissue, lymphatics, nerves and the interstitial cells of **Leydig**, that produce **testosterone**.

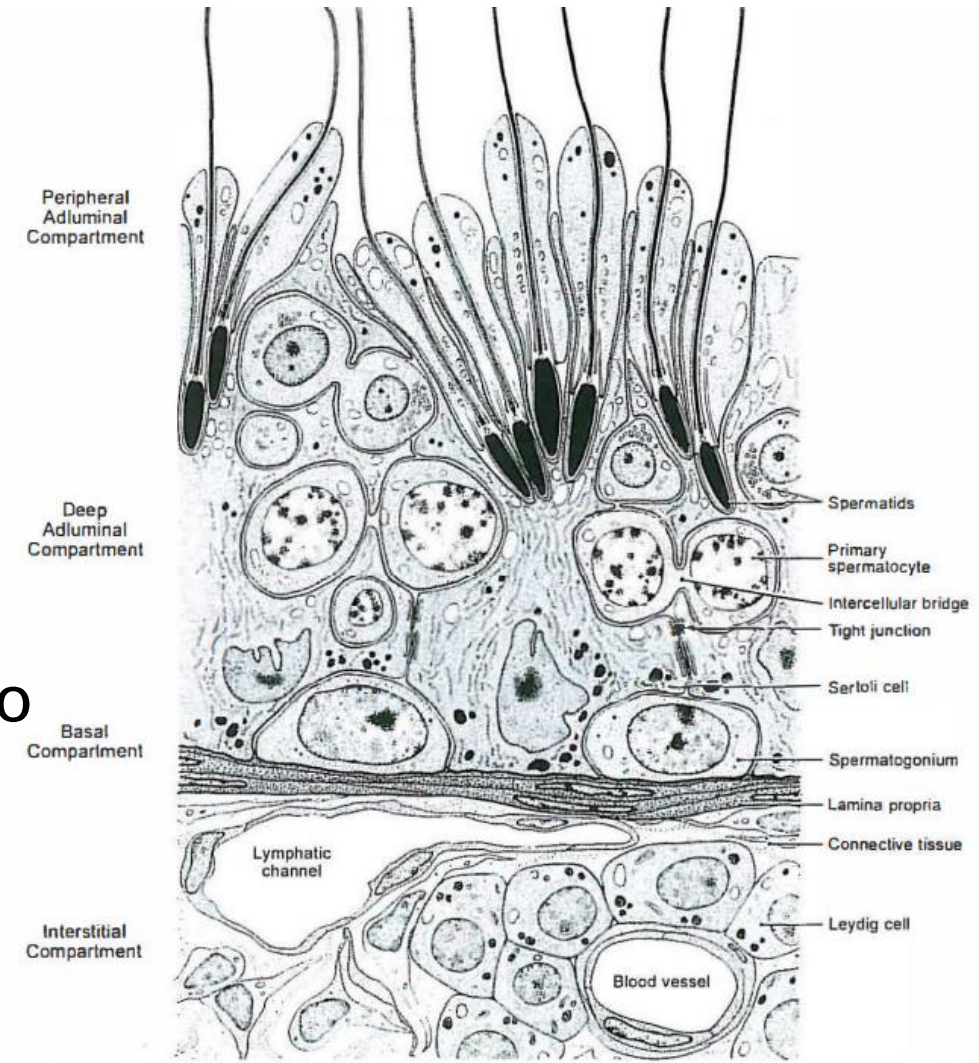




# The Testes



- The **seminiferous tubule** is composed of a basement membrane and a layer of seminiferous epithelium (also called the **germinal epithelium**). The tubule is surrounded by contractile **peritubular cells**. Their contraction and the flow of fluid secreted by **Sertoli cells** allows newly formed spermatozoa to move into the rete tubules .
- The seminiferous epithelium consists of two major regions known as the **basal compartment** and the **adluminal compartment**. Sertoli cells are anchored to the basement membrane and surround the developing population of germ cells



# Seminiferous tubule



## Peripheral Adluminal Compartment

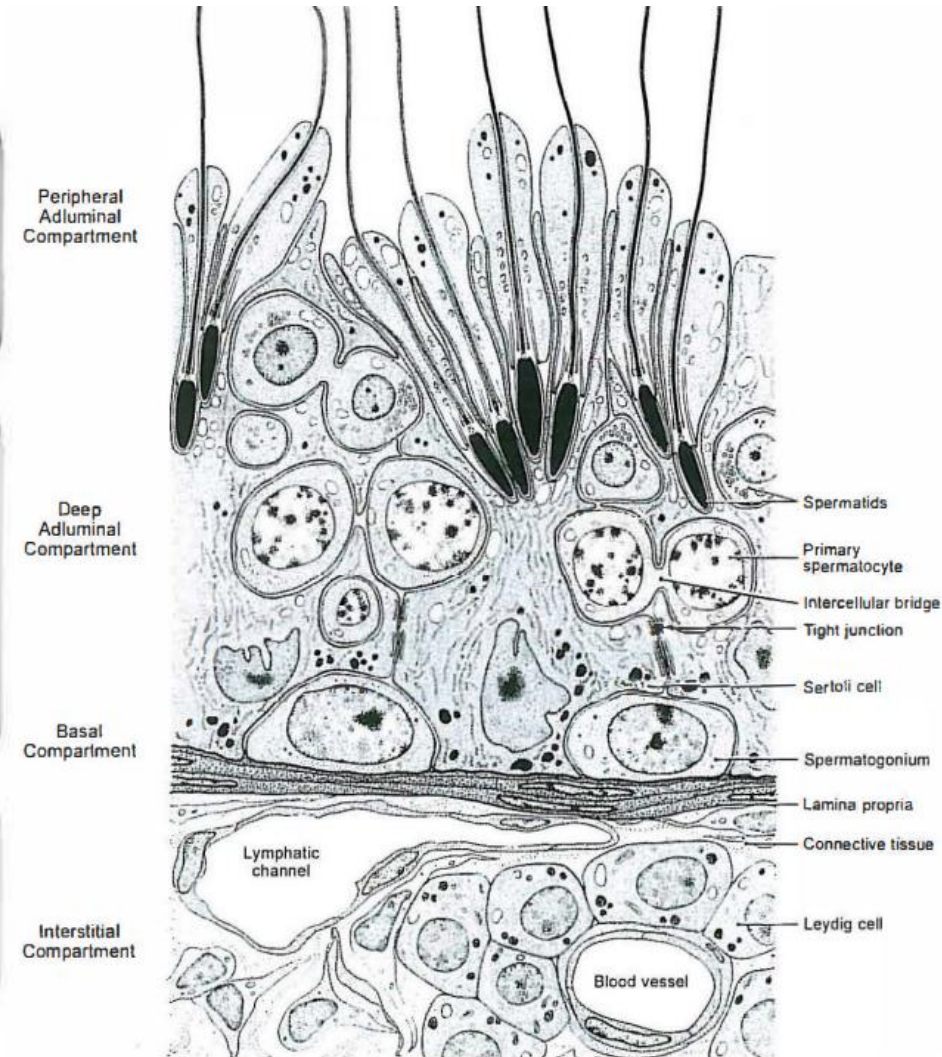
During elongation of the spermatid nucleus, the spermatids are repositioned by the Sertoli cells to become imbedded within long pockets in the cytoplasm of an individual Sertoli cell. When released as a spermatozoon, a major portion of the cytoplasm of each spermatid remains as a residual body (cytoplasmic droplet) within a pocket of the Sertoli cell cytoplasm.

## Deep Adluminal Compartment

The primary spermatocytes are moved from the basal compartment through the tight junctions between adjacent Sertoli cells into the adluminal compartment where they eventually divide to form secondary spermatocytes (not shown) and spherical spermatids. The spermatogonia, primary spermatocytes, secondary spermatocytes and spherical spermatids all develop in the space between two or more Sertoli cells and are in contact with them. Note the intracellular bridges between adjacent germ cells in the same cohort or generation.

## Basal Compartment

Formation of spermatozoa in the seminiferous epithelium starts near the basement membrane. Here a spermatogonium divides to form other spermatogonia and, ultimately, primary spermatocytes. (From Amann, *J.Dairy Sci.* Vol. 66, No. 12, 1983)



# *Seminiferous tubule*

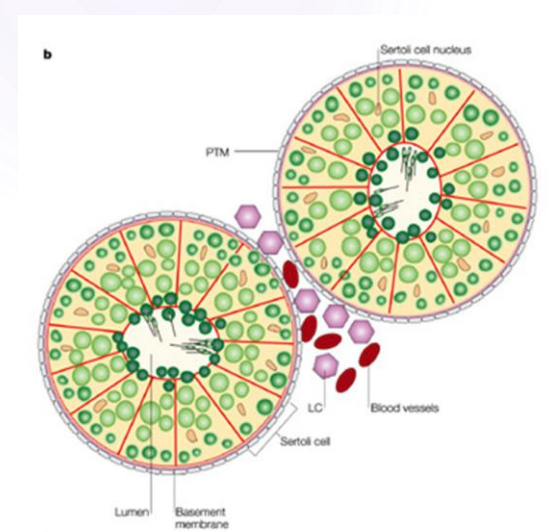
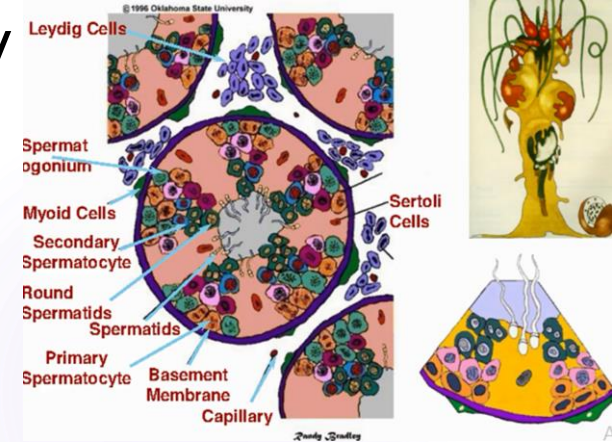
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- Because **Sertoli cells** possess receptors to different hormones (protein and steroid), they have the capability of producing a variety of substances
  - **Androgen binding protein (ABP)**, a testosterone transport protein;
  - **Sulfated glycoproteins (SGP) 1 and 2**,
    - fertility acquisition (**SGP-1**)
    - providing a detergent effect that allows cells and fluids to move through the tubular network of the testis (**SGP-2**);
  - **Transferrin**, an iron transport protein believed to be required for successful spermatogenesis
  - **Inhibin**, as in the female, a suppressor of FSH.

# Blood-Testis Barrier

- Adjacent Sertoli cells are tightly attached to each other by **tight junctions**. The **peritubular cells** surrounding the seminiferous tubule and the **Sertoli cell junctional complexes** form the blood-testis barrier (BTB).
  - The primary purpose is to **prevent autoimmune reactions** from destroying the developing germ cells. The peritubular layer **exclude the immune cells** (macrophages and lymphocytes) and immunoglobulins (antibodies) from the adluminal compartment. these molecules would recognize the developing germinal elements as foreign because they are undergoing meiosis so they generate **immunologic response**.
  - In addition to forming the blood-testis barrier, the Sertoli cell junctional complexes provide a type of **control for transportation of materials** i.e. entering and, at least in part, leaving the adluminal compartment.



# Epididymis

- The function of the epididymis is to provide the environment for final **maturation** of spermatozoa, resulting in acquisition of **motility** and potential **fertility**.
- The epididymis also serves as a storage **reservoir** for spermatozoa. Epididymal function is **androgen dependent**.

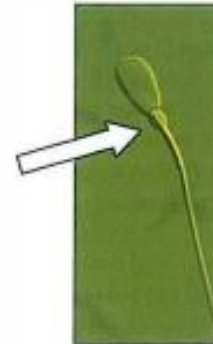
**Head (Caput)**  
25-50 x 10<sup>6</sup> sperm



The head of the epididymis is subdivided into the proximal head (PH) and the distal head (DH). The proximal head reabsorbs a significant amount of rete fluid while the distal head secretes fluid into the lumen of the epididymal duct. Thus, concentration of sperm within the head of the epididymis increases and then decreases significantly.

## Spermatozoal Characteristic

- Not motile
- Not fertile
- Proximal cytoplasmic droplet
- Low disulfide crosslinking

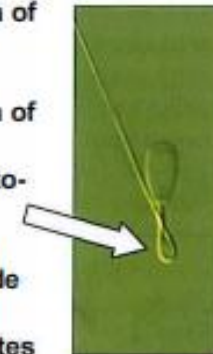


**Body (Corpus)**  
8-25 x 10<sup>9</sup> sperm

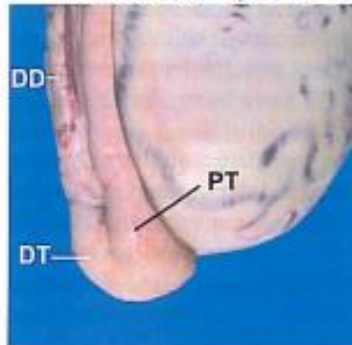


The body of the epididymis (BE) lies parallel to the ductus deferens (DD). Concentrations of sperm throughout the body of the epididymis remain relatively constant.

- Some expression of motility after dilution
- Some expression of fertility
- Translocating cytoplasmic droplet
- Moderate to high degree of disulfide crosslinking
- Can bind to oocytes

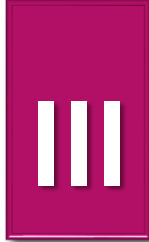
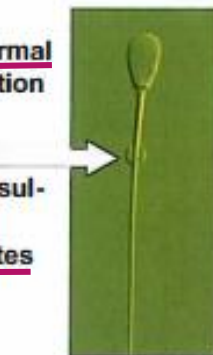


**Tail (Cauda)**  
10-50 x 10<sup>9</sup> sperm



The tail of the epididymis consists of the proximal tail (PT) and the distal tail (DT). Sperm within the distal tail are eligible for ejaculation. Sperm in the proximal tail cannot be moved into an ejaculatory position following sexual stimulation. However, the sperm in the distal tail move through the ductus deferens (DD) and into the pelvic urethra during sexual stimulation.

- Expression of normal motility after dilution
- Fertile potential
- Distal droplet
- High degree of disulfide crosslinking
- Can bind to oocytes



# Accessory Sex Glands

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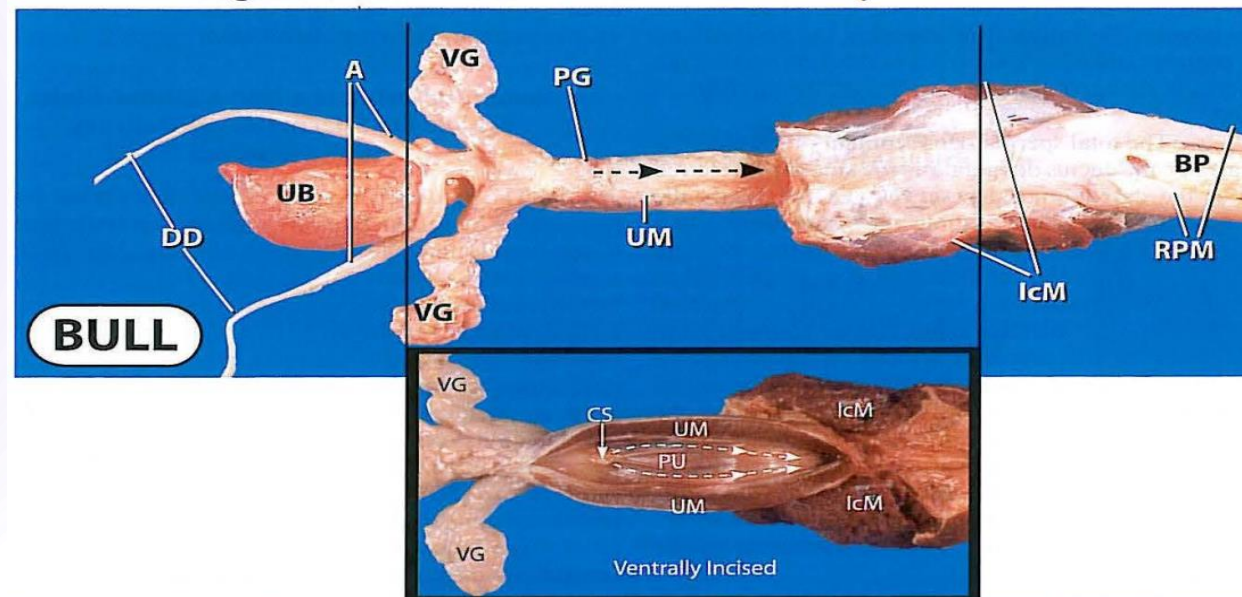


- The epididymis and accessory sex glands are responsible for production of secretions that contribute to the liquid, noncellular portion of semen known as **seminal plasma**.
- Seminal plasma is not required for fertility, but is important in **natural insemination** where a fluid vehicle for delivery of the sperm is needed.
- Spermatozoa that are removed from the **tail of the epididymis** are equally as **fertile** as those that are ejaculated.
- In some species (the boar and stallion), the seminal plasma possesses **special coagulation properties** that plug the female reproductive tract and minimize loss of spermatozoa following copulation and ejaculation.
- The accessory sex glands secrete their products into the lumen of the **pelvic urethra**.

# Accessory Sex Glands



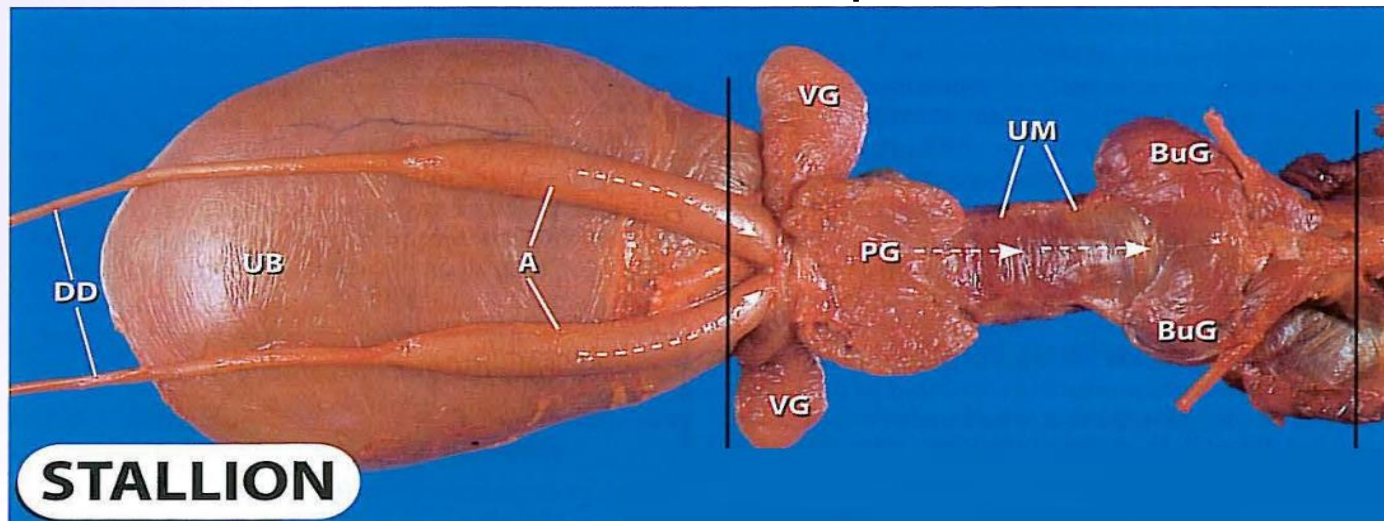
- The **ampullae** are enlargements of the ductus deferens that open directly into the pelvic urethra. The enlargement is the result of a dramatic increase in the mucosa within the ampulla.
- The **vesicular glands** ( ex. seminal vesicles) are paired glands that are dorsocranial to the pelvic urethra. In bulls and boars the vesicular gland contributes to a large proportion of the ejaculate volume.



# Accessory Sex Glands



- The **prostate gland** lies in close proximity to the junction between the bladder and pelvic urethra.
- The prostate may have two structural forms:
  - **Corpus prostate** in which the prostate is outside of the urethralis muscle and is visible as a heart-shaped (boar), or an H-shaped (stallion) structure.
  - **Disseminate prostate** (Urethral gland) in which glandular tissue is distributed along the dorsal and lateral walls of the pelvic urethra.

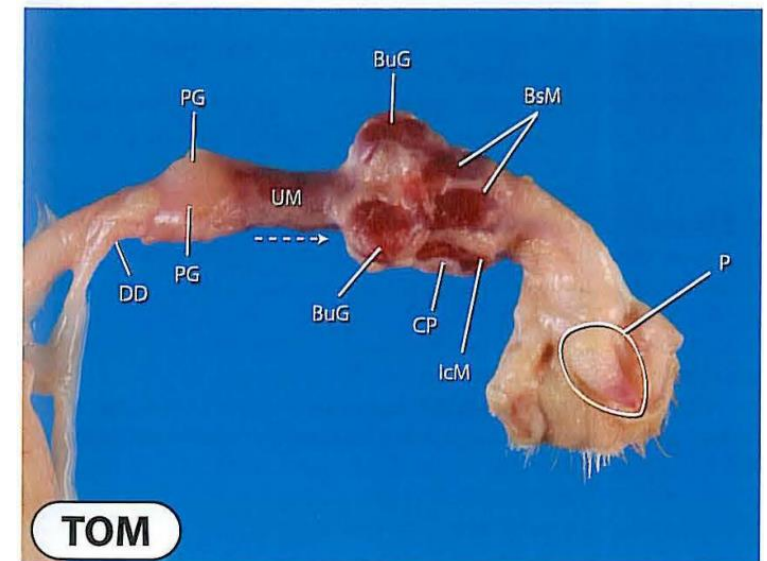
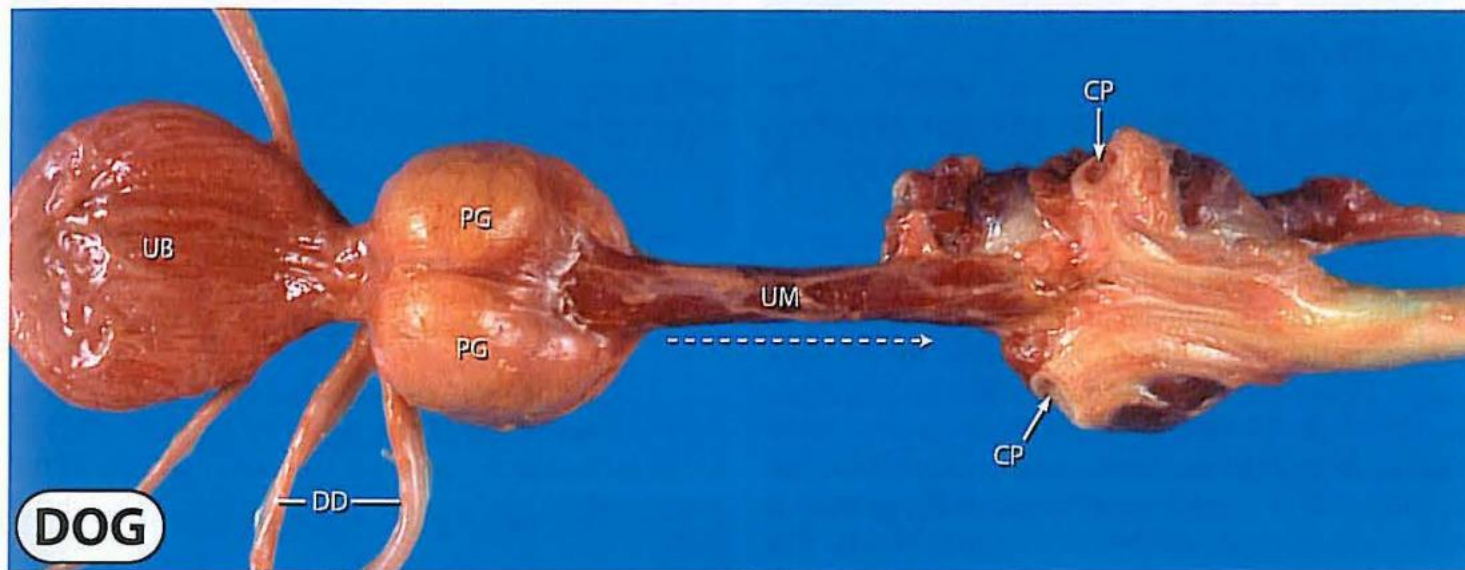




# Accessory Sex Glands



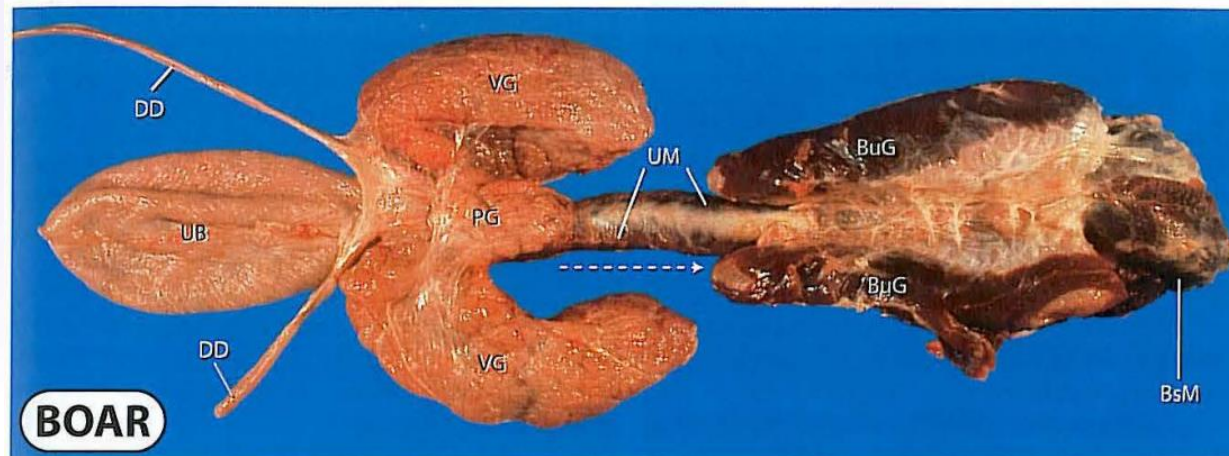
- The **bulbourethral glands** are paired glands located on either side of the pelvic urethra near the ischial arch. These glands are usually small and ovoid shape.
  - These glands produce a viscous secretion that is important because it provides the gel fraction of the ejaculate and causes the seminal plasma to coagulate following ejaculation.



# Accessory Sex Glands



- Secretions of the accessory sex glands contain an immense variety of components and ions, most of which have not been assigned a function. In general, most substances found in blood, including **hormones** and **enzymes**, can be found in seminal plasma.
  - It should be emphasized that with the exception of **fructose** as an energy source, the precise role of the other materials is not known.
- The accessory sex glands are dependent on testosterone for full development and maintenance of their structure and function.



# Journal Club (for more reading)



RESEARCH ARTICLE | VOLUME 42, ISSUE 1, P24-28, JULY 2002 [Download Full Issue](#)

Seminal Secretory Capacity of the Male Accessory Sex Glands in Chronic Pelvic Pain Syndrome (CPPS)/Chronic Prostatitis with Special Focus on the New Prostatitis Classification

Martin Ludwig • Andreas Vidal • Thorsten Diemer • Wolfgang Pabst • Klaus Failing • Wolfgang Weidner

DOI: [https://doi.org/10.1016/S0302-2838\(02\)00224-5](https://doi.org/10.1016/S0302-2838(02)00224-5)

## Abstract

**Objective:** The aim of the study was to evaluate the secretory dysfunction of the male accessory glands in men with inflammatory versus non-inflammatory chronic pelvic pain syndrome (CPPS).

**Methods:** One hundred and twelve consecutive patients symptomatic for chronic pelvic pain were included into the study. All underwent a combined granulocyte analysis in expressed prostatic secretions (EPS) and a four-glass-test followed by ejaculate analysis. Patients were subgrouped according to elevated granulocyte counts in prostatic secretions, leukocytes in semen, or any of both. The content/total enzyme activity of the secretory seminal plasma parameters  $\gamma$ -glutamyl-transferase ( $\gamma$ -GT), fructose, and  $\alpha$ -glucosidase representing the secretory capacity of the prostate gland, the seminal vesicles, and the epididymes, respectively, were investigated.

**Results:** The only significant findings were a reduced total enzyme activity of  $\gamma$ -GT in men stratified according to elevated granulocyte counts in prostatic secretions ( $p=0.022$ ; cutpoint 9.85 U per ejaculate; sensitivity 61.1%, specificity 58.8%, AUC 0.6347) and in men with any inflammatory sign ( $p=0.033$ ; cutpoint 9.9 U per ejaculate, sensitivity 63%, specificity 58.33%, AUC 0.6404).

**Conclusions:** Secretory damage of the prostate gland in men with inflammatory CPPS is demonstrable provided that increased granulocytes in prostatic secretions are part of the diagnostic criteria. However, because of the low sensitivity and specificity of  $\gamma$ -GT it cannot be recommended as diagnostic tool to detect inflammatory disease on the basis of reduced secretory capacity.

► [https://www.europeanurology.com/article/S0302-2838\(02\)00224-5/pdf](https://www.europeanurology.com/article/S0302-2838(02)00224-5/pdf)

# Penis

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- The penis is the **copulatory organ** which compose three parts:
  - base (root) of the penis
  - shaft (the main portion of the penis)
  - glans penis (the specialized distal end)
    - The glans penis is heavily populated with sensory nerves and is the homologue of the female clitoris.
- Bulls, boars and rams have a **fibroelastic penis** with limited erectile tissue encased in a non-expandable, dense connective tissue structure (tuica albuginea).
- In species with a fibroelastic penis, there is a **sigmoid flexure**, an S-shaped configuration along the shaft of the penis.

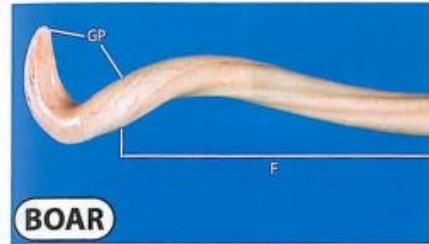
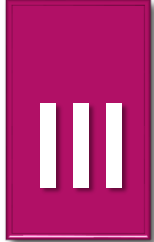
# Penis

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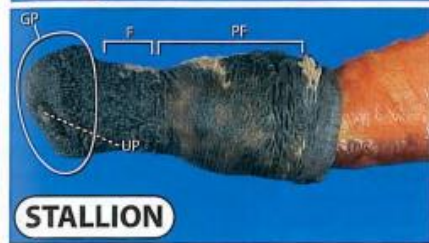


- The shaft of the penis has an area of spongy, erectile tissue known as the **corpus cavernosum** that makes up the majority of the penile interior.
- In the ventral portion of the penis immediately surrounding the penile urethra is another area of spongy erectile tissue called the **corpus spongiosum**
- The cavernous tissue in the dog consists of two morphologically distinct regions.
  - These are **the bulbus glandis** and the **pars longa glandis**. The bulbus glandis forms a turgid bulb during erection that allows the "**copulatory lock**" during the final stages of copulation.
  - The dog penis also has an **os penis** (baculum) that runs through the bulbus glandis and the pars longa glandis.

# Penis

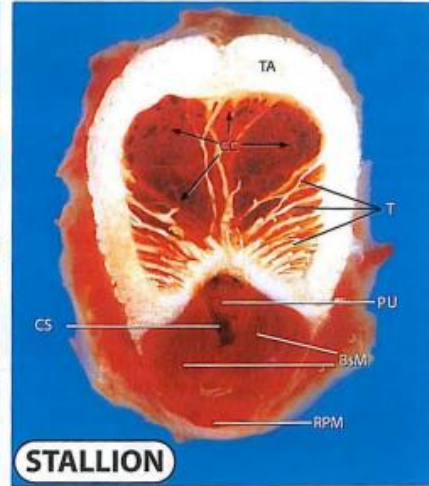


**BOAR**



**STALLION**

- F = Free end of penis
- GP = Glans Penis
- PF = Preputial Fold
- UP = Urethral Process



**STALLION**

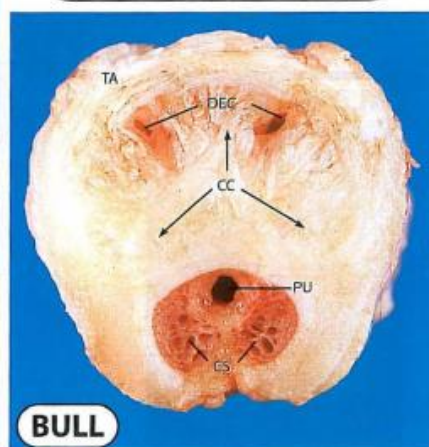
- BsM = Bulbospongiosus Muscle
- CC = Corpus Cavernosum
- CS = Corpus Spongiosum
- DEC = Dorsal Erection Canals
- RPM = Retractor Penis Muscle
- TA = Tunica Albuginea
- T = Trabeculae (from tunica albuginea)
- PU = Penile Urethra



**RAM**



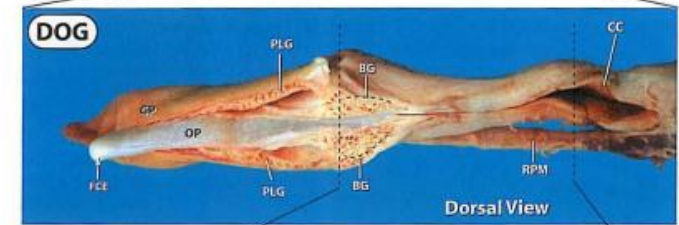
**BULL**



**BULL**

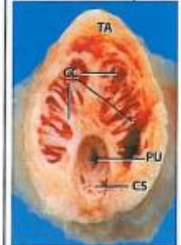
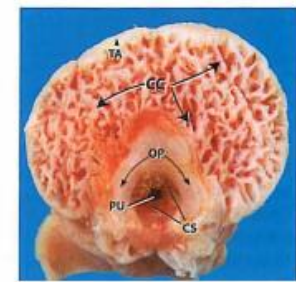


Lateral View

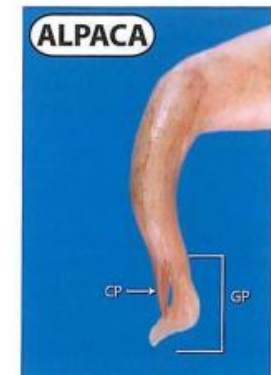


**DOG**

Dorsal View



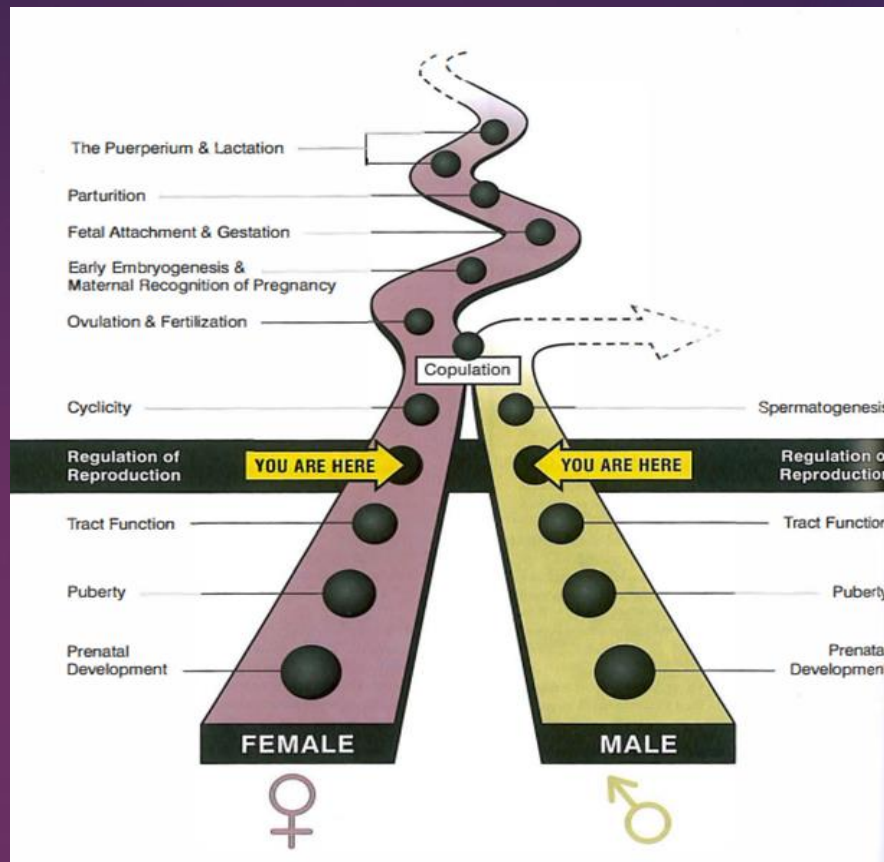
- BG = Bulbus Glandis
- CC = Corpus Cavernosum
- CP = Cartilaginous Process
- CS = Corpus Spongiosum
- FCE = Fibrocartilaginous end of Os Penis
- GP = Glans Penis
- OP = Os Penis
- PLG = Pars longa glandis
- PS = Penile Spines
- PU = Penile Urethra
- RPM = Retractor Penis Muscle
- TA = Tunica Albuginea



**ALPACA**



**TOM**



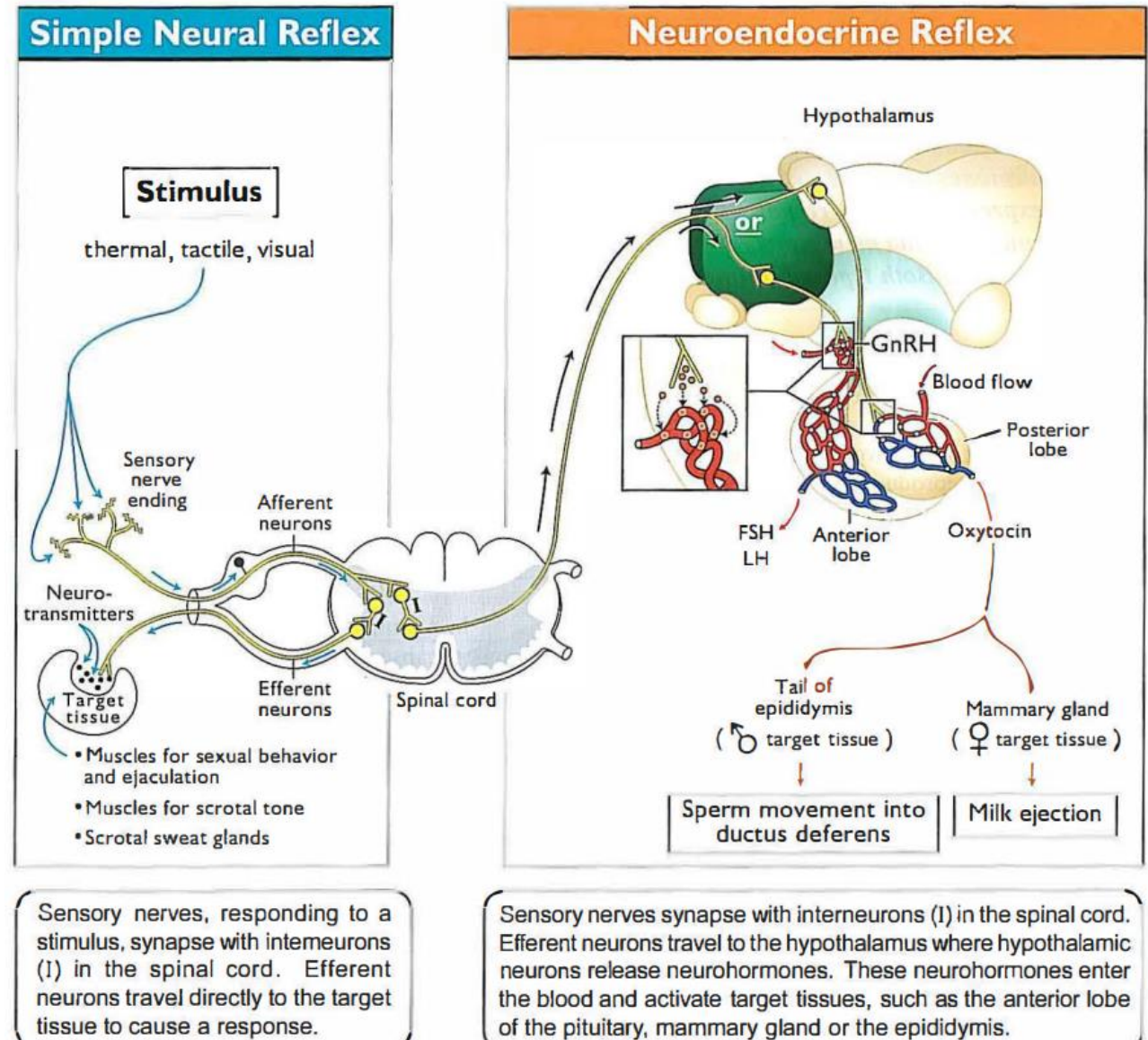
## Section IV

# *The Regulation of Reproduction Nerves, Hormones and Target Tissues*

# Regulation of the reproductive system

## IV

- Reproduction is regulated by a remarkable interplay between **the nervous system** and the **endocrine system**.





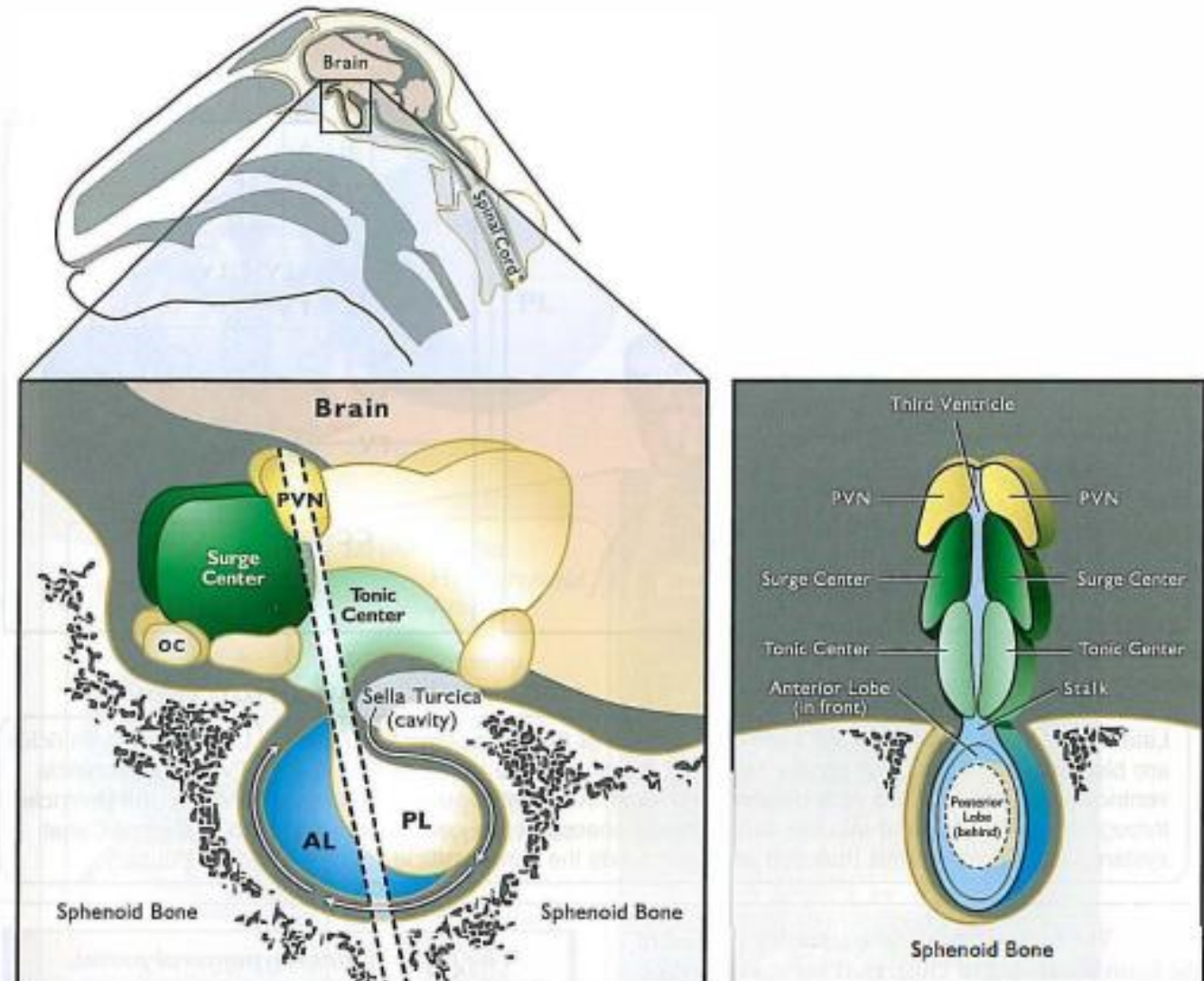
# Hypothalamus and Pituitary

## Saggital view

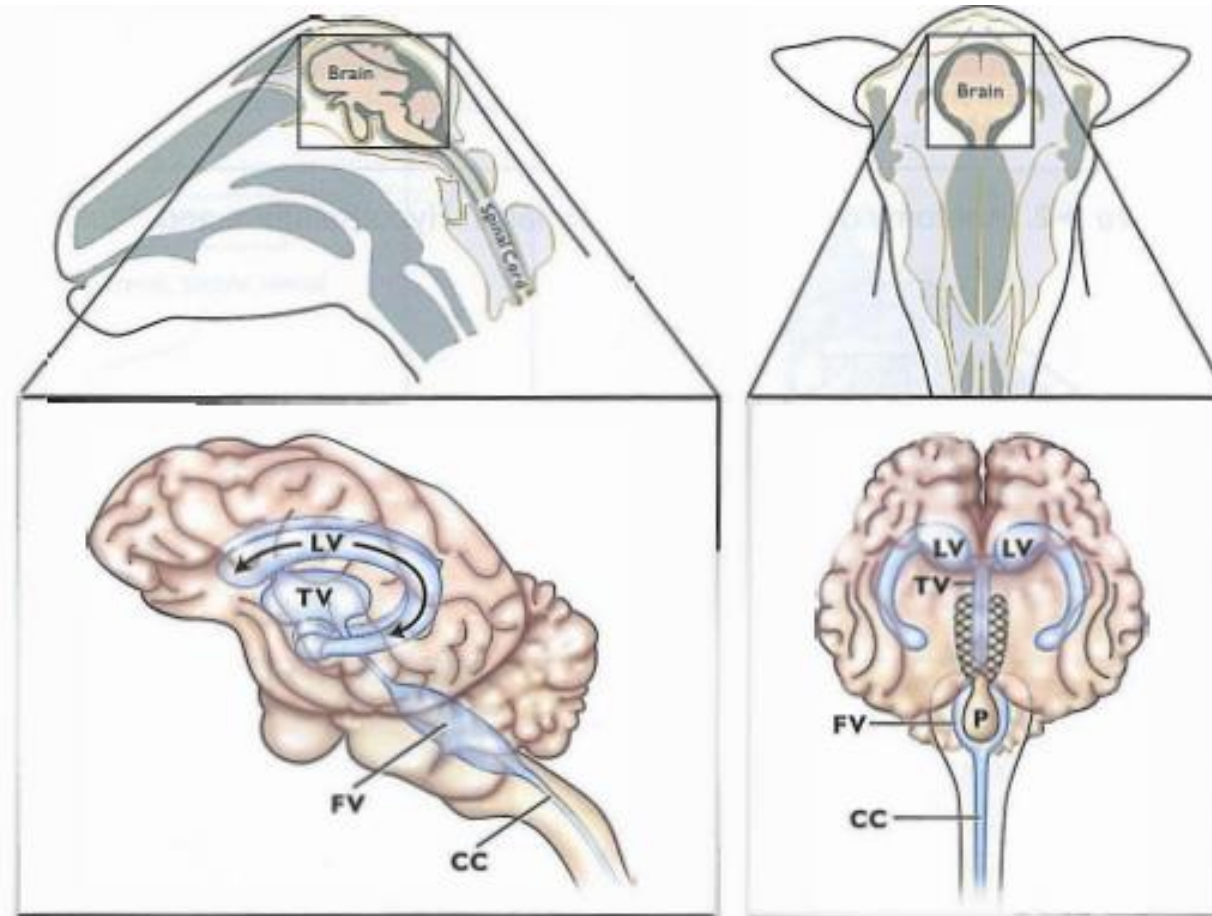
The hypothalamus is a specialized ventral portion of the brain consisting of groups of nerve cell bodies called hypothalamic nuclei that appear as lobules in the figure. The surge center, the tonic center and the paraventricular nucleus (PVN) have direct influence on reproduction. The anterior and posterior lobes of the pituitary are positioned in a depression of the sphenoid bone called the sella turcica.

## Frontal view

This view illustrates the relationship of the paraventricular nucleus (PVN), the surge center and the tonic center to the third ventricle and pituitary. The vertical line in the left panel represents the plane of section shown in the right panel. Notice that the third ventricle (a brain cavity) separates the lateral portions of the hypothalamus. AL = Anterior Lobe of the Pituitary, PL = Posterior Lobe of the Pituitary, OC = Optic Chiasm.



# Ventricular System of the Brain



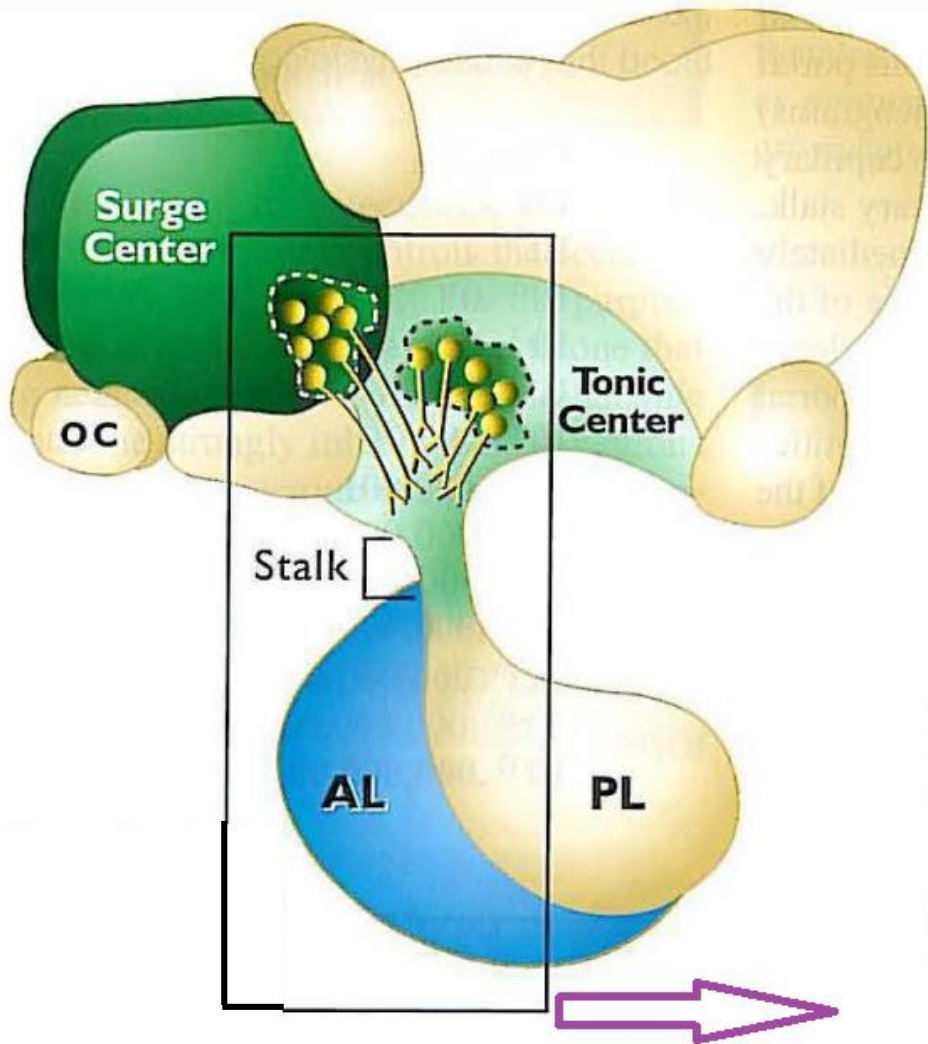
**Lateral view**

**Anterior view**

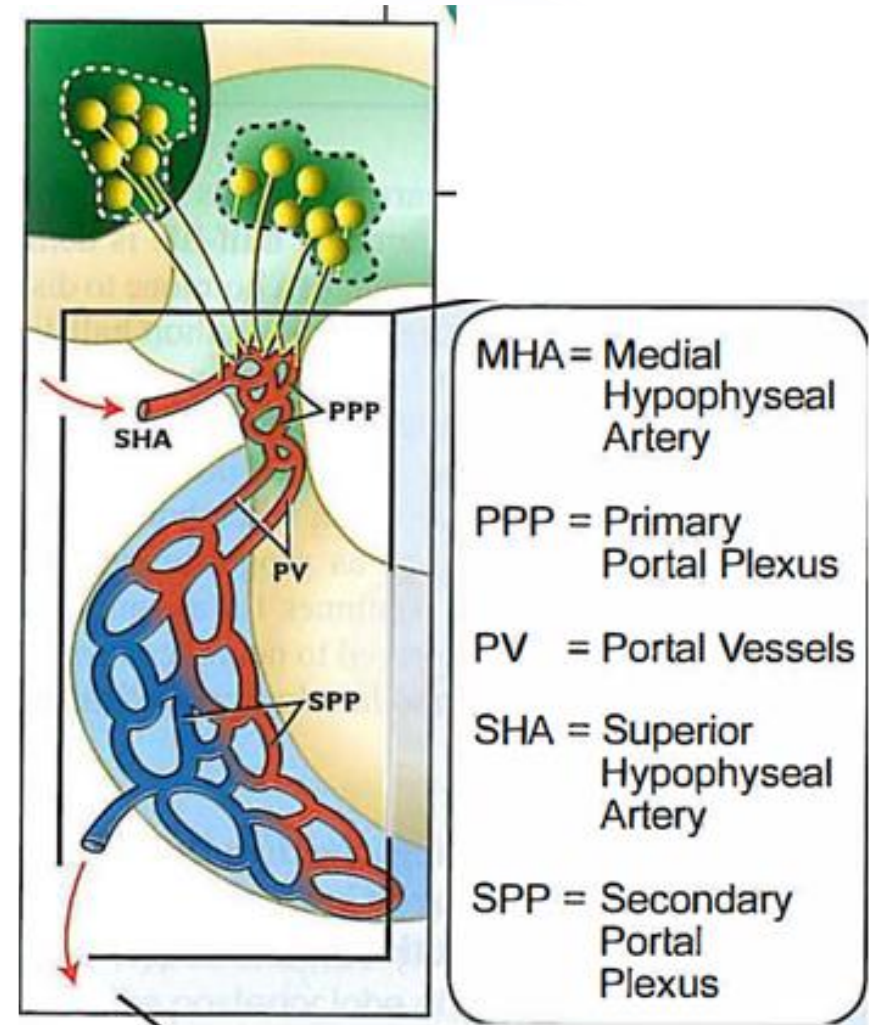
Lateral and anterior views of the ventricular system of the brain. The ventricles are blue-shaded "bags" and appear here as if the brain were transparent. The ventricular system is filled with cerebrospinal fluid that continuously circulates through the ventricles and into the subarachnoid spaces of the central nervous system. The hypothalamus (hatched area) surrounds the third ventricle.

LV = Lateral Ventricles  
TV = Third Ventricle  
FV = Fourth Ventricle  
CC = Central Canal  
P = Pituitary

# The Hypothalamo-Hypophyseal Portal System IV



Axons from neurons in the surge center and the tonic center extend to the stalk region where their endings terminate upon blood vessels of the hypothalamo-hypophyseal portal system. This portal system consists of: the superior hypophyseal artery; the primary portal plexus, (where the surge center and tonic center neurons terminate); the medial hypophyseal artery that supplies part of the anterior lobe of the pituitary (AL); the portal vessels that transport blood containing releasing hormones; and the secondary portal plexus that delivers blood (and releasing hormones) to the cells of the anterior lobe.



# *PVN and the PL of the Pituitary*

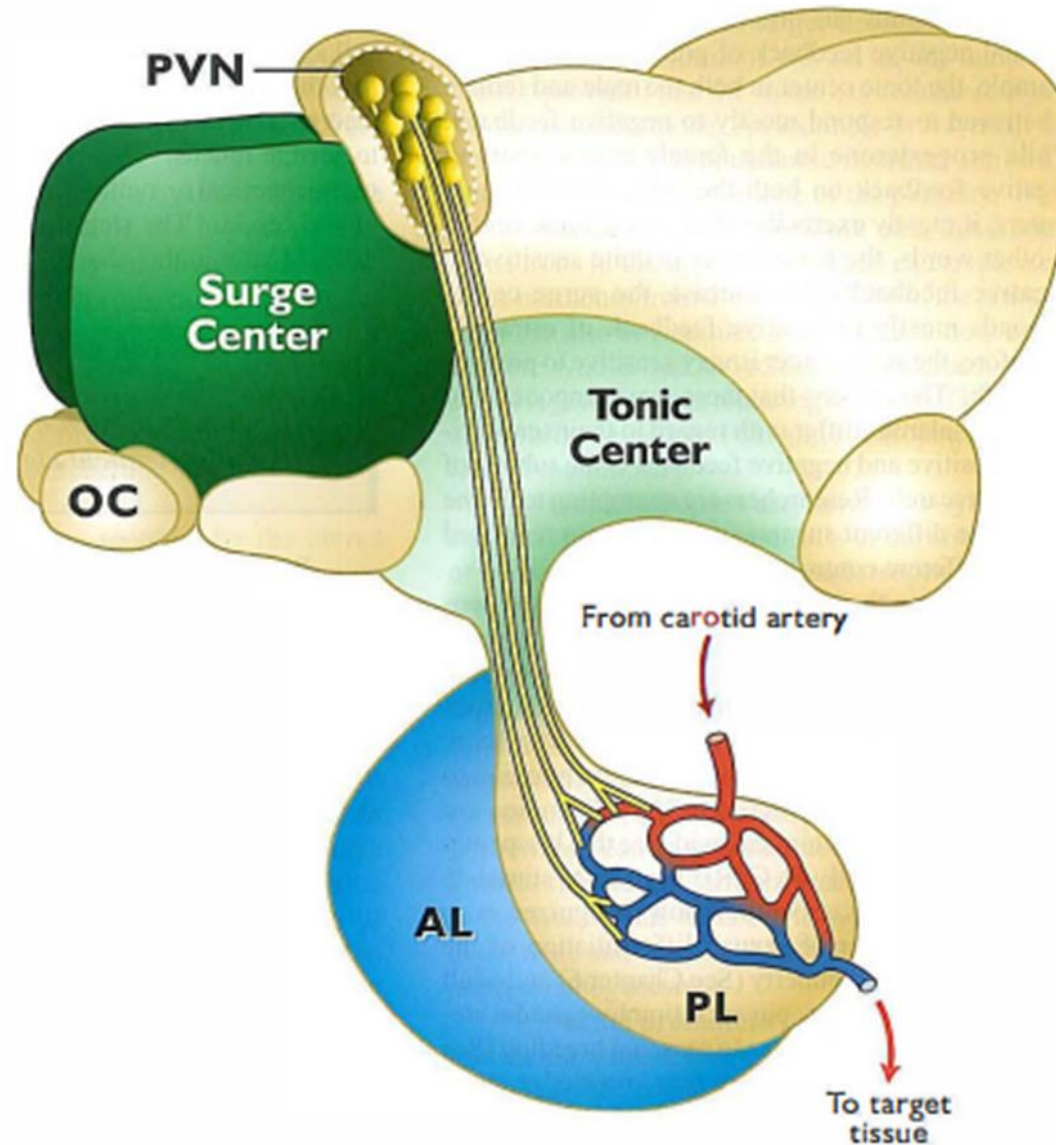
Axons from neurons originating in the hypothalamus (PVN) extend into the posterior lobe of the pituitary where they release their neurohormones into a capillary plexus.

AL = Anterior Lobe of the Pituitary

OC = Optic Chiasm

PL = Posterior Lobe of the Pituitary

PVN = Paraventricular Nucleus



# *Endocrine control vs. Neural control*

---

- In contrast to neural regulation, the endocrine system relies on **hormones** to cause responses. A hormone is a substance produced by a gland that acts on a remote tissue (target tissue) to bring about a change in the target tissue. These changes involve alterations in **metabolism**, **synthetic activity** and **secretory activity**.
- Hormones are characterized as having relatively **short half-lives** which is important because once the hormone is secreted and released into the blood and causes a response, it is degraded so that further responses do not occur.
- Compared to neural control, hormonal control is **slower** and has durations of minutes, hours or even days.

# *Positive and Negative Feedbacks*

## IV

- Almost all reproductive functions are controlled by **positive** and **negative feedback** mechanisms.
- These mechanisms control the secretion of **GnRH** that in-turn controls the secretion of the gonadotropins **FSH** and **LH**.
  - **Progesterone** strongly inhibits GnRH neurons and therefore when progesterone is high, GnRH neurons secrete only basal levels of GnRH. Such basal secretion while allowing for some follicular development will not allow sufficient follicular development for the secretion of high levels of estradiol.

$\uparrow P_4 \rightarrow \downarrow \text{GnRH} \rightarrow \downarrow \text{FSH \& LH} =$   
*Incomplete follicular development*

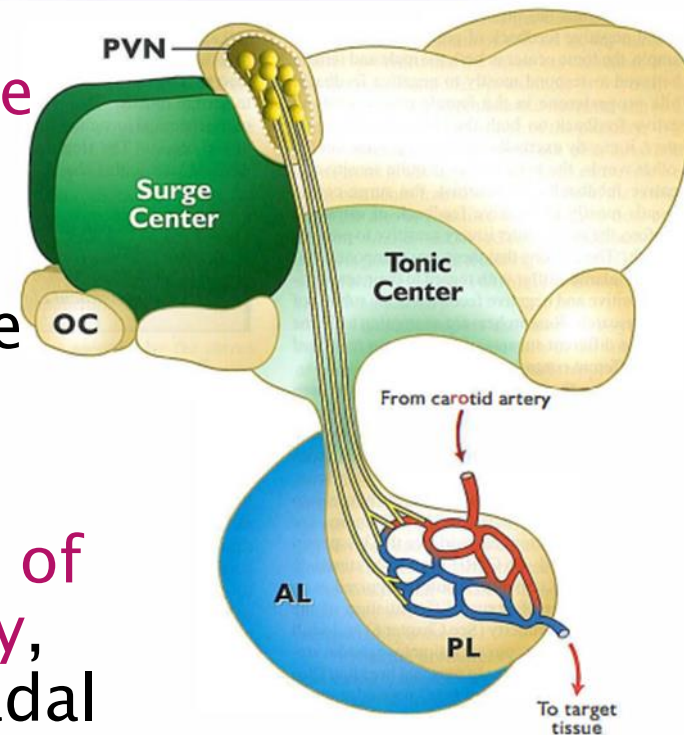
$\uparrow E_2 \rightarrow \uparrow \text{GnRH (surge)} \rightarrow \text{LH surge} =$   
*Ovulation*

- The female contains a **surge center** that is responsible for secreting large quantities of GnRH that induce ovulation. The surge center will not release large quantities of GnRH until there is positive feedback by **estradiol**.

# Positive and Negative Feedbacks

## IV

- The **tonic center** in both the male and female is believed to respond mostly to **negative feedback**. While progesterone in the female exerts a strong negative feedback on both the surge and the tonic centers, it mostly exerts its effect on the tonic center.
- In contrast, the **surge center** responds mostly to **positive feedback** of estradiol.
- A new class of neuropeptides called **kisspeptins** has emerged as the possible "gatekeepers" for GnRH release which are secreted by hypothalamic neurons in the **periventricular, preoptic and arcuate** nuclei.
- Kisspeptin is now recognized as an important regulator of **sexual differentiation of the brain**, the timing of **puberty**, and adult regulation of **gonadotropin secretion** by gonadal steroids, especially as it relates to **seasonal breeding**.



# *Reproductive Hormones*

---

Reproductive hormones can be classified according to their:

- **Source of origin**
  - Hypothalamic hormones (GnRH),
  - Pituitary hormones (FSH, LH, PL, OT),
  - Gonadal hormones (estrogens, progesterone, inhibin, testosterone, oxytocin, relaxin),
  - Uterine hormones (PGF<sub>2α</sub>),
  - Placental hormones (hCG, eCG),
  - Mammary gland bioactive factor (lactocrine signaling)
    - Lactocrine transmission of relaxin and its effects on development of the neonatal female reproductive tract



# Reproductive Hormones

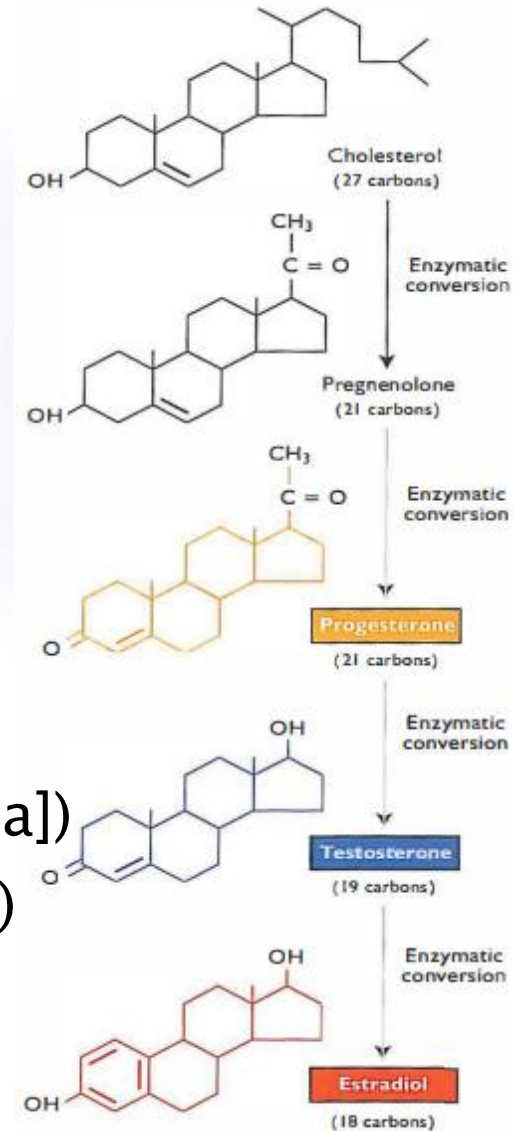
## IV

- **Mode of action**

- Neurohormones (OT)
- Releasing hormones (GnRH)
- Gonadotropins (FSH, LH)
- Sexual promoters (estrogens, progesterone, testosterone, hCG, eCG, placental lactogen)
- General metabolic hormones (thyroxin, adrenal corticoids, GH)
- Luteolytic hormones ( $\text{PGF}_{2\alpha}$ )

- **Biochemical structure**

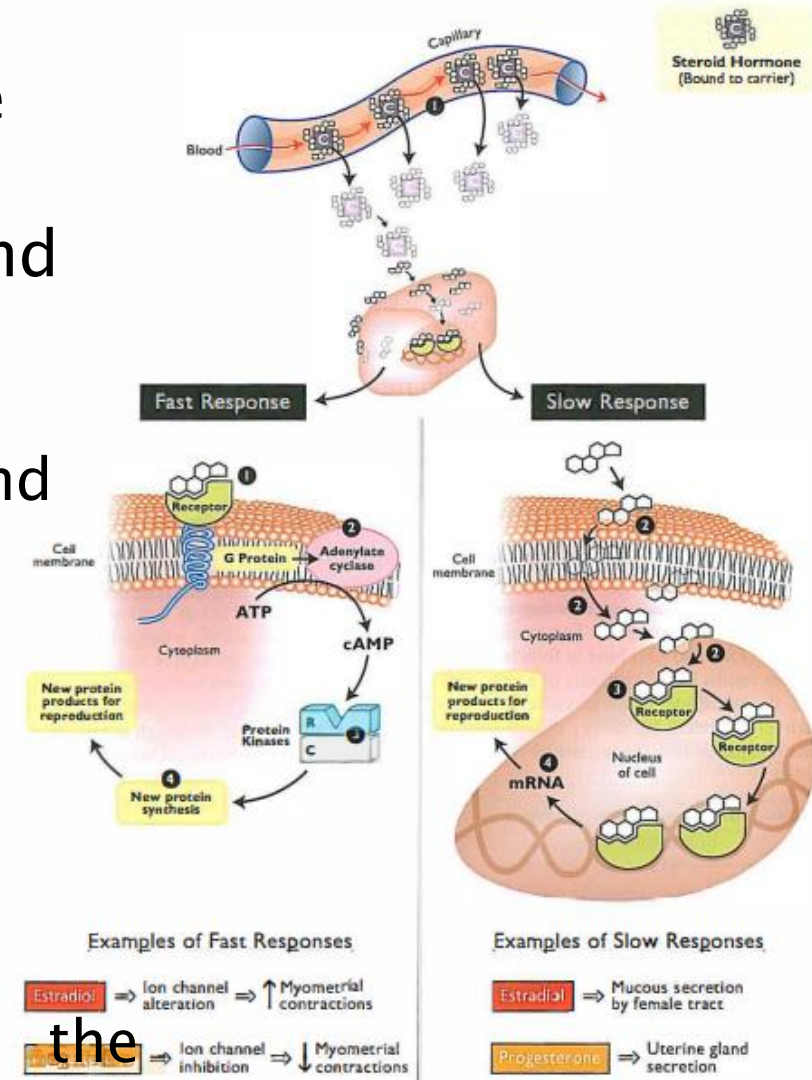
- Peptides (GnRH, PL, Relaxin [from CL of pregnancy and placenta])
- Glycoproteins with  $\alpha$  and  $\beta$  subunits (inhibin, activin, follistatin)
- Steroids (progesterone, testosterone, estradiol)
- Prostaglandins ( $\text{PGF}_{2\alpha}$ ,  $\text{PGE}_2$ )



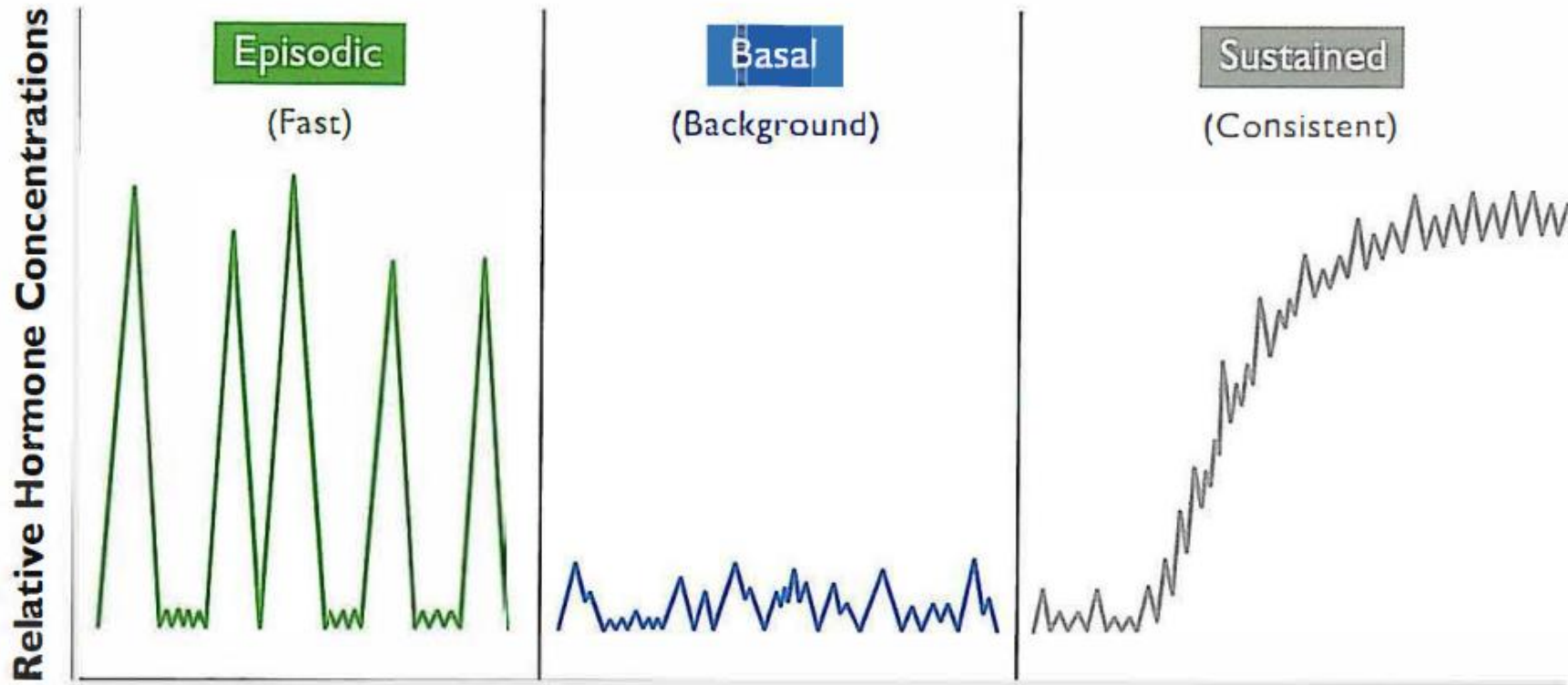
# Mechanisms of Steroid Hormone Action

## IV

- **Step 1 - Steroid Transport**
  - bind to a variety of plasma proteins which carry the steroids in the blood and interstitial fluid
- **Step 2 - Movement Through the Cell Membrane and Cytoplasm**
  - diffuses through the plasma membrane and when enters the cell, it diffuses through the cytoplasm and into the nucleus
- **Step 3 - Binding to Nuclear Receptor**
  - The steroid-receptor complex (transcription factor) initiates DNA-directed messenger RNA synthesis (transcription)
- **Step 4 - mRNA and Protein Synthesis**
  - specific proteins are synthesized that will enhance the reproductive process



# Hormonal Secretion Patterns



Episodic secretion is generally associated with hormones under nervous control. When nerves of the hypothalamus fire, neuropeptides are released in a sudden burst or pulse.

In a basal secretion pattern, the hormone stays low but fluctuates with low amplitude pulses.

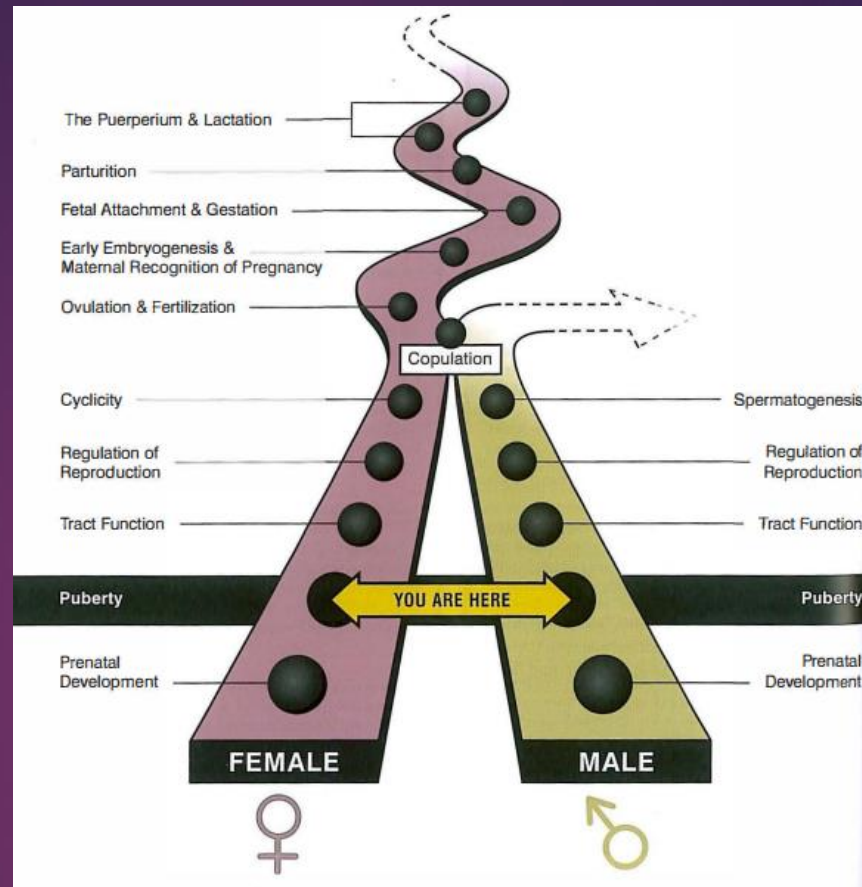
In the sustained hormone release profile, the hormone remains elevated, but in a relatively steady fashion for a long period (days to weeks). Steroids tend to be secreted in this manner.

| Name of Hormone (Abbrev.)                              | Biochemical Classification      | Source   | Male Target Tissue   |
|--|---------------------------------|--|--|
| <b>Gonadotropin Releasing Hormone (GnRH)</b>           | Neuropeptide (decapeptide)      | Hypothalamic surge and tonic centers   | Anterior lobe-pituitary (gonadotroph cells)  |
| <b>Luteinizing Hormone (LH)</b>                        | Glycoprotein                    | Anterior lobe (pituitary) (gonadotroph cells)  | Testis (interstitial cells of Leydig)  |
| <b>Follicle Stimulating Hormone (FSH)</b>              | Glycoprotein                    | Anterior lobe-pituitary (gonadotroph cells)  | Testis (Sertoli cells)   |
| <b>Prolactin (PRL)</b>                                 | Protein                         | Anterior lobe-pituitary (lactotroph cells)   | Testis and brain   |
| <b>Oxytocin (OT)</b>                                   | Neuropeptide (octapeptide)      | Synthesized in the hypothalamus, stored in the posterior lobe-pituitary; synthesized by corpus luteum. | Smooth muscle of epididymal tail, ductus deferens and ampulla                            |
| <b>Estradiol (E<sub>2</sub>)</b>                       | Steroid                         | Granulosa cells of follicle, placenta, Sertoli cells of testis   | Brain<br>Inhibits long bone growth   |
| <b>Progesterone (P<sub>4</sub>)</b>                    | Steroid                         | Corpus luteum and placenta   |  |
| <b>Testosterone (T)</b>                                | Steroid                         | Interstitial cells of Leydig, cells of theca interna in female   | Accessory sex glands, tunica dartos of scrotum, seminiferous epithelium, skeletal muscle |
| <b>Inhibin</b>   | Glycoprotein                    | Granulosa cells (female) Sertoli cells (male)  | Gonadotrophs of anterior lobe-pituitary  |
| <b>Prostaglandin F<sub>2α</sub> (PGF<sub>2α</sub>)</b> | Prostaglandin (C-20 fatty acid) | Uterine endometrium, vesicular glands  | Epididymis   |
| <b>Relaxin (RLN or RLX)</b>                            | Protein Polypeptide             | Corpus luteum, placenta prostate   | Sperm and male tract   |
| <b>Human chorionic gonadotropin (hCG)</b>              | Glycoprotein                    | Trophoblast of blastocyst (chorion)  |  |
| <b>Equine chorionic gonadotropin (eCG)</b>             | Glycoprotein                    | Chorionic girdle cells   |  |
| <b>Placental lactogen</b>                              | Protein                         | Placenta   |  |

| Female Target Tissue   | Male Primary Action   | Female Primary Action   |
|--|---|---|
| Anterior lobe-pituitary (gonadotroph cells)                                | Release of FSH and LH from anterior lobe-pituitary                                    | Release of FSH and LH from anterior lobe-pituitary  |
| Ovary (cells of theca interna and luteal cells)                            | Stimulates testosterone production  | Stimulates ovulation, formation of corpora lutea and progesterone secretion                                     |
| Ovary (granulosa cells)  | Sertoli cell function   | Follicular development and estradiol synthesis  |
| Mammary cells, corpus luteum in some species (rat and mouse)               | Can induce maternal behavior in females and males                                     | Lactation, maternal behavior and corpora lutea function (some species)  |
| Myometrium and endometrium of uterus, myoepithelial cells of mammary gland | PGF <sub>2α</sub> synthesis and pre-ejaculatory movement of spermatozoa               | Uterine motility, promotes uterine PGF <sub>2α</sub> synthesis, milk ejection                                   |
| Hypothalamus, entire reproductive tract and mammary gland                  | Sexual behavior   | Sexual behavior, GnRH, elevated secretory activity of the entire tract, enhanced uterine motility               |
| Uterine endometrium, mammary gland, myometrium, hypothalamus               |   | Endometrial secretion, inhibits GnRH release, inhibits reproductive behavior, promotes maintenance of pregnancy |
| Brain, skeletal muscle, granulosa cells                                    | Anabolic growth, promotes spermatogenesis, promotes secretion of accessory sex glands | Substrate for E <sub>2</sub> synthesis, abnormal masculinization (hair patterns, voice, behavior, etc.)         |
| Gonadotrophs of anterior lobe-pituitary                                    | Inhibits FSH secretion  | Inhibits FSH secretion  |
| Corpus luteum, uterine myometrium, ovulatory follicles                     | Affects metabolic activity of spermatozoa, causes epididymal contractions             | Luteolysis, promotes uterine tone and contraction, ovulation  |
| Pelvic ligaments, cervix, mammary gland, nipples                           |   | Softening of pelvic ligaments, cervix, connective tissue remodeling in tract                                    |
| Ovary  | Sperm motility, tract growth  | Facilitate production of progesterone by ovary  |
| Ovary  |   | Causes formation of accessory corpora lutea   |
| Mammary gland of dam   |   | Mammary stimulation of dam  |

# Section V

## *Puberty*



# Puberty

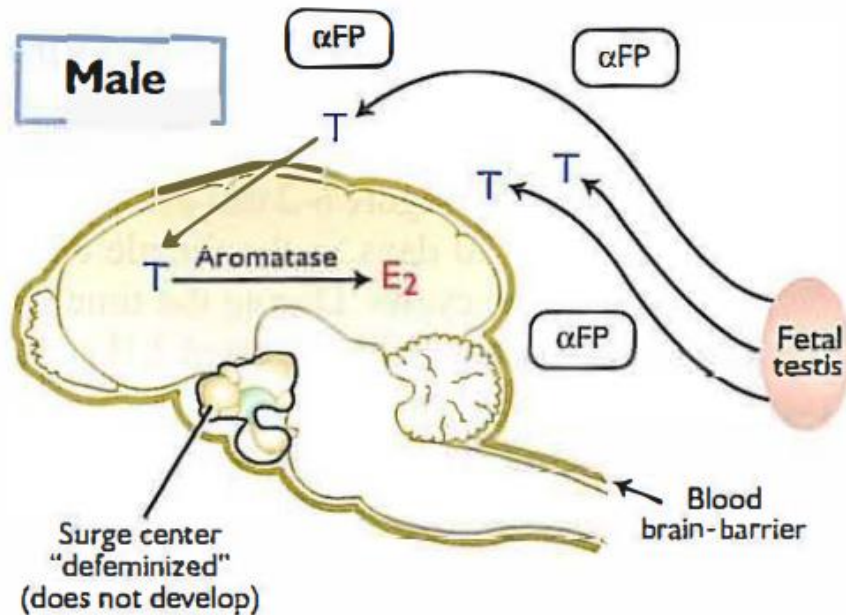
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- **Puberty** is the acquisition of reproductive competence.
- It is a **process** that occurs over time, not an **event**.
- The **onset** of puberty depends on the ability of specific **hypothalamic neurons** to produce **GnRH** in sufficient quantities to promote and support gametogenesis.
- In the **female**, hypothalamic GnRH neurons must develop the ability to **respond to estradiol positive feedback** before they can cause sufficient quantities of GnRH to induce ovulation.
- Development of hypothalamic GnRH neurons is influenced by **genetic** and **environmental** factors and their **interactions**.

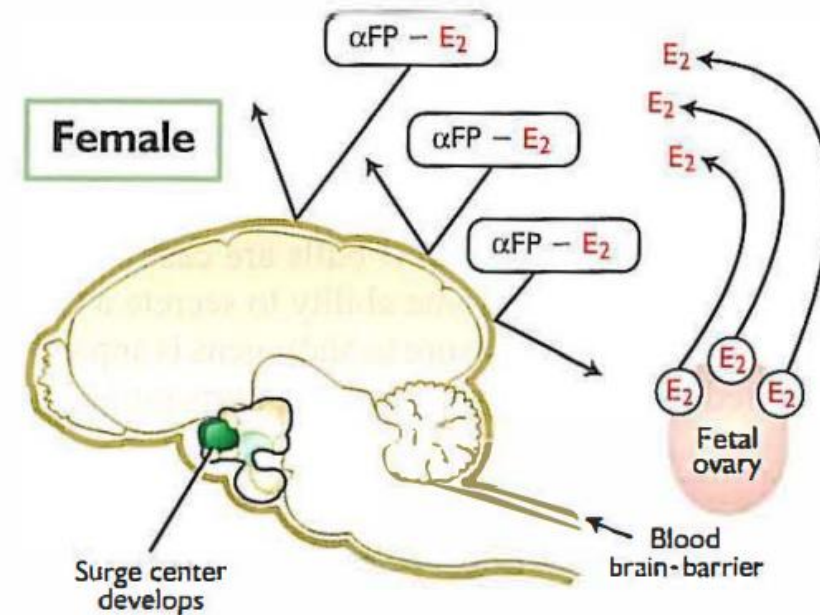
# Male hypothalamus vs. Female hypothalamus

V

- During prenatal development in the male, **testosterone** from the fetal testis "**defeminizes**" the brain thus **minimizing surge center** function.



In the male, Testosterone freely enters the brain because  $\alpha$ -FP does not bind it. Testosterone is aromatized into estradiol and the male brain is "defeminized". Therefore, a GnRH surge center **does not** develop.



In the female,  $\alpha$ -FP prevents E<sub>2</sub> from entering the brain. The hypothalamus is thus "feminized" and the surge center develops.

# *LH Secretory Patterns in Males vs. Females*

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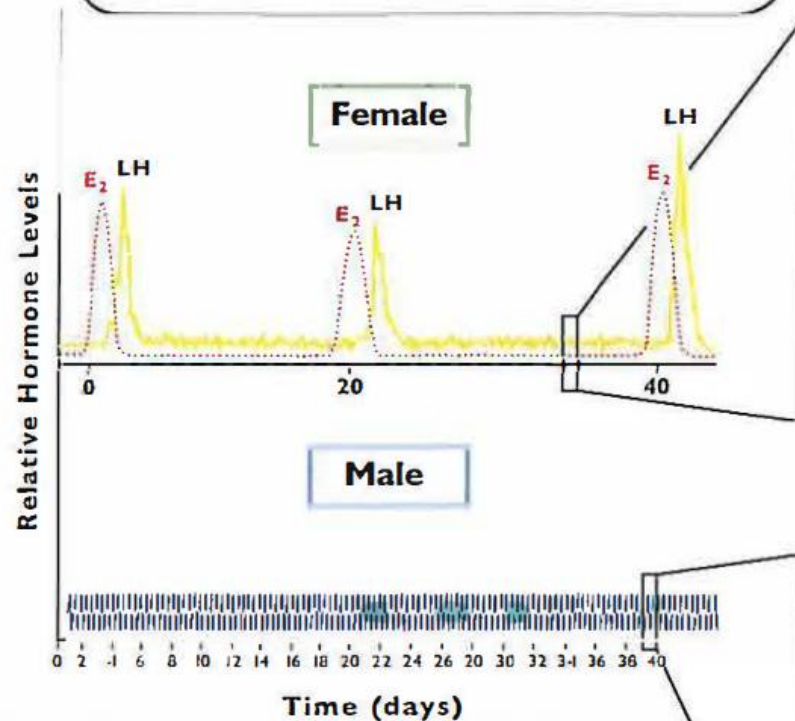
V

- LH does not surge in the **male**, but maintains a **pulsatile** pattern of secretion.
  - These pulses occur **every 2 to 6 hours** in the postpubertal male. This steady GnRH pulsatile rhythm results in steady pulses of LH and, in turn, steady pulsatile secretion of **testosterone**.
- In contrast, estradiol and **LH surge** about every 20 days in the **female** depending on the length of the cycle.
  - During the time between the surges, low amplitude, repeated LH pulses are present.

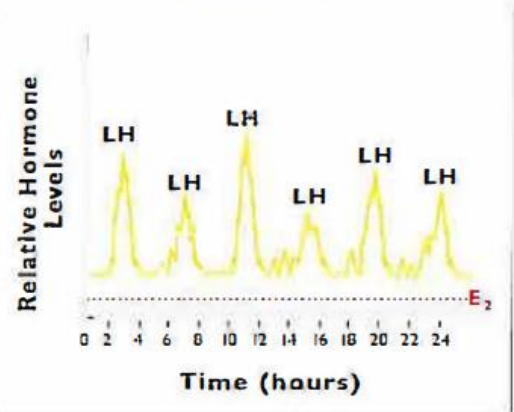


# LH Secretory Patterns in Males vs. Females

Females have high amplitude preovulatory episodes of LH once every several weeks and basal pulsatile episodes between the large preovulatory surges.



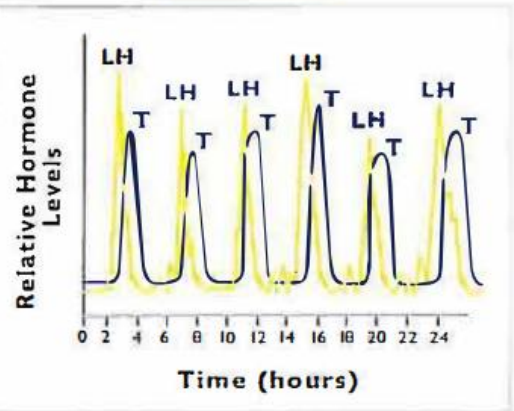
Female



Male

Males have small LH episodes that occur every 2 to 6 hours. Testosterone is secreted soon after each LH episode.

Male



# *The Onset of Puberty*

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- Generally, puberty can be defined in both the male and female as the **ability** to accomplish **reproduction** successfully.
- The fundamental requirement for puberty is the **secretion of GnRH** at the appropriate frequency and quantities to stimulate **gonadotropin** release by the pituitary
- Gonadotropins promote **gametogenesis**, **steroidogenesis** and the **development of reproductive tissues**.
- The most important "drivers" of pubertal onset are the ability of **presynaptic neurons** to provide information to the **GnRH neurons**.
  - Function of these presynaptic neurons may be influenced by: **1)** plane of nutrition, **2)** exposure to certain environmental or social cues and **3)** the genetics of the individual.

# The Onset of Puberty - Criteria

- Several criteria can be used to define puberty in domestic animals.
- **In female:**
  - Age at **first estrus** (heat): The age at which the female becomes sexually receptive.
  - Age at **first ovulation**: This can be accomplished using palpation or ultrasonography of the ovary per rectum in animals. Also, laparoscopy and endoscopy can be used to determine when ovulation has occurred.
  - Age at which a female can **support pregnancy** without deleterious effects.

- **In male:**
  - Age when **behavioral traits** are expressed
  - Age at **first ejaculation**
  - Age when **spermatozoa first appear** in the ejaculate
  - Age when the ejaculate contains a **threshold number of spermatozoa**

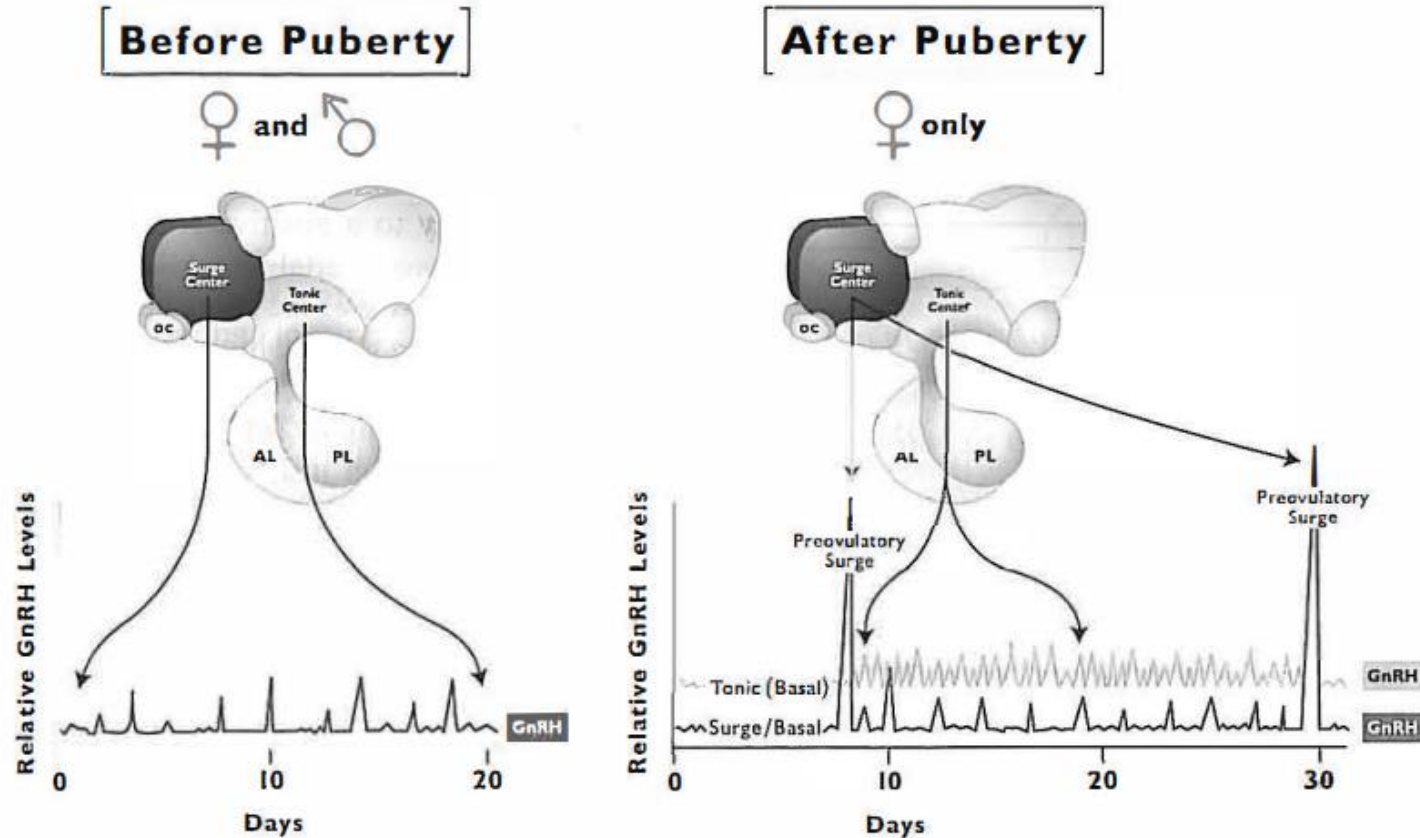
| <u>Species</u>      | <u>Male</u>   | <u>Female</u> |
|---------------------|---------------|---------------|
| Alpaca <sup>2</sup> | 2-3 yrs       | 1 yr          |
| Bovine              | 11 mo (7-18)  | 11 mo (9-24)  |
| Camel <sup>2</sup>  | 3-5 yrs       | 3 yrs         |
| Canine <sup>1</sup> | 9 mo (5-12)   | 12 mo (6-24)  |
| Equine              | 14 mo (10-24) | 18 mo (12-19) |
| Feline              | 9 mo (8-10)   | 8 mo (4-12)   |
| Llama <sup>2</sup>  | 2-3 yrs       | 6-12 mo       |
| Ovine               | 7 mo (6-9)    | 7 mo (4-14)   |
| Porcine             | 7 mo (5-8)    | 6 mo (5-7)    |

# *The Onset of Puberty - Mechanisms*

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- The major factor limiting onset of puberty is the failure of the hypothalamus to secrete sufficient quantities of GnRH to cause gonadotropin release.
- In the male, the onset of puberty is brought about because of decreased hypothalamic sensitivity to negative feedback by testosterone/estradiol.
- In the prepubertal female, the surge center is quite sensitive to the positive feedback of estradiol. But, the surge center cannot release "ovulatory quantities" of GnRH because the ovary cannot secrete high levels of estradiol.
- At low concentrations of estradiol, the tonic center has a high sensitivity to negative feedback and therefore does not secrete high levels of GnRH and gonadotropins remain low.
- During the pubertal transition, however, the negative feedback sensitivity by the tonic center to estradiol decreases and consequently higher and higher amounts of GnRH are secreted causing an increase in pulse frequency of LH.

# The Onset of Puberty - Mechanisms



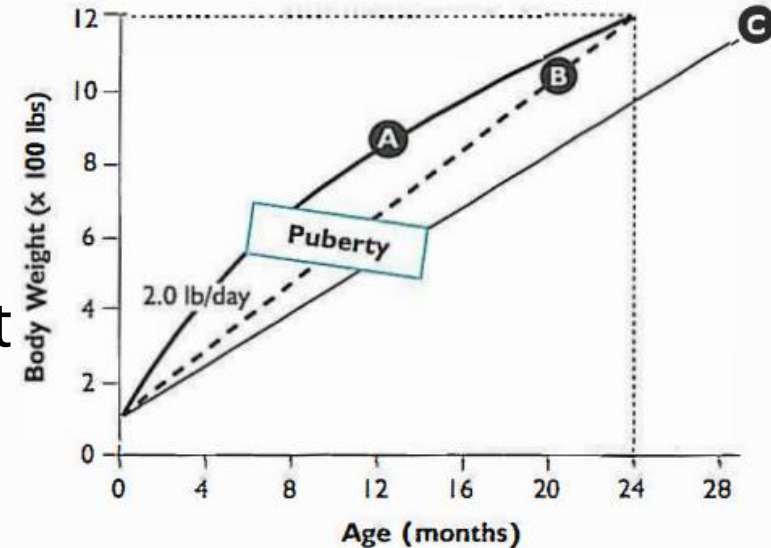
Before puberty in both the female and male, GnRH neurons in the tonic center and the surge center of the hypothalamus release low amplitude and low frequency pulses of GnRH.

After puberty in the female, the tonic center controls basal levels of GnRH, but they are higher than in the prepubertal female because the pulse frequency increases. The surge center controls the preovulatory surge of GnRH. The male does not develop a surge center.

# The Onset of Puberty - Mechanisms

V

- There is evidence to indicate that initiation of **high frequency GnRH pulses** is under the influence of **glucose** and **free fatty acid** concentrations in the blood.
- The impact of nutrition on the age of pubertal onset in dairy heifers has been shown in different studies.
- **Leptin** is a hormonal peptide, discovered in 1994, that is secreted by **adipocytes** (fat cells) and its **receptors** have also been discovered in the anterior lobe of the pituitary and **hypothalamus**
- Leptin may be an important **signal** that "notifies" key hypothalamic neurons that influence GnRH secretion that **nutritional status** is **adequate** because a threshold degree of "fatness" has been achieved

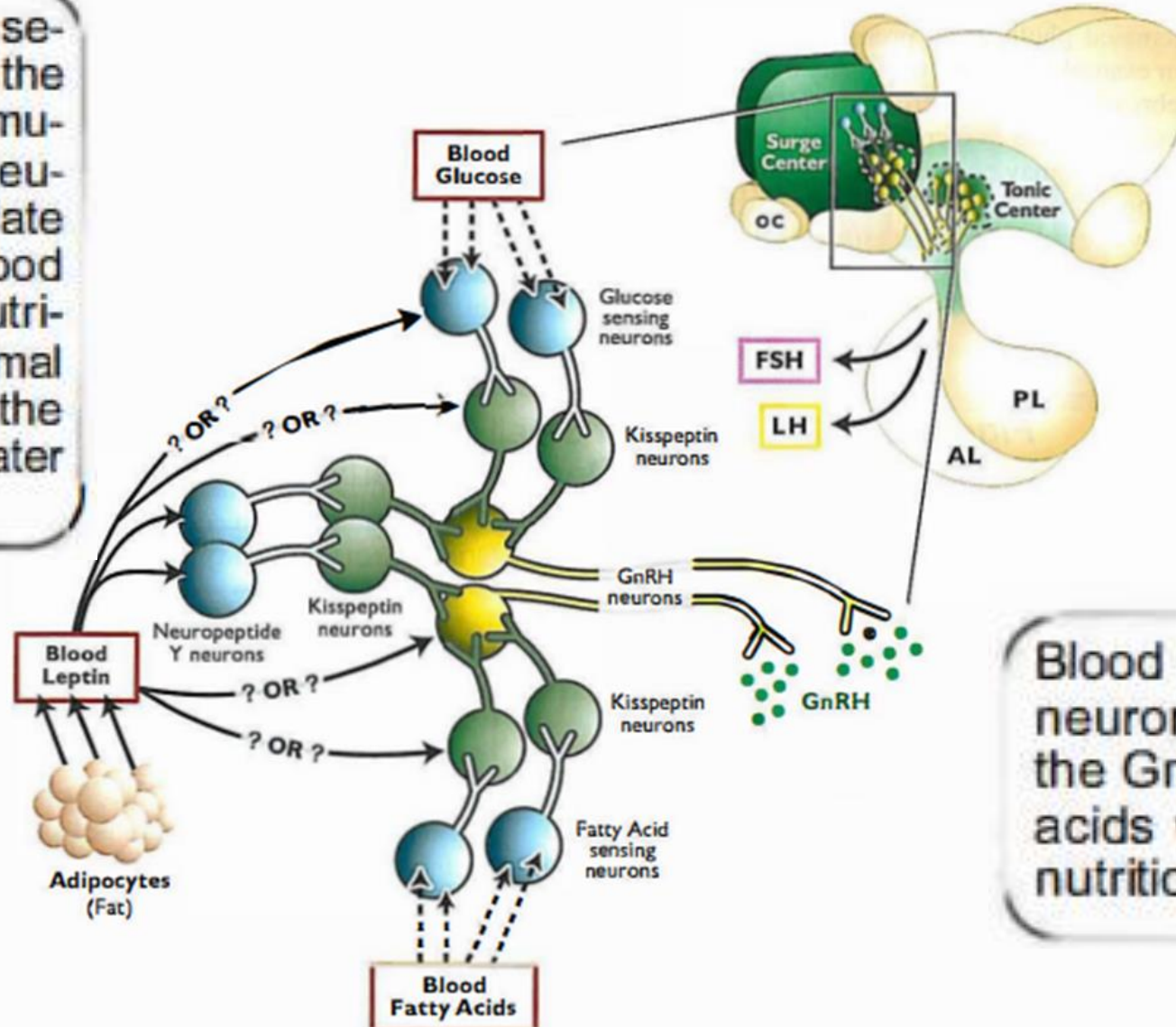


- A = High plane of nutrition (2.0 lb/day average daily gain)
- B = Moderate plane of nutrition (1.5 lb/day average daily gain)
- C = Low plane of nutrition (1.2 lb/day average daily gain)

Age at first parturition should be 24 months and the primiparous heifer should weigh 1,200 lb.

# Possible Influence of Metabolic Signals on Puberty V

Adipocytes (fat cells) secrete leptin that enters the blood. Leptin may stimulate neuropeptide Y neurons or directly stimulate GnRH neurons. Blood leptin reflects the nutritional status of the animal because the greater the amount of fat the greater the amount of leptin.



Blood glucose concentrations, another indicator of metabolic status, might stimulate glucose sensing neurons that in turn stimulate GnRH neurons.

Blood fatty acids may stimulate neurons that in turn stimulate the GnRH neurons. Blood fatty acids would be an indicator of nutritional status of the animal.

# Journal Club *(for more reading)*

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## Metabolic signals in human puberty: effects of over and undernutrition

G A Martos-Moreno <sup>1</sup>, J A Chowen, J Argente

Affiliations + expand

PMID: 20026379 DOI: 10.1016/j.mce.2009.12.017

### Abstract

Puberty in mammals is associated with important physical and psychological changes due to the increase in sex steroids and growth hormone (GH). Indeed, an increase in growth velocity and the attainment of sexual maturity for future reproductive function are the hallmark changes during this stage of life. Both growth and reproduction consume high levels of energy, requiring suitable energy stores to face these physiological functions. During the last two decades our knowledge concerning how peptides produced in the digestive tract (in charge of energy intake) and in adipose tissue (in charge of energy storage) provide information regarding metabolic status to the central nervous system (CNS) has increased dramatically. Moreover, these peptides have been shown to play an important role in modulating the gonadotropic axis with their absence or an imbalance in their secretion being able to disturb pubertal onset or progression. In this article we will review the current knowledge concerning the role played by leptin, the key adipokine in energy homeostasis, and ghrelin, the only orexigenic and growth-promoting peptide produced by the digestive tract, on sexual development. The normal evolutionary pattern of these peripherally produced metabolic signals throughout human puberty will be summarized. The effect of two opposite situations of chronic malnutrition, obesity and anorexia, on these signals and how they influence the course of puberty will also be discussed. Finally, we will briefly mention other peptides derived from the digestive tract (such as PYY) that may be involved in the regulatory link between energy homeostasis and sexual development.



# *Other Factors affecting Puberty*

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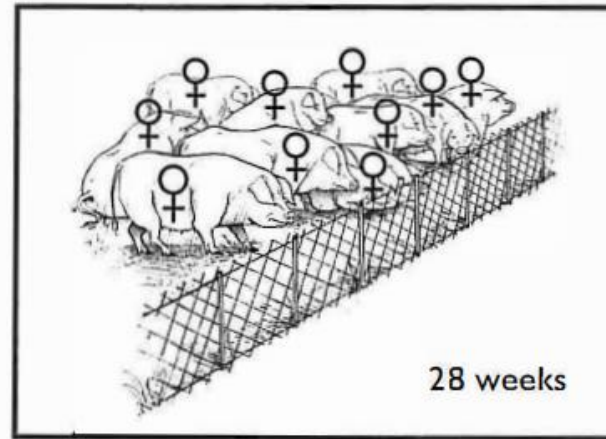
- **Season of birth** can influence the age of puberty
  - In natural photoperiods, spring-born (February-March) **lambs** receiving adequate nutrition attain puberty during the subsequent fall (September-October). The age at puberty is about 5 to 6 months after birth. In contrast, fall-born lambs do not reach puberty until about 10 to 12 months.
  - **Heifers** born in autumn tend to reach puberty earlier than those born in spring. Exposure during the second six months of their life to long photoperiods and spring/summer-like temperatures hastens the onset of puberty.
  - In the **bitch** there is little seasonality associated with the onset of puberty.
  - In the **queen** increased photoperiod prompts the onset of puberty.

# Other Factors affecting Puberty

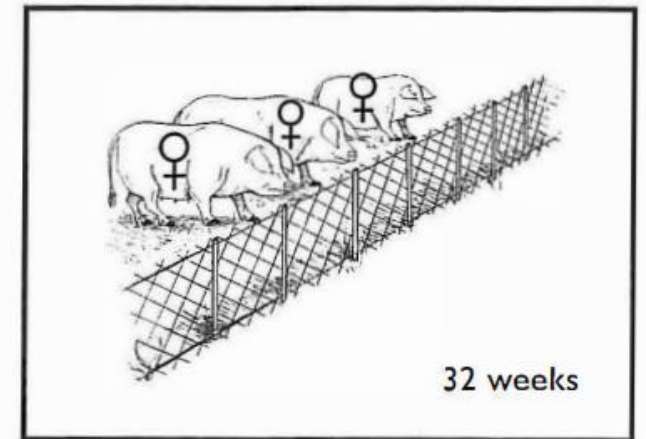
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- **Social cues** significantly impact the onset of puberty in many mammalian species.
  - Such mediation is caused by olfactory recognition of **pheromonal substances** present in the urine.
  - Enhancement of the onset of puberty by the presence of the male has been demonstrated in the ewe, sow and cow.

Large Groups (>10) = Normal Puberty



Small Groups (2-3 gilts) = Delayed Puberty



Exposure to a Boar = Accelerated Puberty

